The Best Available Science Supports Most Probable Number (MPN) Testing Methods for Type Approval of Ballast Water Management Systems

John J. Cullen

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21st International Conference on Aquatic Invasive Species
April 27-31, 2019
Montréal, QC, Canada

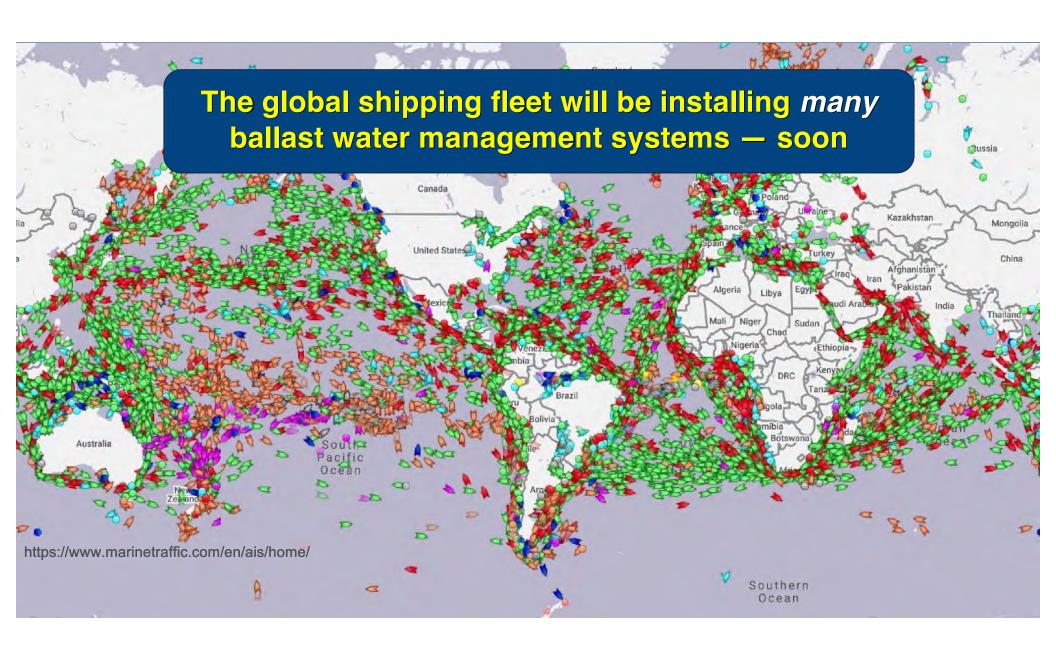




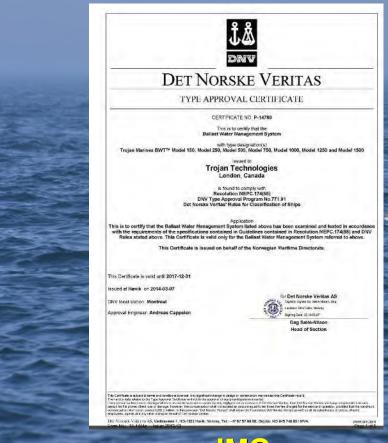
Cullen ICAIS Montréal — 201

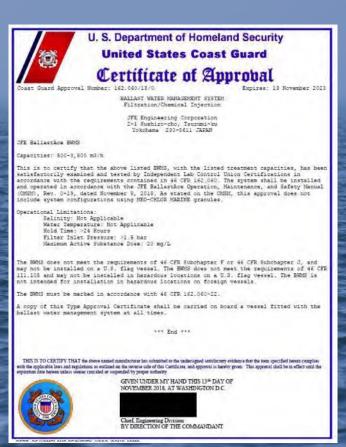
Cullen, J.J., 2019. The best available science supports most probable number (MPN) testing methods for type approval of ballast water management systems, 21st International Conference on Aquatic Invasive Species, Montréal, QC, Canada, pp. 1-37. doi: 10.5281/zenodo.3531993

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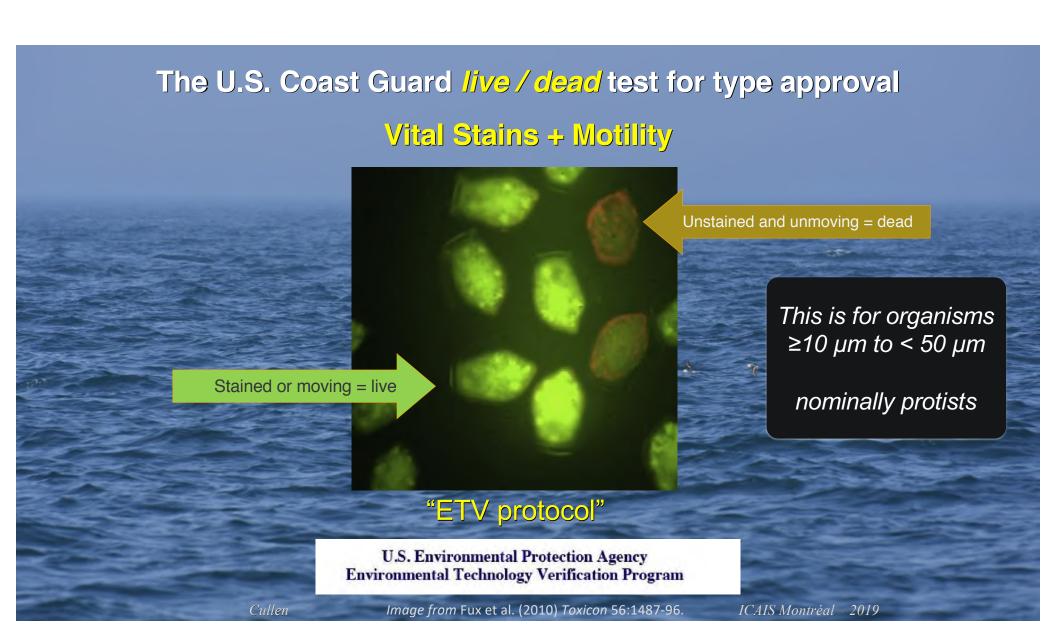
BWMS type approval is required



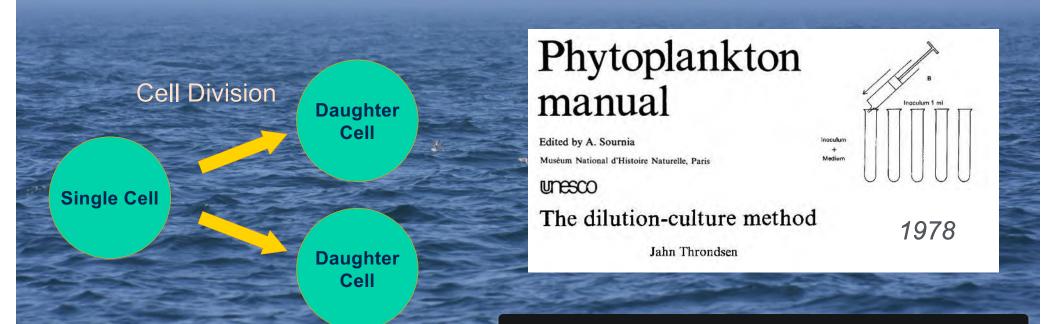


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USCG

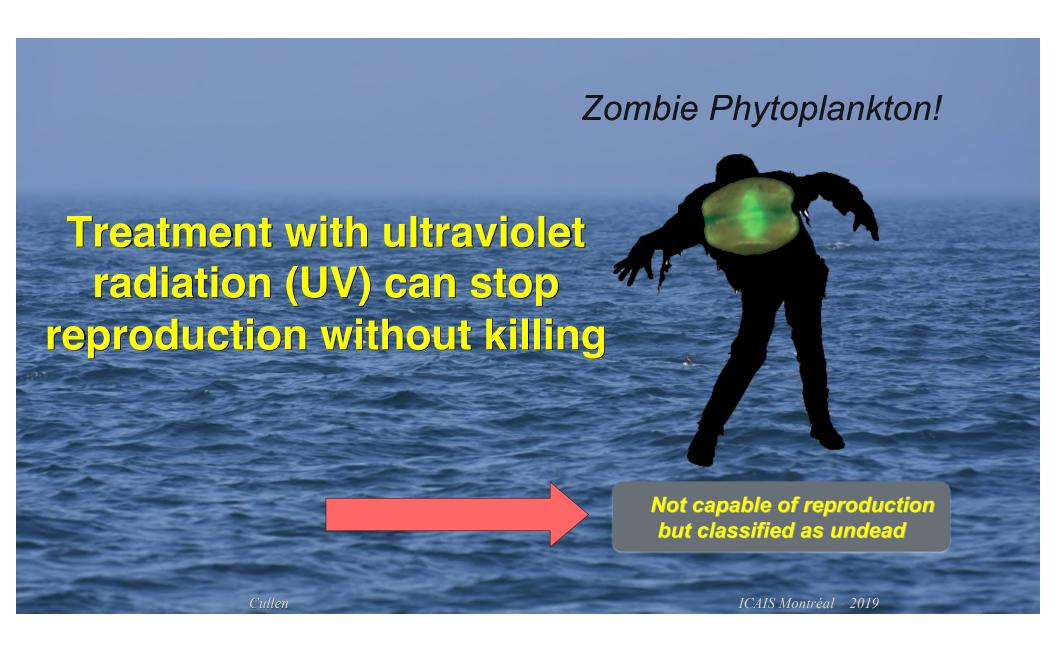


Most Probable Number Dilution Culture (MPN) enumerates viable cells — those capable of reproduction



Heterotrophs are assessed by motility





Super-simple Summary:

Treat with low-energy UV

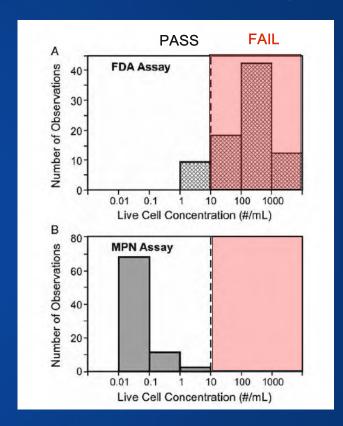
Test with MPN: PASS

Test with stains: FAIL

Correct answer: PASS

See also Blatchley et al, 2018. Environ Sci Technol 52, 8075-8086.

UV-Treated Discharge



Wright and Welschmeyer 2015. J. Mar. Eng. Tech. 14, 9-18

Super-simple solution for the USCG:

Accept MPN Dilution Culture + Motility and align with the IMO

Document prepared with consent of the United States delegation, led by the USCG.

(January 2017)



BWM.2/Circ.61 Annex, page 1

ANNEX

GUIDANCE ON METHODOLOGIES THAT MAY BE USED FOR ENUMERATING VIABLE ORGANISMS FOR TYPE APPROVAL OF BALLAST WATER MANAGEMENT SYSTEMS

Table: Methodologies that may be used for enumerating viable organisms for type approval of BWMS

| Methodologies for enumerating viable organisms | Organism size class or indicator | Assessed criteria of viability | Examples of how the methodologies are applied | Applicability to ballast water treatment technologies |
|---|--|---|---|---|
| FDA/CMFDA + Motility | Viable organisms ≥ 10 µm to < 50 µm | Membrane integrity, enzyme activity, motility | PPR 4/7, appendix 1; PPR 4/INF.10 | Suitable for assessing treatment technologies intended to kill or remove organisms |
| MPN Dilution Culture + Motility | Viable organisms ≥ 10 µm to < 50 µm | Reproduction capacity, motility | PPR 4/7, appendix 2 | Suitable for assessing all treatment technologies |

Consensus agreement



BWM.2/Circ.61 Annex, page 1

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Cullen

USCG cannot test low-energy UV treatment systems suitably

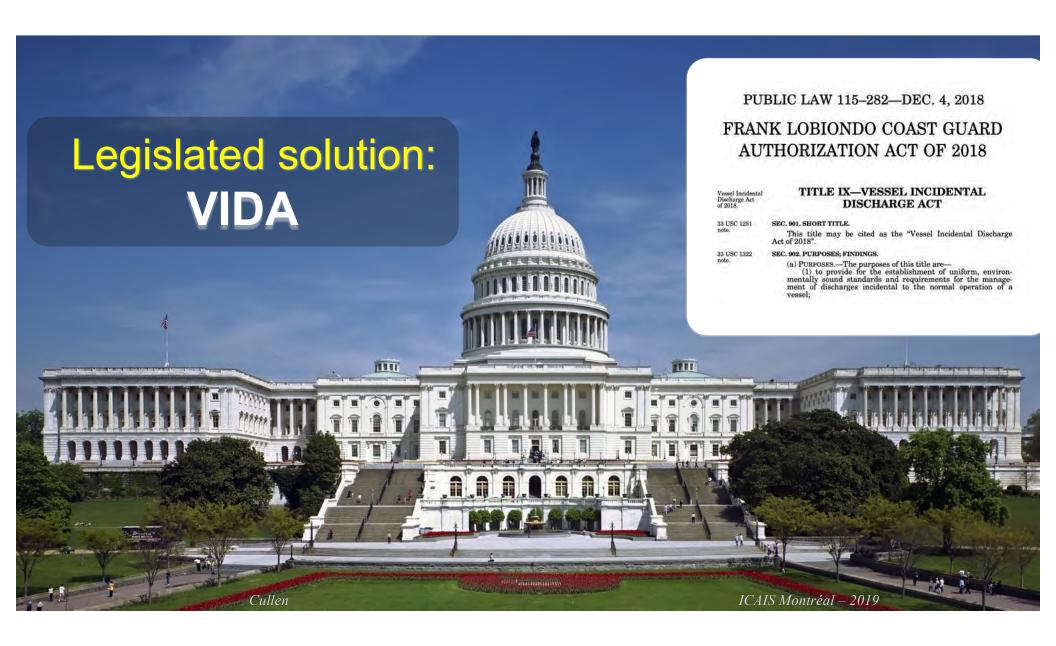


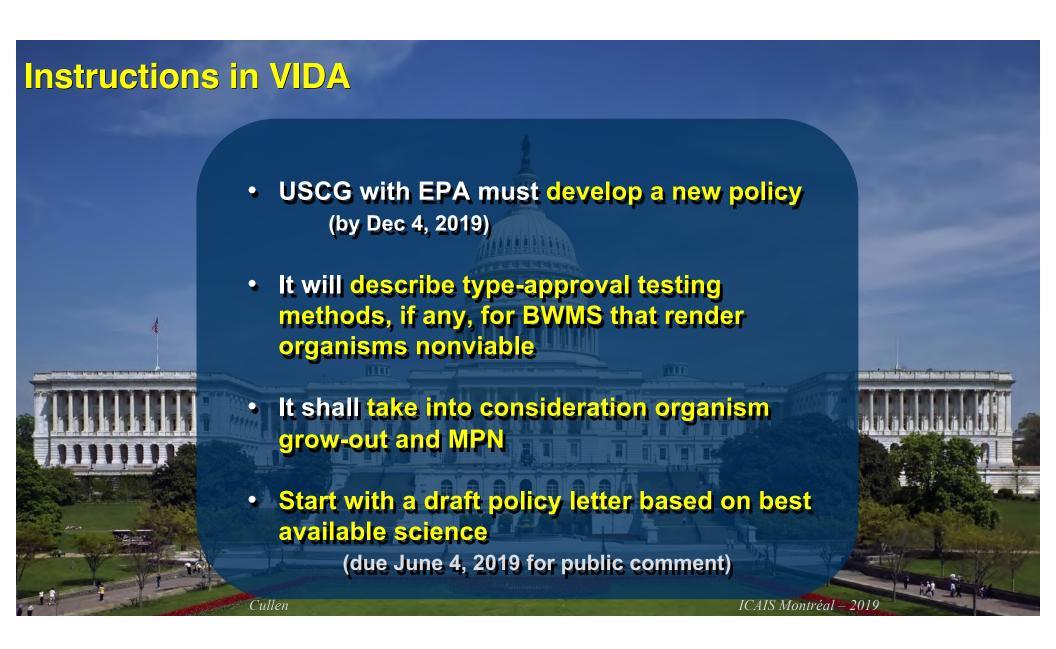
BWM.2/Circ.61 Annex, page 1

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Stains are not suitable for low-energy UV







United States Coast Guard

2763 Martin Lüther King Jr. Ave. S.E., Washington, DC 20593-7509 Staff Symbol: CS-ODES Phone: 202-272-1433 Fax: 202-372-8382 Email: environmental standards@usog.m

16714 CG-OES Policy Letter No. 01-19 26 July 2019



To: Distribution

Subj: ACCEPTANCE OF TYPE APPROVAL TESTING PROTOCOLS FOR BALLAST WATER MANAGEMENT SYSTEMS (BWMS) THAT RENDER ORGANISMS IN BALLAST WATER NONVIABLE.

Ref: (a) Title IX of the Frank LoBiondo Coast Guard Authorization Act of 2018 "Vessel Incident Discharge Act (VIDA)" 33 U.S.C. 1322(p)

(b) Title 46 Code of Federal Regulations (CFR) Part 162.060

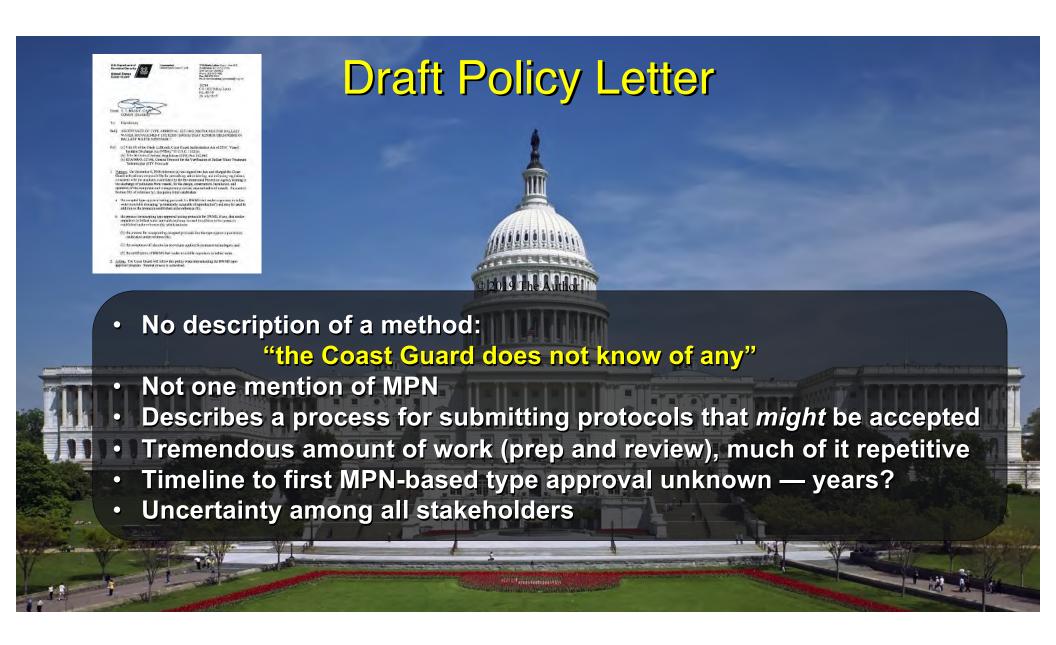
(c) EPA/600/R-10/146, Generic Protocol for the Verification of Ballast Water Treatment Technologies (ETV Protocol)

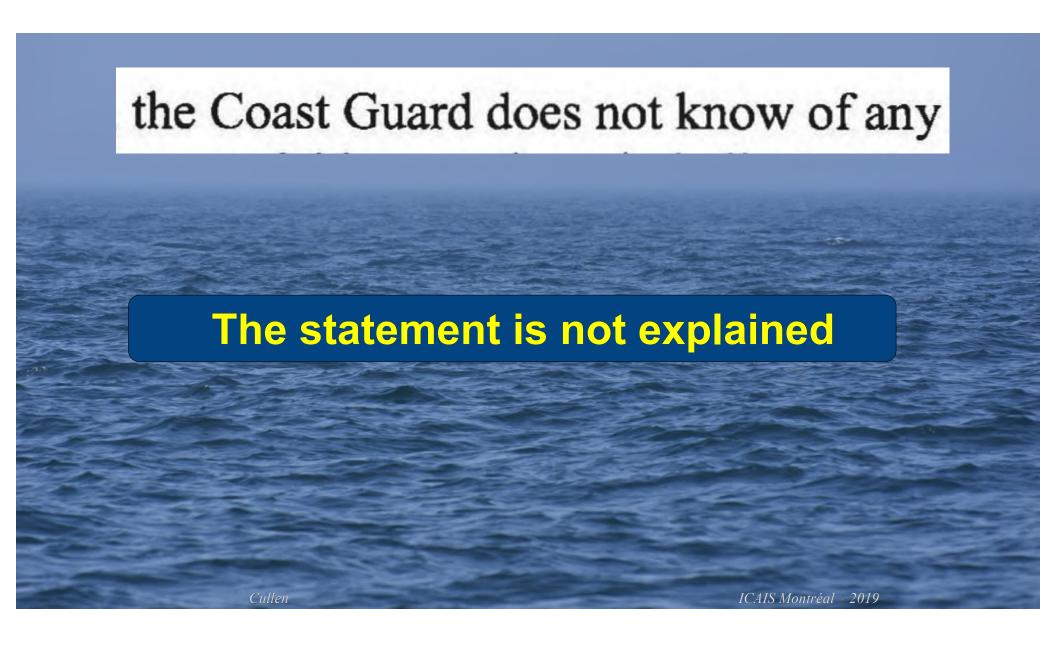
- Purpose. On December 4, 2018 reference (a) was signed into law and charged the Coast
 Guard with primary responsibility for prescribing, administering, and enforcing regulations,
 consistent with the standards established by the Environmental Protection Agency relating to
 the discharge of pollutants from vessels, for the design, construction, installation, and
 operation of the equipment and management practices required onboard vessels. Pursuant to
 Section 903 of reference (a), this policy letter establishes:
- the accepted type-approval testing protocols for BWMS that render organisms in ballast water nonviable (meaning "premanently incapable of reproduction") and may be used in addition to the protocols established under reference (b);
- the process for accepting type-approval testing protocols for BWMS, if any, that render organisms in ballast water nonviable and may be used in addition to the protocols established under reference (b), which includes:
 - the process for incorporating accepted protocols into the type-approval procedures established under reference (b):
 - (2) the acceptance of laboratories to evaluate applicable treatment technologies; and
 - (3) the certification of BWMS that render nonviable organisms in ballast water.
- Action. The Coast Guard will follow this policy when implementing the BWMS typeapproval program. Internet release is authorized.

Draft Policy Letter Surprises Many

...the Coast Guard does not know of any type-approval testing protocols for BWMS that render nonviable organisms in ballast water that are based on best available science.

Cullen









Phytoplankton manual

Edited by A. Sournia

Muséum National d'Histoire Naturelle, Paris

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The dilution-culture method

1978



Si Cite The Drains Sci. Deared 2018 52 8025-8080

The Biological Basis for Ballast Water Performance Standards: "Viable/Non-Viable" or "Live/Dead"?

Ernest R. Blatchley III.* 10 John J. Cullen. Brian Petri. Keith Bircher, and Nicholas Welschmeyer

J Appl Phycol (2018) 30:1073-1094 DOI 10.1007/s10811-017-1254-8

J Appl Phycol (2016) 28:279-298 DOI 10:1007/s10811-015-0601-x

Received: 31 January 2015 / Accepted: 21 April 2015 / Published online: 24 May 2015 © The Author(s) 2015. This article is published with open access at Springerlink.com

On the use of the serial dilution culture method to enumerate viable phytoplankton in natural communities of plankton subjected to ballast water treatment

John J. Cullen 1 · Hugh L. MacIntyre 1



Jahn Throndsen

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journal homepage: www.

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John J. Cullen

Department of Oceanography, Dallousie University, Halling, Nove Scotia 83H 4R2, Canad

Best available science disagrees

Enumerating viable phytoplankton using a culture-based Most Probable Number assay following ultraviolet-C treatment

> 8) 1090-1100 lenceDirect Bulletin

UV Huences required for compliance with pallast water discharge standards using two approved methods for algal viability assessment



Kim Lundgreen ", Henrik Holbech", Knud Ladegaard Pedersen", Gitte Ingelise Petersen , Rune Røjgaard Andreasen^c, Christaline George^d, Guillaume Drillet^{d,e}, Martin Andersen^c

SUB-COMMITTEE ON POLLUTION PREVENTION AND RESPONSE 4th session Agenda item 7



PPR 4/7 12 October 2016 Original: ENGLISH

REVIEW OF THE GUIDELINES FOR APPROVAL OF BALLAST WATER **MANAGEMENT SYSTEMS (G8)**

Analysis methods for determining the viability of organisms in the 10 to 50 µm size class

Submitted by Denmark and Norway



pubs.acs.org/journal/esticu

Algal DNA Repair Kinetics Support Culture-Based Enumeration for Validation of Ultraviolet Disinfection Ballast Water Treatment Systems

Natalie M. Hull, Mythili R. Isola, Brian Petri, Po-Shun Chan, and Karl G. Linden 19

⁷Civil, Environmental, and Architectural Engineering, University of Colorado, Boulder, Colorado 80309, United States "Trojan Technologies, London, ON NSV 4T7, Canada



Phytoplankton manual

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Policy Analysis bubsacs.org/est

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Enumerating viable phytoplankton using a culture-based Most



The dilution-culture method

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Science of the Total Environment 627 (2018) 18

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journal homepage: www.elsevier.com/

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John J. Cullen

Department of Oceanography, Dalhousie University, Halljox, Nove Scotia 83H 482, Canac

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Quick review of the best ***

available science

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n 135 (2018) 1090-1100

UV fluences required for compliance with ballast water discharge standards using two approved methods for algal viability assessment

Kim Lundgreen^{a, a}, Henrik Holbech^a, Knud Ladegaard Pedersen^a, Gitte Ingelise Petersen^b, Rune Røjgaard Andreasen^c, Christaline George^d, Guillaume Drillet^{d, e}, Martin Andersen^f

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Algal DNA Repair Kinetics Support Culture-Based Enumeration for Validation of Ultraviolet Disinfection Ballast Water Treatment Systems

Natalie M. Hull, Mythili R. Isola, Brian Petri, Po-Shun Chan, and Karl G. Linden*

°Civil, Environmental, and Architectural Engineering, University of Colorado, Boulder, Colorado 80309, United States °Trojan Technologies, London, ON NSV 4T7, Canada

Documentation freely available on the permanent public record

The best available science describing type-approval testing methods and protocols for ballast water management systems that render nonviable organisms in ballast water

John J. Cullen

Department of Oceanography, Dalhousie University, Halifax, Nova Scotia B3H 4R2, Canada

© 2019 The Author. This report was submitted to the United States Coast Guard and U.S. Environmental Protection Agency on 4 March 2019 and first posted online 1 May 2019; doi: 10.5281/zenodo.2656597.

MPN Dilution Culture has been used for decades to enumerate viable phytoplankton with unknown culturing requirements

Preliminary Studies of Nanoplankton and Ultraplankton Systematics and Abundance by a Quantitative Culture Method.

E. W. Knight-Jones, Marine Biological Station, Bangor. J. du Conseil

1951

Phytoplankton manual

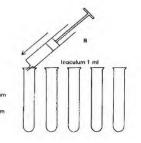
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On the use of the serial dilution culture method to enumerate viable phytoplankton in natural communities of plankton subjected to ballast water treatment 2015

John J. Cullen 1 · Hugh L. MacIntyre 1

MPN Dilution Culture + Motility has been used for years in type-approval testing of BWMS and has been fully endorsed by USCG-approved laboratories

USCG-accepted Independent Laboratory:



Contributed to 14 of the 21 USCG type approvals

Contributed to 16 type approvals for Norway (IMO)

USCG-accepted testing facilities:







MPN since 2010

MPN since 2010

MPN since 2006

Cullen

Suitable testing protocols have been published They are based on best available science

SUB-COMMITTEE ON POLLUTION PREVENTION AND RESPONSE 4th session Agenda item 7



PPR 4/7 12 October 2016 Original: ENGLISH

REVIEW OF THE GUIDELINES FOR APPROVAL OF BALLAST WATER MANAGEMENT SYSTEMS (G8)

Analysis methods for determining the viability of organisms in the 10 to 50 µm size class

Submitted by Denmark and Norway

An Analysis Method for

Determining the Viability of Organisms \geq 10 μ m and < 50 μ m

Using MPN Dilution Culture + Motility

A protocol proposed to the USCG as part of the comments to the CG-OES Policy Letter No. 01-19, dated 26 July 2019

Submitted by

DNV GL (Independent Laboratory to the USCG for type approval testing of BWTS)

DHI (laboratory in Denmark for analyzing the biological efficacy of BWTS)

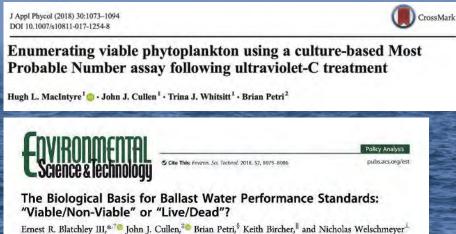
NIVA (laboratory in Norway for analyzing the biological efficacy of BWTS)

GBRC (laboratory in California for analyzing the biological efficacy of BWTS)

Cullen

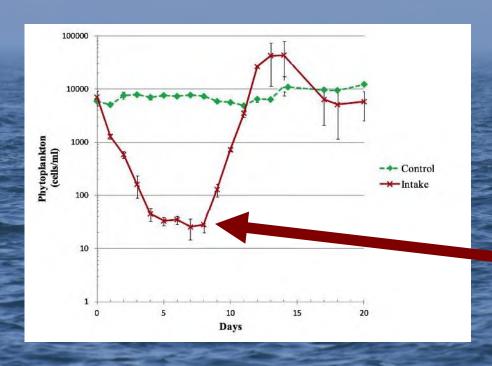
MPN-based methods assess permanent loss of viability





- Damage from UV is repaired quickly or not at all
- The MPN Dilution Culture method promotes any repair that might occur
- Two-week incubation is enough to render the risk of delayed recovery negligible

Is "regrowth" evidence of delayed recovery?



Growth first detected eight days after treatment

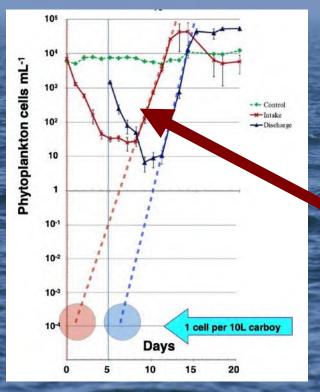
Liebich et al., 2012. Aquatic Invasions 7, 29-36 Shown by Cullen and MacIntyre, ICAIS 2016

ICAIS Montréal – 2019

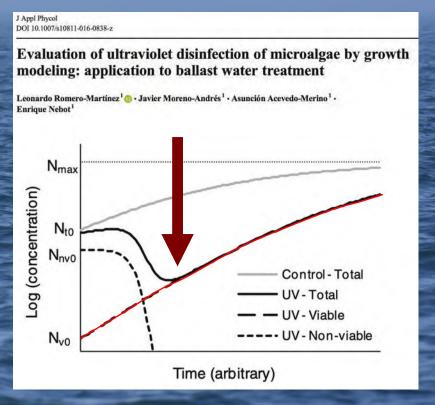
Cullen

Delayed detection is not a demonstration of delayed growth "nonviable means permanently nonviable"

Apparent "regrowth" is survivors emerging



Liebich et al., 2012. Aquatic Invasions 7, 29-36 Shown by Cullen and MacIntyre, ICAIS 2016 *Cullen* Prediction of growth modeling with no delayed recovery



MPN + Motility has been thoroughly validated

J Appl Phycol (2018) 30:1073-1094 DOI 10.1007/s10811-017-1254-8

Agenda item 7



Enumerating viable phytoplankton using a culture-based Most Probable Number assay following ultraviolet-C treatment

Hugh L. MacIntyre 10 · John J. Cullen 1 · Trina J. Whitsitt 1 · Brian Petri 2

SUB-COMMITTEE ON POLLUTION PREVENTION AND RESPONSE 4th session



PPR 4/7 12 October 2016 Original: ENGLISH

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Analysis methods for determining the viability of organisms in the 10 to 50 µm size class

Submitted by Denmark and Norway

6th IMarEST Ballast Water Technology Conference, 12-13 January 2017, London, UK https://doi.org/10.24868/bwtc6.2017.010



The case for using the Most Probable Number (MPN) method in ballast water management system type approval testing

John J. Cullen^a, Hugh L. MacIntyre^a

^aDepartment of Oceanography, Dalhousie University, Halifax, Nova Scotia, Canada B3H 4R2,

Journal of Applied Phycology (2019) 31:491-503 https://doi.org/10.1007/s10811-018-1541-z



Inter-laboratory validation of the serial dilution culture—most probable number method for enumerating viable phytoplankton

Hugh L. MacIntyre ¹ ... John J. Cullen ¹ · Shannah Rastin ¹ · Magdalena Waclawik ¹ · Kimberly J. Franklin ¹ · Nicole Poulton ² · Laura Lubelczyk ² · Kate McPhee ² · Tammi L. Richardson ³ · Elise Van Meerssche ³ · Brian Petri ⁴

Science of the Total Environment 627 (2018) 1602-1626



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Science of the Total Environment



journal homepage: www.elsevier.com/locate/scitotenv

Review

Quantitative framework for validating two methodologies that are used to enumerate viable organisms for type approval of ballast water management systems



John J. Cullen

Department of Oceanography, Dalhousie University, Halifax, Nova Scotia B3H 4R2, Canada

J. Phycol. 52, 572-589 (2016)

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DOI: 10.1111/jpy.12415

CLASSIFICATION OF PHYTOPLANKTON CELLS AS LIVE OR DEAD USING THE VITAL STAINS FLUORESCEIN DIACETATE AND 5-CHLOROMETHYLFLUORESCEIN DIACETATE 1

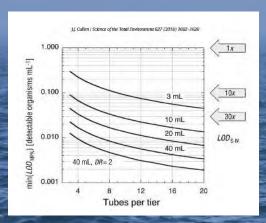
Hugh L. MacIntyre2 and John J. Cullen

Department of Oceanography, Dalhousie University, PO Box 15000, Halifax, Nova Scotia B3H 4R2, Canada

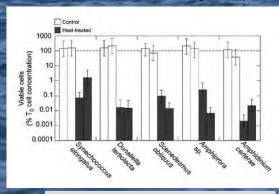
Cullen

Quantitative and rigorous

Limit of detection

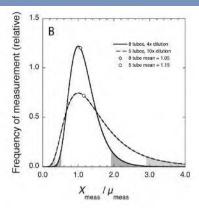


Bias (false positive/negative)

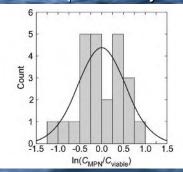


see next slide for reference

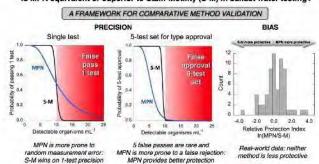
Precision



Interlaboratory comparison Reproducibility



Is MPN equivalent or superior to Stain-Motility (S-M) in ballast water testing?



Expert assessment of bias

| | Bias on number of living 10 to 50 µm organisms | Probable magnitude |
|--|--|--|
| Autotrophs | | |
| Inclusion of > 50 µm autotrophs Water samples are not screened through a 50 µm filter to remove > 50 µm autotrophs, which can result in an overeatimation bias. | Overestimate | In many cases this bias is expected to be small, as > 50 µn autotrophs are typically rare (< 3% Chi in > 50 µm size fraction, Welsonmeyer, personal communication). Additionally, the majority of BWMS include a filtration step designed to remove the organisms in the > 50 µm size |
| | | class down to < 10 organisms/m². Where filtration is part of the treatment, the Discharge Treated sample will have very few organisms that ar > 50 µm, reducing the potential overestimation bias. |
| Inclusion of <10 µm autotrophs Samples may or may not be filtered on a 10 µm filter and resuspended, to remove < 10 µm autotrophs. Where 10 µm filtration is not employed, < 10 µm autotrophs will be present in the sample. This will result in an overestimation bias. | Overestimate | This bias can be large depending upon the mixtures of taxa in a given sample. |
| Non-growing autotrophs Some species of autotrophs may not be able to grow (reproduce) in the supplied culture conditions (media, temperature, light). This can result in an underestimation bias. | Underostimate | This bias can be minimized by providing oplimal incubation conditions (general growth media; preparation of media using sterlized local water to which the community is already acclimated, incubation temperatures near ambient, low to intemperatures and under the soft and the state of the state |

Validated to a higher standard than for FDA/CMFDA + Motility

USCG-2019-0477-0007

Comment to the USCG draft policy letter CG-OES No. 01-19
John J. Cullen

https://www.regulations.gov/document?D=USCG-2019-0477-0007

| Performance attributes | Validation data provided? | | |
|---|------------------------------------|--------------------------------|--|
| | FDA/CMFDA + Motility validation | MPN + Motility documents | |
| Range of resolution | NO | YES | |
| Lower limit of detection | NO | YES | |
| Upper limit | Not applicable | YES | |
| Precision | NO | YES | |
| Repeatability (base method, laboratory) | МО | YES | |
| Reproducibility (base method, laboratory) | NO | YES | |
| Trueness / bias (base method, laboratory) | NO | YES | |
| Trueness / bias (natural samples) | YES (partial) | YES (partial) | |
| Quantitative acceptance criteria for validation | NO | YES | |
| Quantitative comparative validation | Not applicable at the time | YES (equivalent protection) | |
| Status of Method Validation (USCG) | Accepted | Not accepted | |

Equivalent or better protection

Science of the Total Environment 627 (2018) 1602-1626

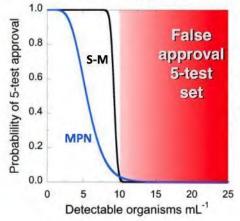
Is MPN equivalent or superior to Stain-Motility (S-M) in ballast water testing?

A FRAMEWORK FOR COMPARATIVE METHOD VALIDATION



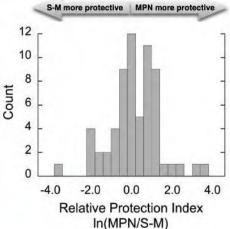
MPN is more prone to random measurement error: S-M wins on 1-test precision

5-test set for type approval

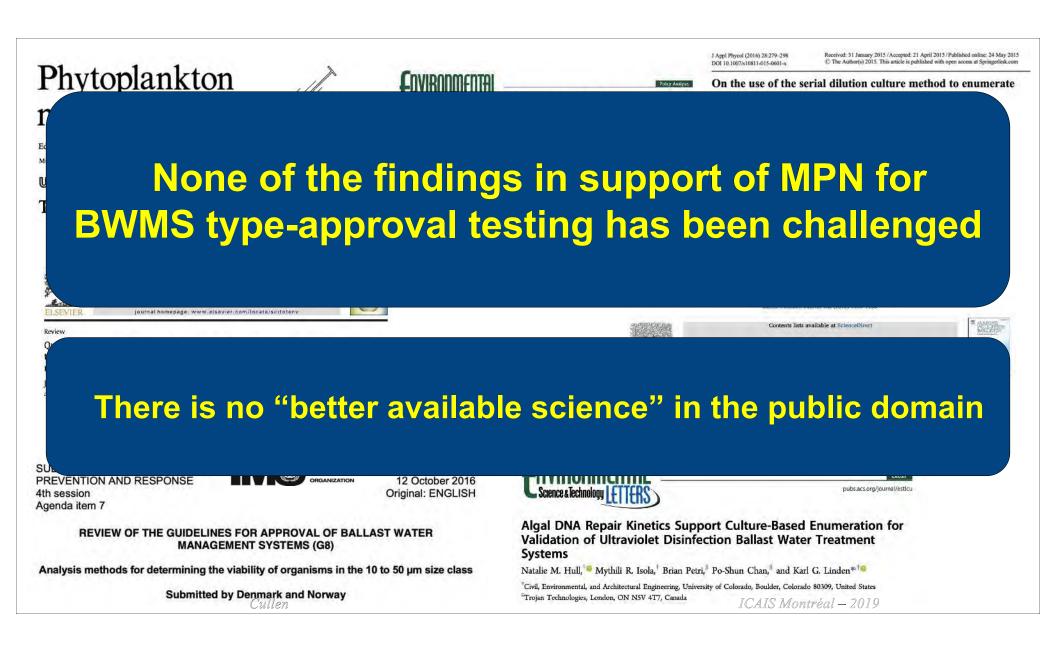


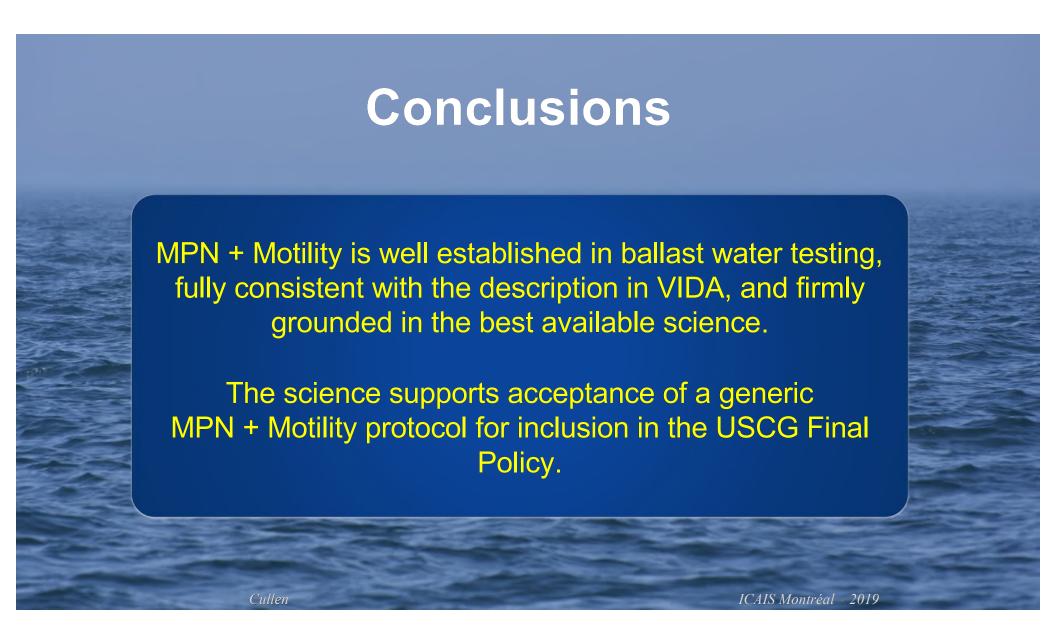
5 false passes are rare and MPN is more prone to a false rejection: MPN provides better protection

BIAS



Real-world data: neither method is less protective





Benefits of accepting MPN + Motility and moving on with protection of coastal environments

- Alignment with IMO and Canadian ballast water policy
- Equivalent or better protection of the environment
- Consistent and predictable framework for shipowners
- Path for approval of low-energy UV treatment
- Eliminate unnecessary and burdensome work
 - manufacturers, laboratories, regulators, contractors, lawyers...
- More resources for Homeland Security and better ballast water treatment

Editorial:



Moving on would be good

Thank you