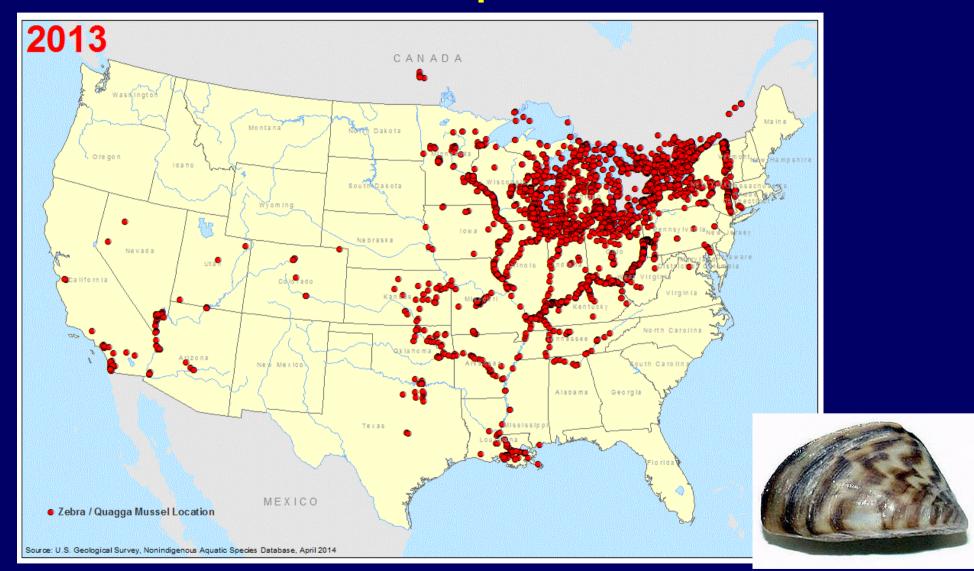
# Feasibility and Efficacy of Three Methods of Zebra Mussel Larvae Detection in Lake Winnipeg

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GLIER, University of Windsor



# **Dreissena spread**



## **Lake Winnipeg**

- Manitoba
- Two basins
- Zebra mussels
  - 2013 detected in south basin
  - 1 veliger reported in the north basin in October 2015
- Low abundance throughout



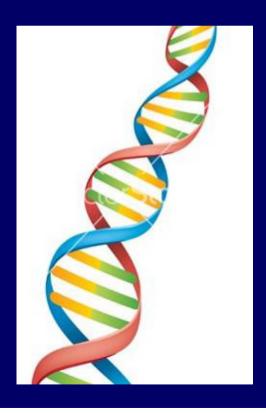
Janusz (2015), personal communication

## **Project goals**

- Detection of zebra mussels in Lake Winnipeg
- Comparison of methods of detection





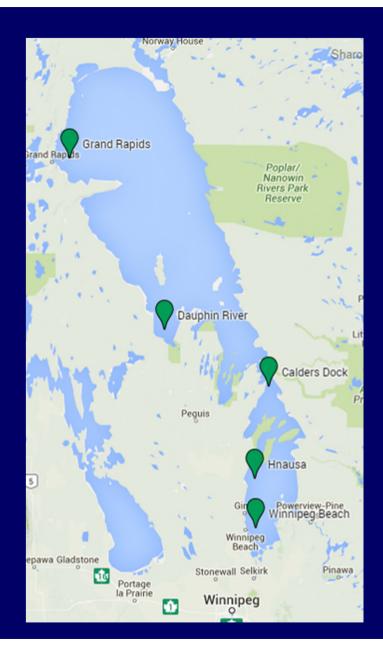


#### **Null Hypotheses**

- Increased sampling intensity does not change probability of detection of NIS
- No difference in detection success in the south and north basin
- Different methods of detection would result in the same success rates

#### **Methods**

- July 2015
- Sampling sites
  - Grand Rapids
  - Dauphin River
  - Calder's Dock
  - Hnausa
  - Winnipeg Beach



#### **Methods**

- 100 plankton tows at each site
  - 64µm plankton nets
- Plankton collected and stored in 95% ethanol
- Filtered out plankton larger than 300µm
- Concentrated into 50mL



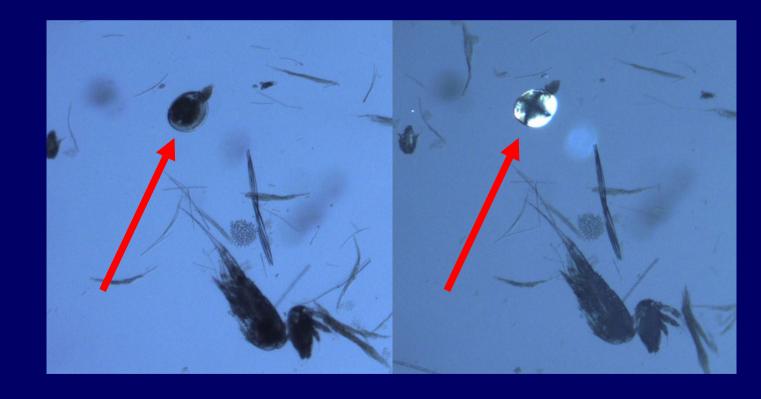




# **Conventional Microscopy**

- Cross-polarized light microscopy
  - Entire sample analyzed and enumerated
  - Distinct shape





Frischer et al. (2012); Johnson (1995)

## Flow Cytometry and Image Analysis

- Flow Cytometer And Microscopy (FlowCAM)
  - Fitted with XPL (cross-polarized lenses)
  - Subsample (3/50 of filtered)
  - Images of each particle is captured and stored for analysis

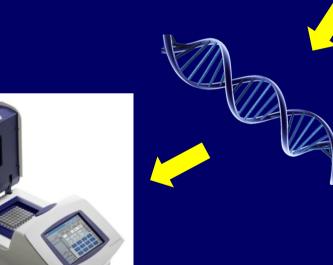




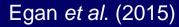


## **Environmental DNA (eDNA)**

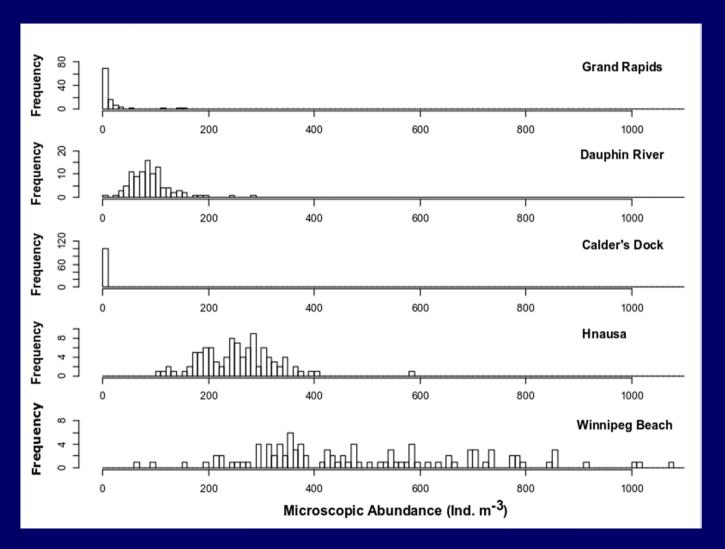
- DNA detection
  - Subsample (1/50) of filtered plankton
  - DNA extraction
  - PCR amplified with species-specific primers

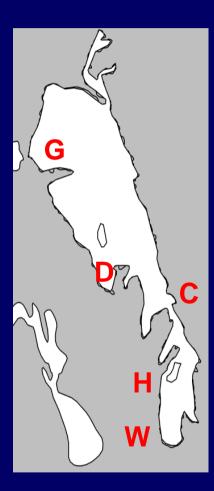




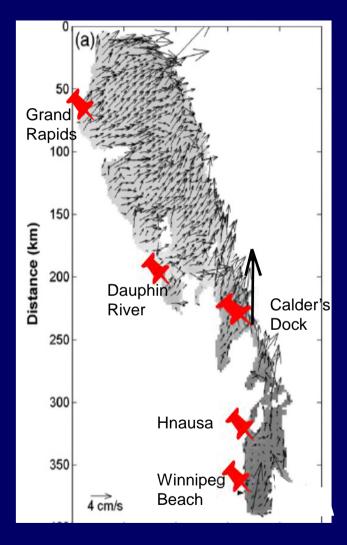


# **Microscopy: Abundance**

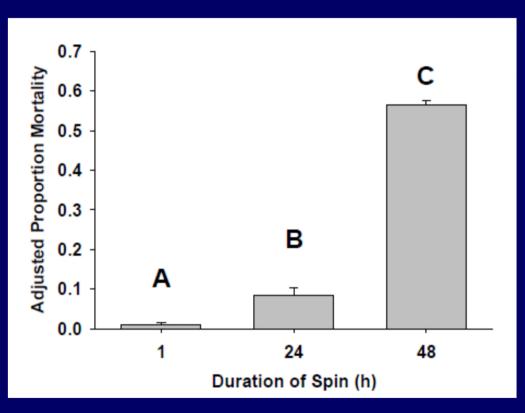




#### **Flow Rate**

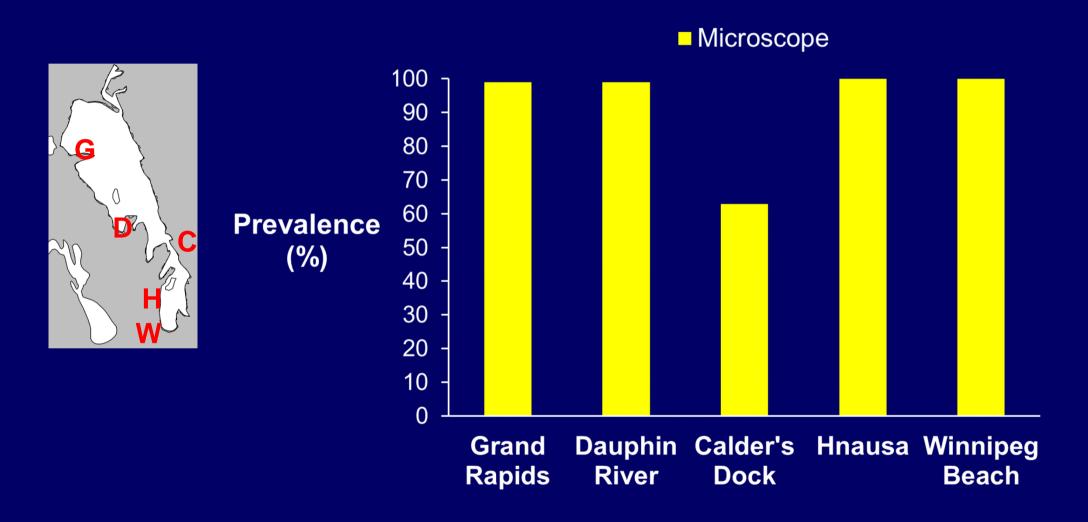


#### **Increased Mortality**

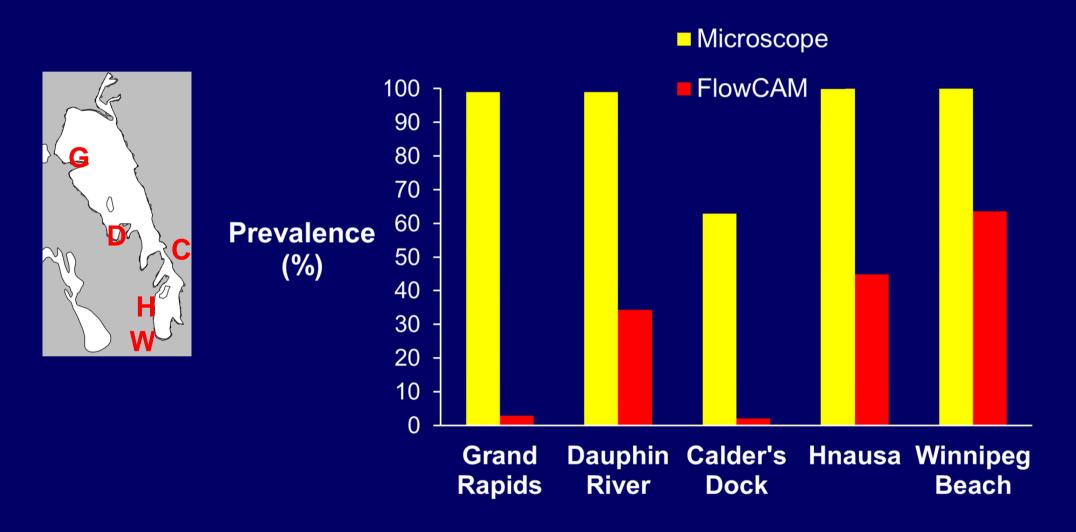


Horvath & Crane (2010)

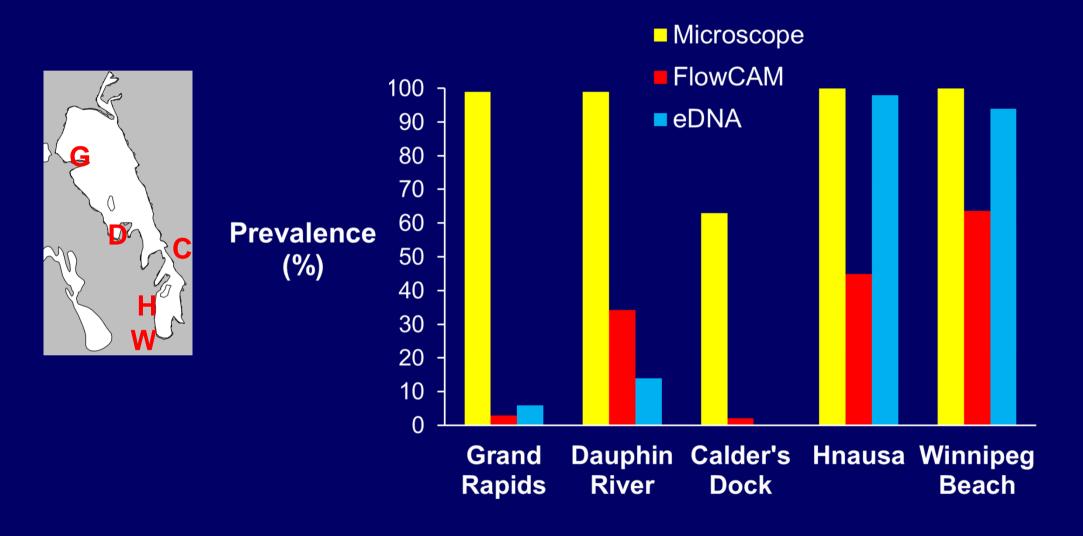
# **Veliger Prevalence in Samples**

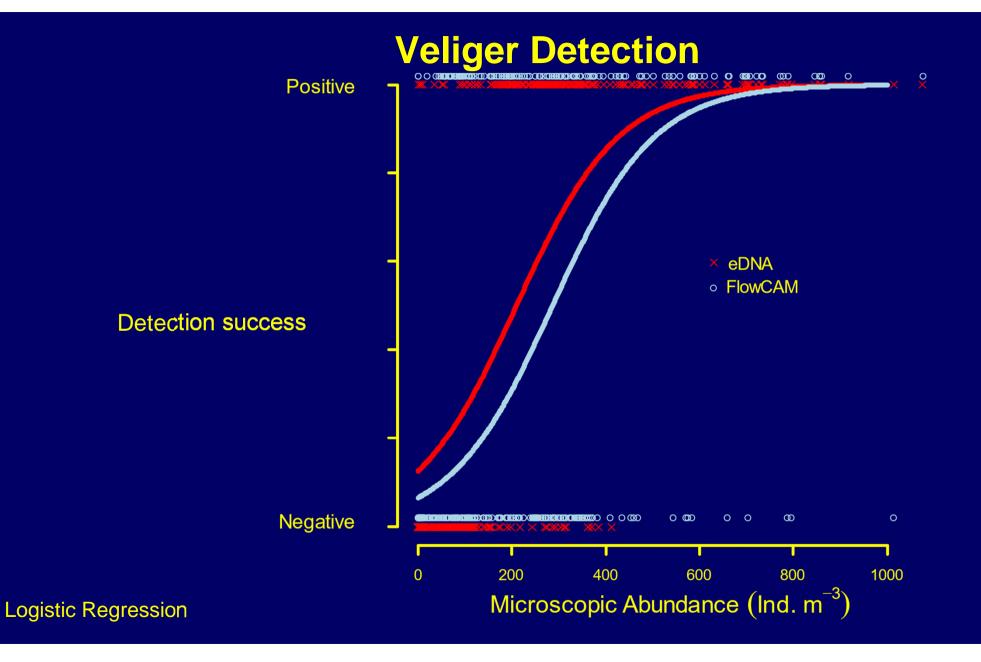


# Veliger Prevalence in Samples



# Veliger Prevalence in Samples





# **Cost and time**

	Microscopy	FlowCAM	eDNA
Start up cost (CAD\$)	10,441.60	81,499.73	5,835.00

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Time (Hours)	198	167	24

#### **Limitations**

- Possible artifact of subsampling
- FlowCAM volume restricted by time taken to process
- eDNA volume restricted by equipment

## **Significance**

- Changes in sampling strategy
- Detection methods
  - Microscopy mid-range in cost and longest time taken, but highest sensitivity
  - eDNA lowest cost and fastest relatively high sensitivity

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- Dr. Doug Haffner
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- Ian MacIsaac
- Sarah-Jayne Collins









Multiple Stressors and Cumulative Effects in the Great Lakes:

An NSERC CREATE Program to Develop Innovative Solutions through International Training Partnerships