



# eDNA: Bridging the Gap Between Science and Management

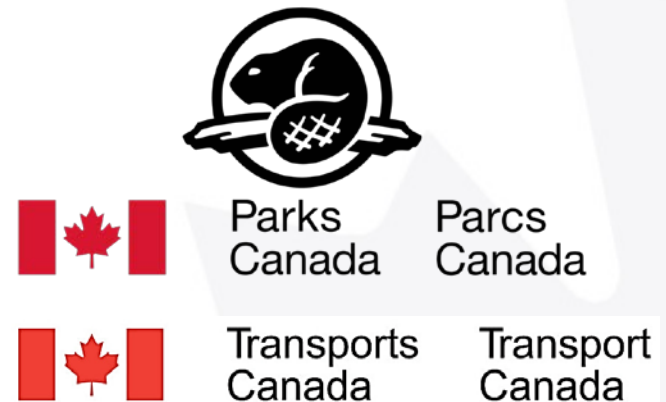
**Stephanie Sardelis**

National Aquatic Invasive Species Advisor, Fisheries and Oceans Canada (DFO)  
International Aquatic Invasive Species Conference 2019

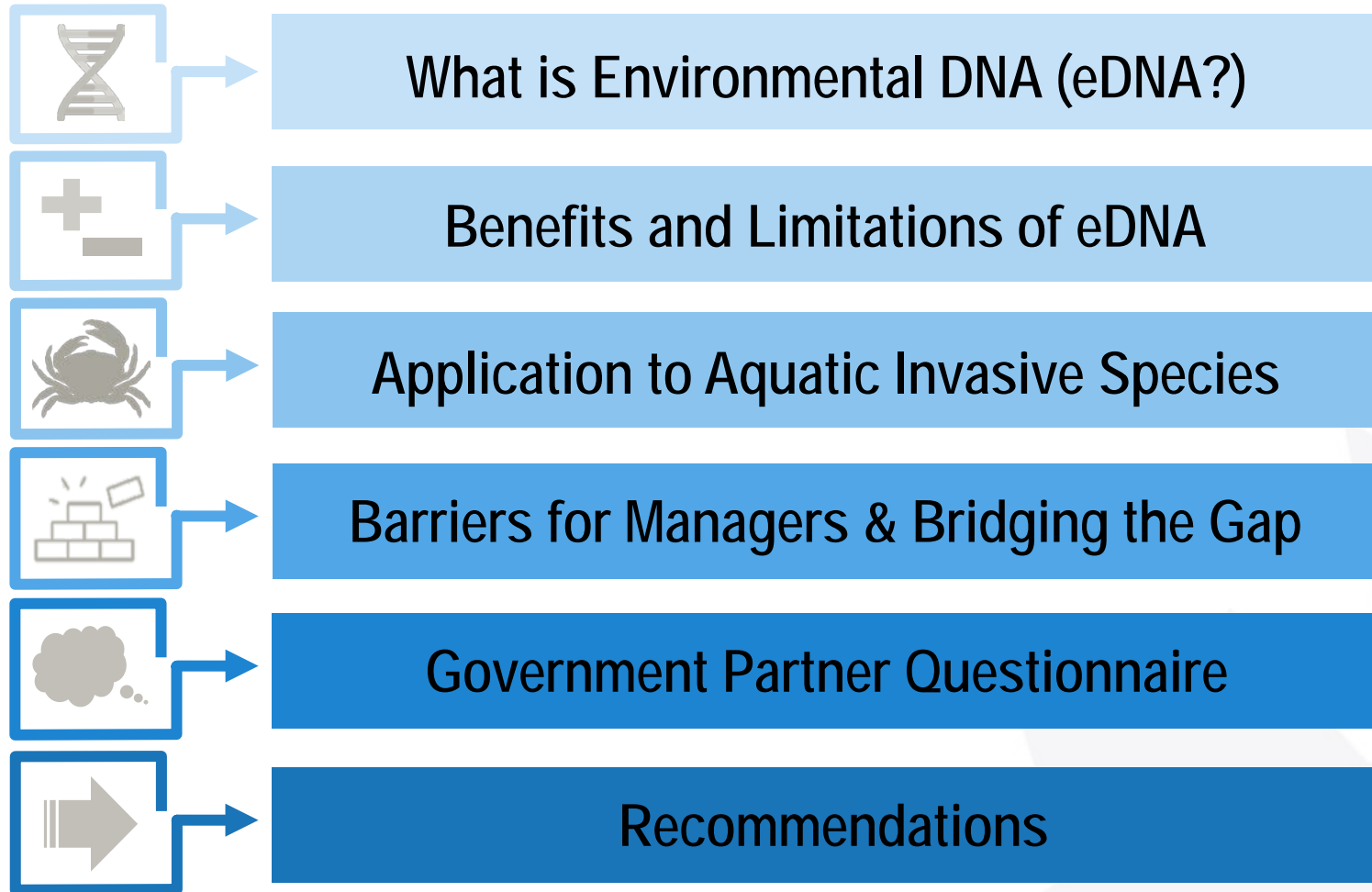
<https://fishbio.com/field-notes/conservation/traces-left-behind>

# Collaborators

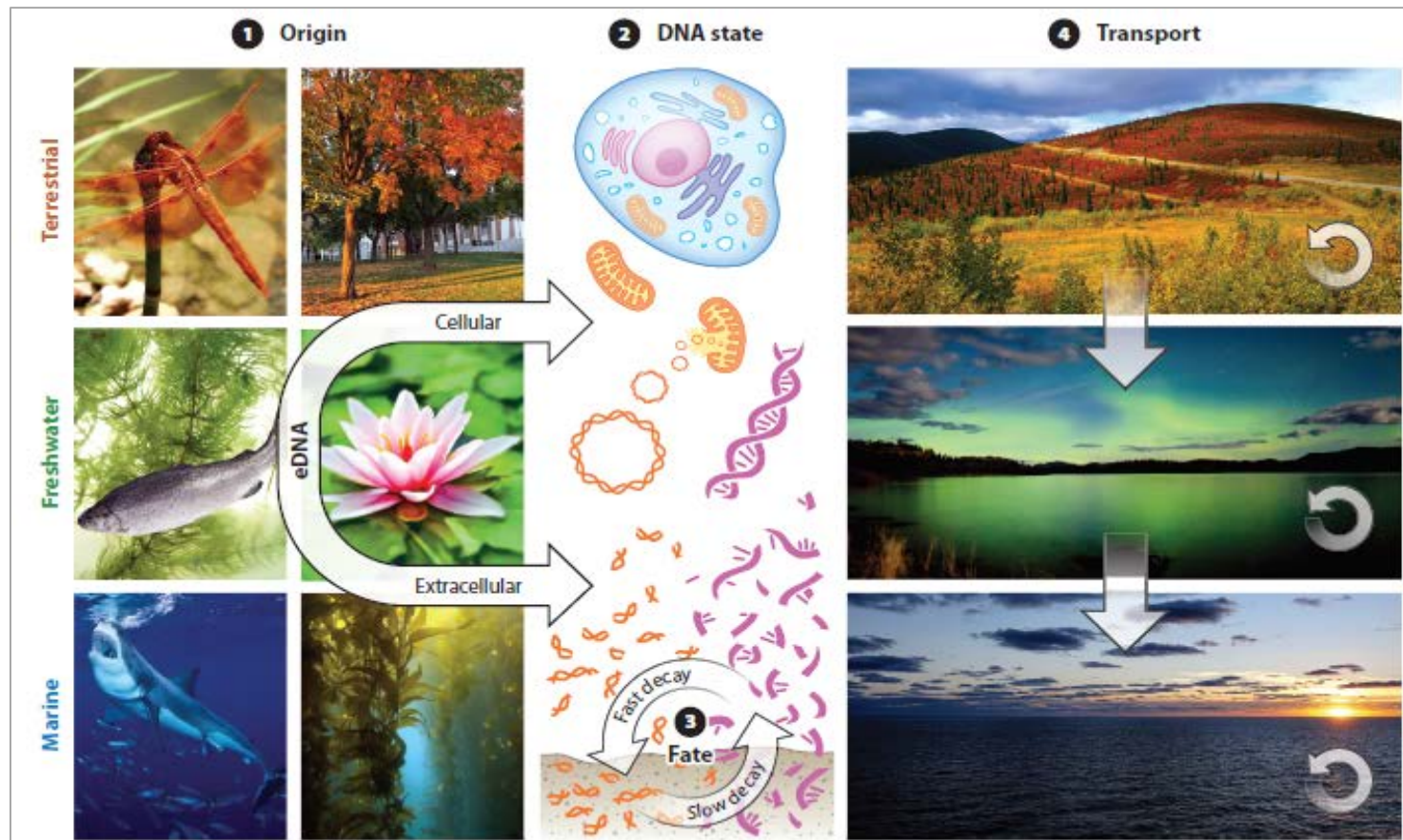
## National Aquatic Invasive Species Committee (NAISC)



# Overview



# What is Environmental DNA (eDNA)?



Credit: Annu. Rev. Ecol. Evol. Syst. 2018. 49:209-230



# Benefits and Limitations of eDNA

Benefits	Limitations
Detection in timely manner improves prevention	Technology is still developing
Many species can be detected simultaneously	Variation in sampling locations/environments; not possible to strictly standardize sampling and analysis protocols
Avoids direct contact with sensitive species	
Less effort than conventional survey methods	Should not be used as a sole indicator of presence
Can cover large/remote areas	Lack of reporting standards and standardized terminology

# Application to Aquatic Invasive Species

## Prevention

- Surveillance of commercial bait and live fish trades (Nathan et al. 2015; Roy et al. 2017)

## Early Detection

- Identifies areas to focus monitoring
- Ballast water screening
- Sea Lamprey in Great Lakes and Zebra Mussels in Lake Winnipeg (Gingera et al. 2016, 2017)

## Response

- Facilitates decision making to take timely action

## Control & Mgmt.

- May help confirm eradication success after intensive effort
- Long-term monitoring of Asian Carps in Chicago Area Waterway System (USFWS)

# What Can eDNA Tell Us?

## A positive eDNA detection tells us:

- Water sample contains genetic material from a target species

## A positive detection *does not* tell us:

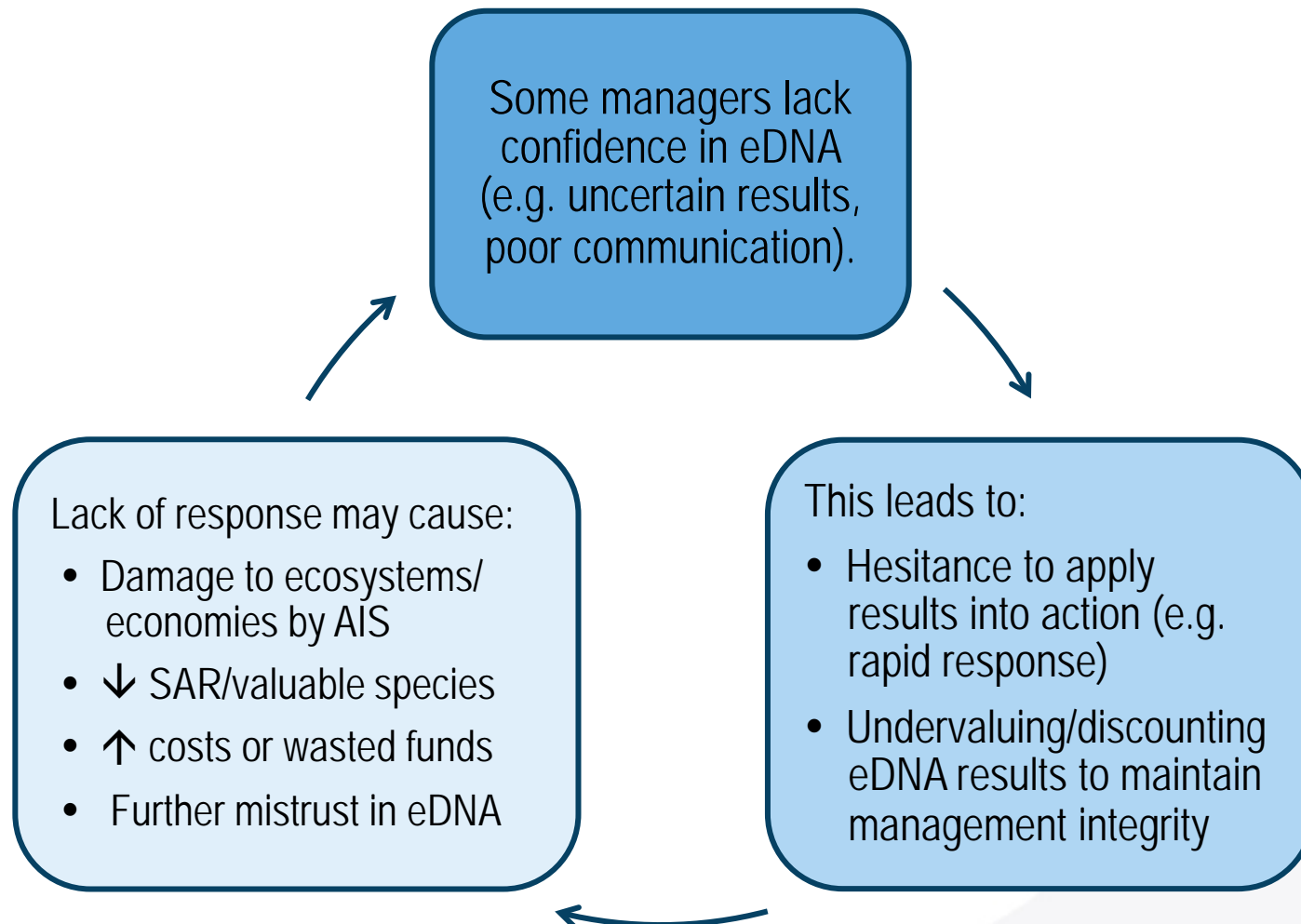
- If the organism is living or dead
- When the organism was present
- Abundance or concentration of organisms in the sample area



\* Positive detections can be false (e.g. sample contamination)

\*\* Spatial/temporal replication for certainty is often needed before taking action

# Barriers for Managers



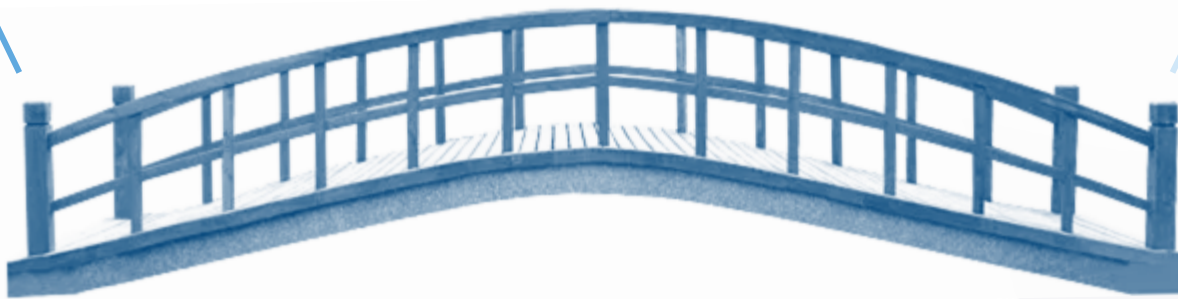


# Bridging the Gap

How should we **respond** to positive eDNA detections?

What **reporting best practices** can scientists and managers develop so results can be effectively applied to decision-making?

**Uptake** by FPT and cross-border partners/scientists/stakeholders



# Recommendations

- Uncertainty must be communicated clearly by scientists to managers
- Reporting standards can help managers interpret results to inform decisions
- Standardized language may further reduce confusion

Reporting Standards	Best Practices	Standard Terminology
<p><i>May include, but not limited to:</i></p> <ul style="list-style-type: none"> <li>• Error sources, direction and magnitude</li> <li>• Minimum information across different methodologies, species and environments</li> <li>• Report format</li> <li>• Mandatory content for consistency across studies                             <ul style="list-style-type: none"> <li>• Descriptions of methodology</li> <li>• Sample size</li> <li>• Statistical power</li> <li>• Uncertainty</li> </ul> </li> </ul>	<p>Develop a communication plan between managers and scientists before projects start</p> <p>Communicate the implications and limitations of eDNA early and clearly to the public</p> <ul style="list-style-type: none"> <li>• e.g. positive is bad, negative is good; eDNA <math>\neq</math> fish, more eDNA <math>\neq</math> more fish</li> </ul>	<p>The repeated use of the word “positive” is not clear and can be misinterpreted.</p> <p>Define:</p> <ul style="list-style-type: none"> <li>• Positive detection</li> <li>• Suspect positive</li> <li>• Confirmed positive</li> <li>• Weak positive</li> <li>• False positive</li> <li>• etc.</li> </ul>

# Conclusion

- eDNA presents a unique opportunity to detect AIS easily, quickly, and cost-effectively, facilitating action to preventing their introduction and spread.
- However, managers must be able to interpret eDNA results if they are expected to inform decision-making.
- Communication between scientists and managers must improve to maximize the applicability of eDNA. In turn, this will improve communicating decisions to the public.
- Our recommendations are complementary to the scientific community continuing to refine methodologies and sampling protocols, etc.



# References

- Baillie et al. 2019. Environmental DNA and its applications to Fisheries and Oceans Canada: National needs and priorities. Can. Tech. Rep. Fish. Aquat. Sci. 3329: xiv + 84 p.
- Gingera et al. 2016. Detection and identification of lampreys in Great Lakes streams using environmental DNA. Journal of Great Lakes Research, 42(3), 649-659. doi: 10.1016/j.jglr.2016.02.017
- Gingera et al. 2017. Environmental DNA as a detection tool for zebra mussels *Dreissena polymorpha* (Pallas, 1771) at the forefront of an invasion event in Lake Winnipeg, Manitoba, Canada. Management of Biological Invasions, 8(3), 287-300. doi: 10.3391/mbi.2017.8.3.03Annu. Rev. Ecol. Evol. Syst. 2018. 49:209-230
- Jerde et al. 2011. "Sight-unseen" detection of rare aquatic species using environmental DNA. Conservation Letters, 4(2), 150-157. doi: 10.1111/j.1755-263X.2010.00158.x
- Nathan et al. 2015. The use of environmental DNA in invasive species surveillance of the Great Lakes commercial bait trade. Conservation Biology, 29(2), 430-439. doi: 10.1111/cobi.12381
- Roy et al. 2017. Development of environmental DNA (eDNA) methods for detecting high-risk freshwater fishes in live trade in Canada. Biological Invasions, 20, 299-231. doi: 10.1007/s10530-017-1532-z

# Contact

## **Stephanie Sardelis**

National Aquatic Invasive Species Advisor

Fisheries and Oceans Canada

[Stephanie.Sardelis@dfo-mpo.gc.ca](mailto:Stephanie.Sardelis@dfo-mpo.gc.ca)

613-293-2495

## **Susan Roe**

National Manager of Aquatic Invasive Species

Fisheries and Oceans Canada

[Susan.Roe@dfo-mpo.gc.ca](mailto:Susan.Roe@dfo-mpo.gc.ca)

613-240-8089