

Implementing DNA metabarcoding as cost-effective tool to provide biological data for port baseline survey



Presented by : Anaïs Rey (Ph.D student)
Supervised by : Dr Naiara Rodríguez-Ezpeleta and Dr Oihane C. Basurko

Introduction: Non-indigenous Species introduced with ballast water into ports



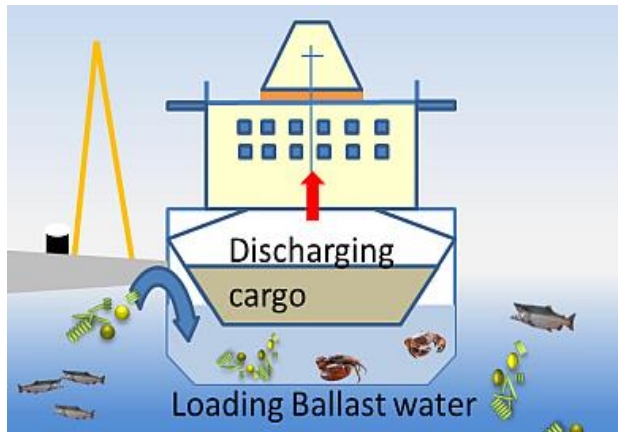
- Shipping carries about **90% of world trade**

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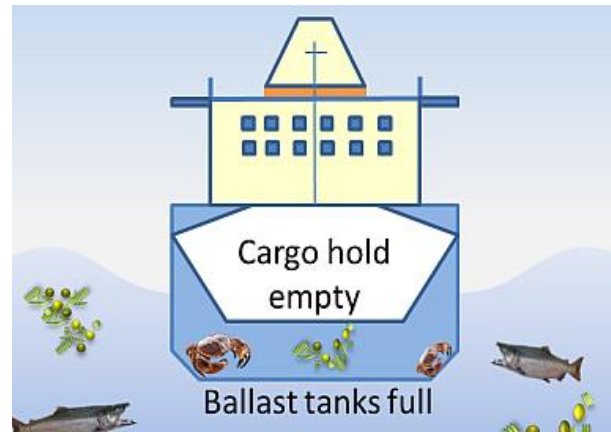


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- **10 billion tones of ballast water** are moved globally each year
- **7,000 species** are daily transferred around the world with ballast water

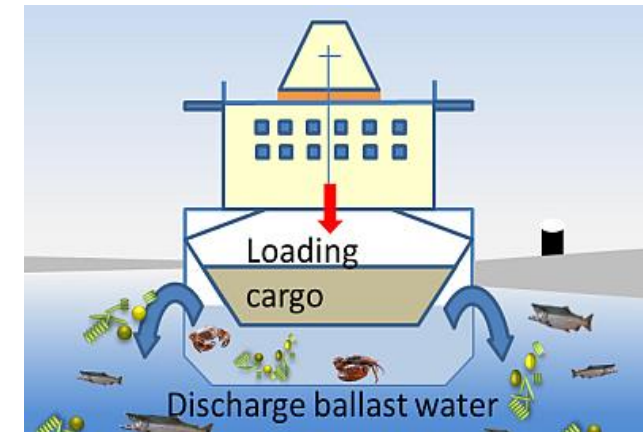
At source port



During voyage



At recipient port



<https://www.nioz.nl/ballastwater-en>

Introduction: Non-indigenous Species introduced with ballast water into ports



- Shipping carries about **90% of world trade**
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- Major vector of **introduction of Non-Indigenous Species, pathogens and harmful algae**

Economical impact

Zebra mussel (*Dreissena polymorpha*)



- Costs for management \$500 millions/year in Great Lakes

Environmental impact

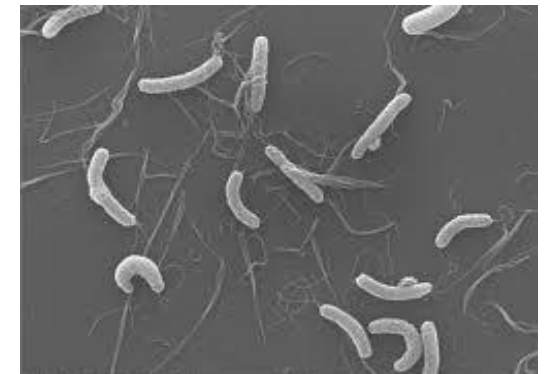
Chinese crab (*Eriocheir sinensis*)



- Food web altered
- Native biodiversity reduced
- Physical disturbances

Social impact

Cholera (*Vibrio cholera*)



- Human disease

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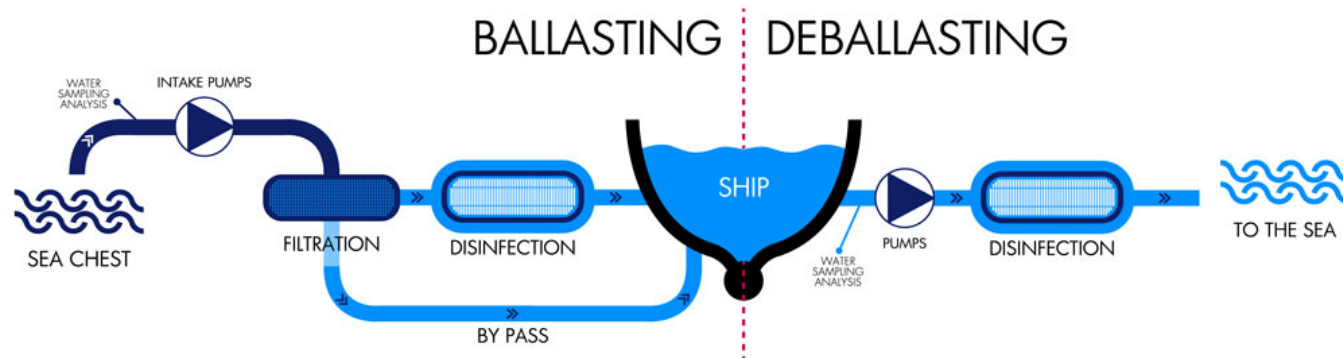


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- To prevent further biological invasions, the **Ballast Water Management Convention** has been established and is currently into force



All ships in international travel must manage their ballast waters and sediments:

- Ballast Water Exchange in open sea
- Ballast Water Treatment Systems



Technology in accordance with existing law regulations (IMO D - 2 STANDARDS)
simple innovative method of treatment of ballast water for any type of vessel

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Ballast water monitoring needed:

Developing tools that will provide biological data to assess the compliance to the Ballast Water Management Convention

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Focus on DNA-based detection of species as a solution:

Assessing the usefulness of genetic tools for ballast water and ports monitoring



Contents lists available at [ScienceDirect](#)

Journal of Sea Research

journal homepage: www.elsevier.com/locate/seares

The challenges and promises of genetic approaches for ballast water management

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Introduction: Port baseline surveys



Monitoring applications extracted from Guidelines

- ✓ Risk assessment for granting exemptions: port baseline survey & target species detection

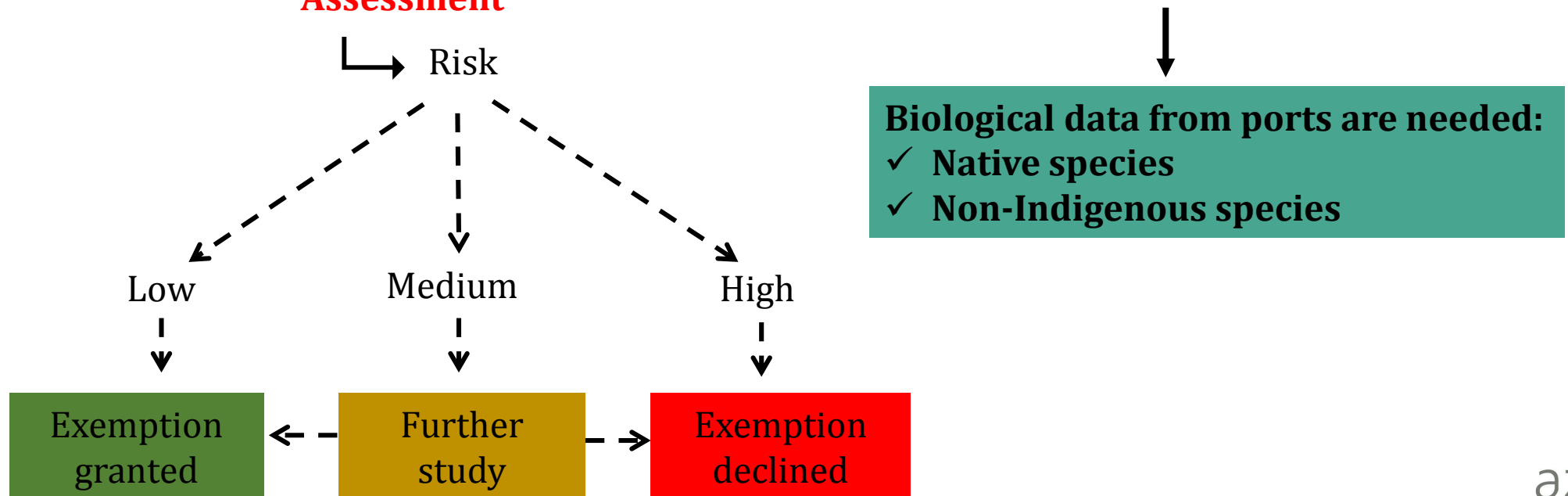
Example:

Shipowner looking an exemption for the specific voyage Rotterdam (Netherland) and Le Havre (France)

↳ Ask for an exemption

↳ **Scientific Risk Assessment**

→ Is there a risk of harmful organisms introduction ?



Objectives

To ease complex and expensive port baseline surveys

➔ **By developing of a protocol based on the use of DNA metabarcoding**

- The taxonomic assignment of individuals from an environmental sample based on their DNA sequences

➔ **Where specific objectives will be assessed:**



❖ **Reliability of the DNA metabarcoding port baseline survey to:**

- Detect Non-Indigenous Species
- Retrieve the biological communities of the port

❖ **The potential of using environmental DNA to retrieve similar communities**

❖ **The importance of seasonal sampling**

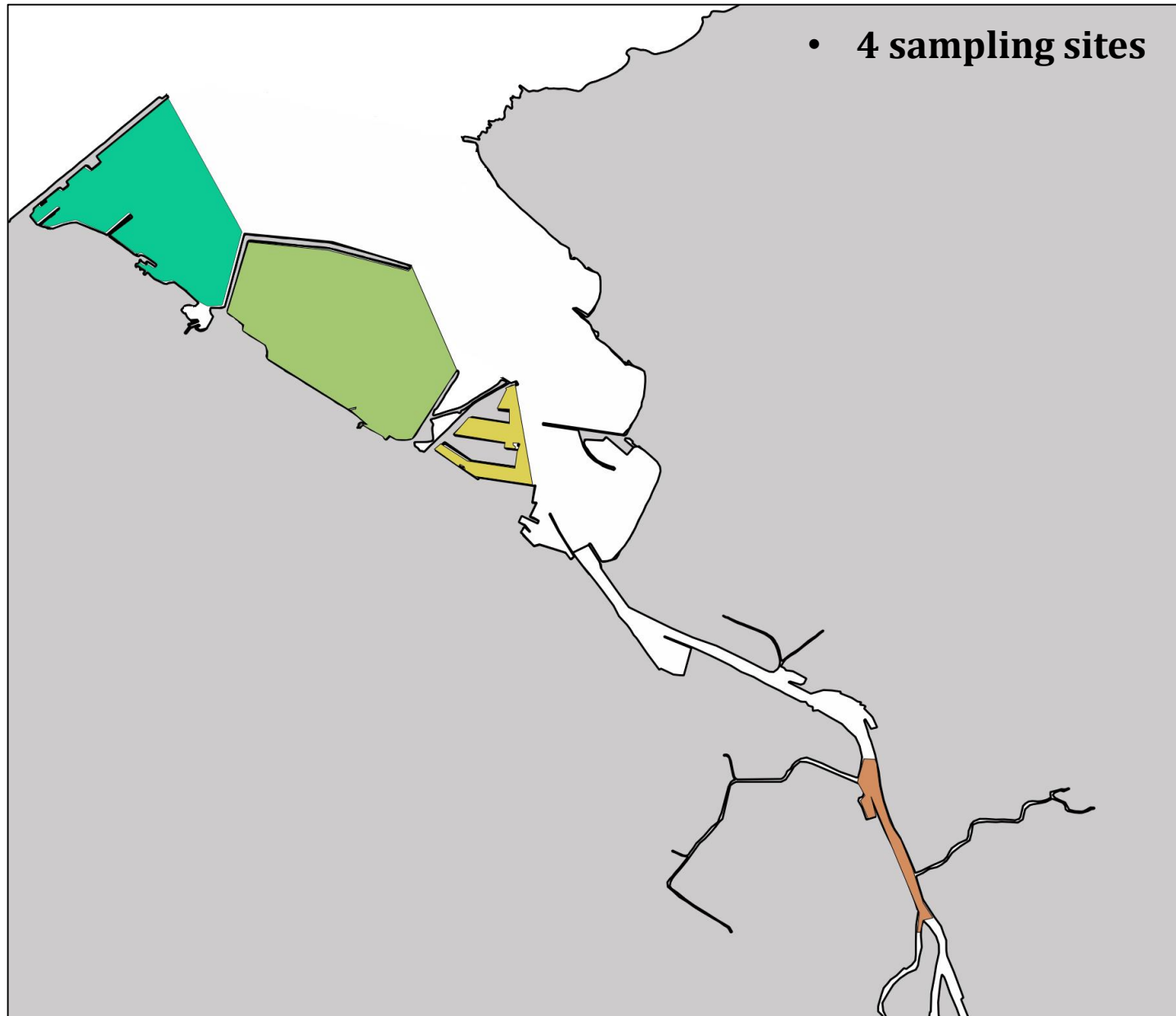
Materials and methods: The port studied

→ Port of Bilbao, open on the Atlantic ocean (Bay of Biscay), Northern Spain, Europe



<http://www.breakbulk.com/bilbao-port-traffic-rises>

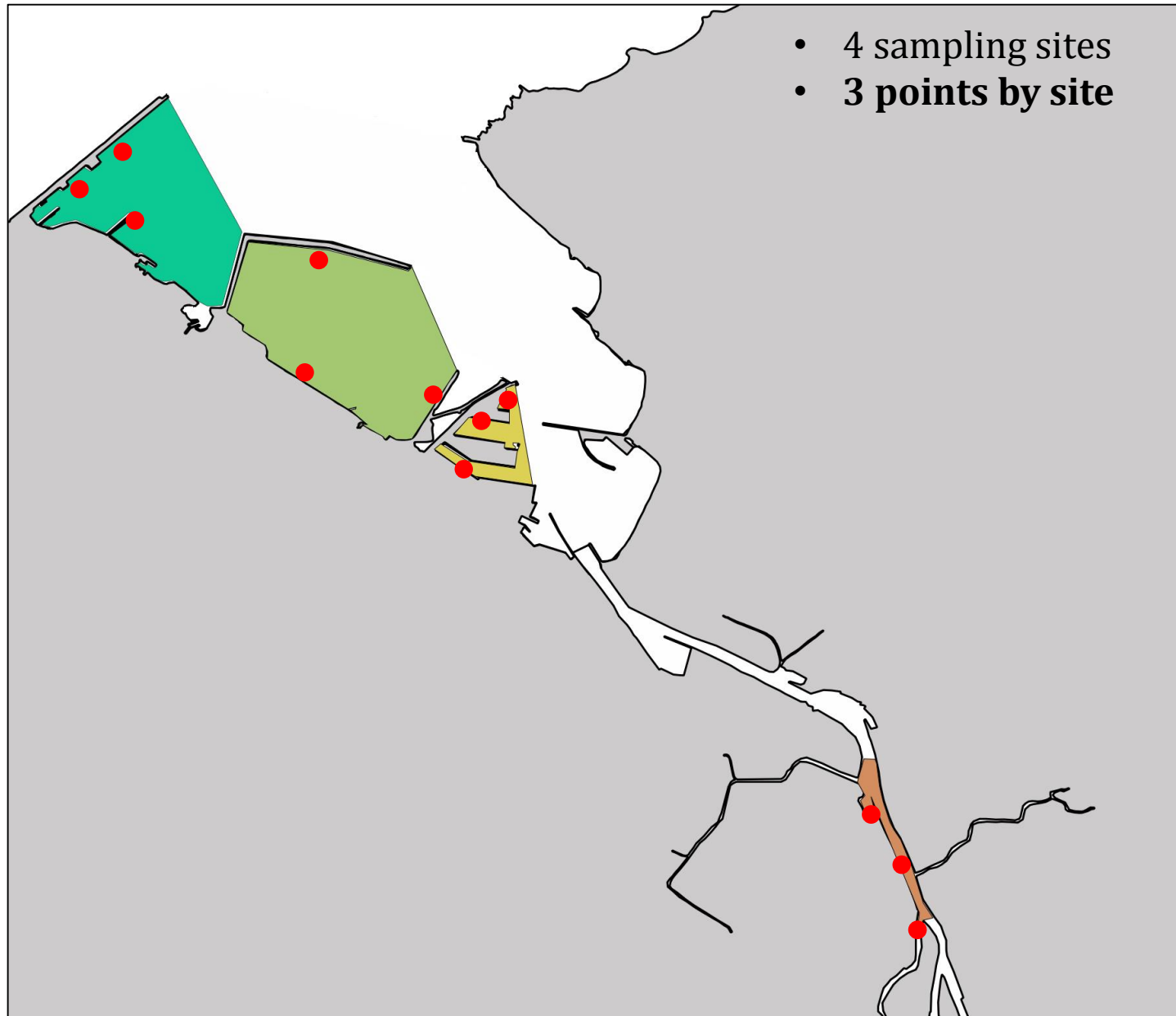
Materials and methods: The sampling design



4 Sampling periods:

Autumn 2016
Winter 2017,
Spring 2017
Summer 2017

Materials and methods: The sampling design



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Winter 2017,
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Materials and methods: The sampling design



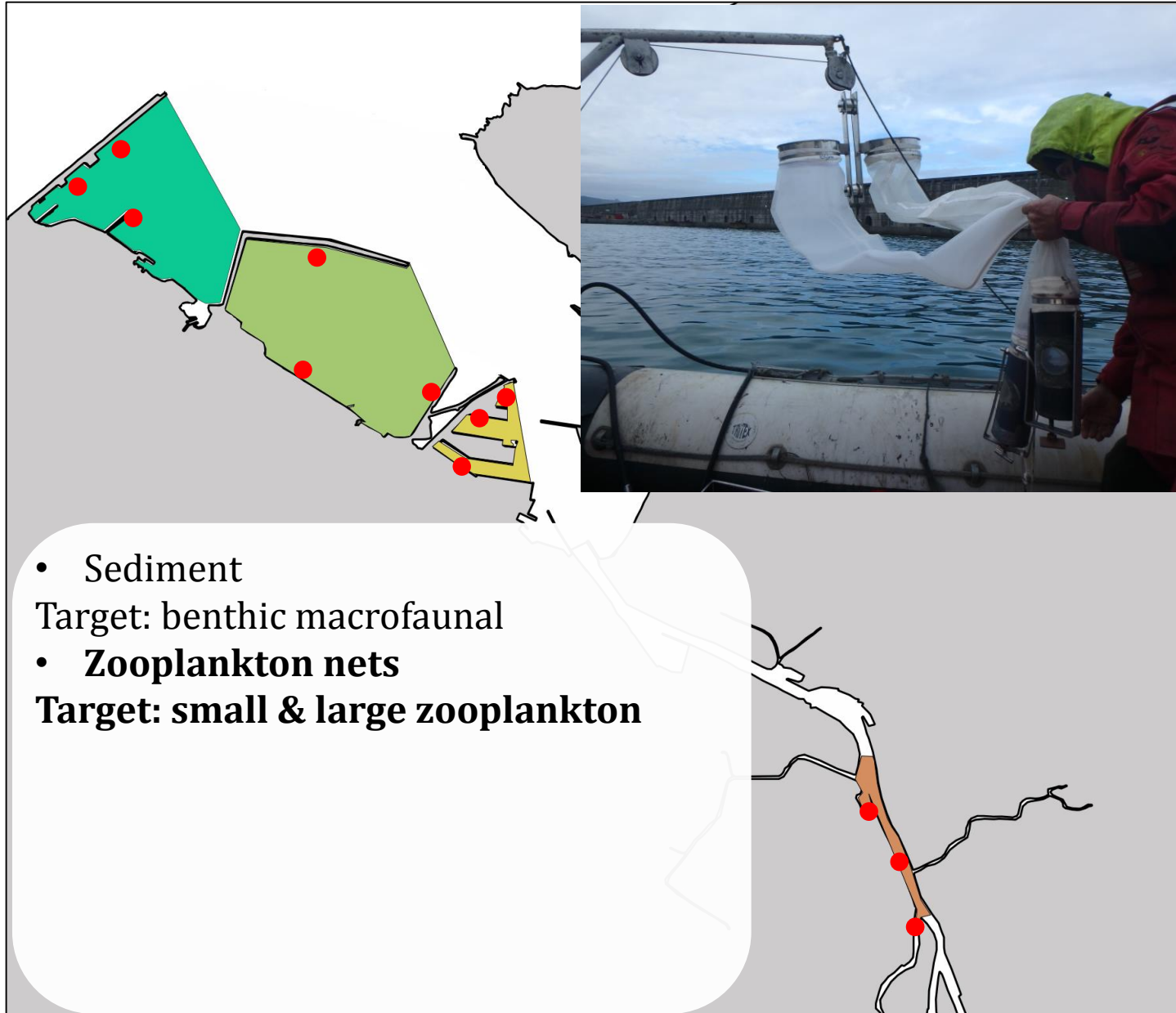
- **Sediment**
Target: benthic macrofaunal



4 Sampling periods:

Autumn 2016
Winter 2017,
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Materials and methods: The sampling design



4 Sampling periods:

Autumn 2016

Winter 2017,

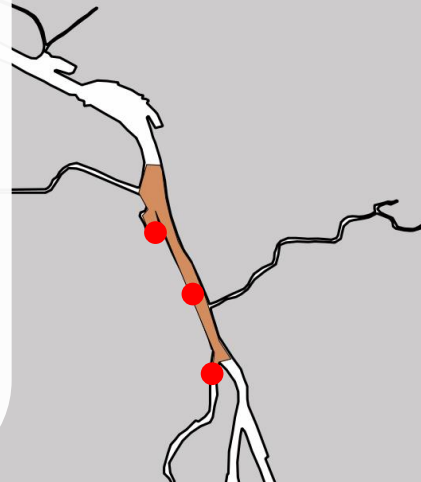
Spring 2017

Summer 2017

Materials and methods: The sampling design

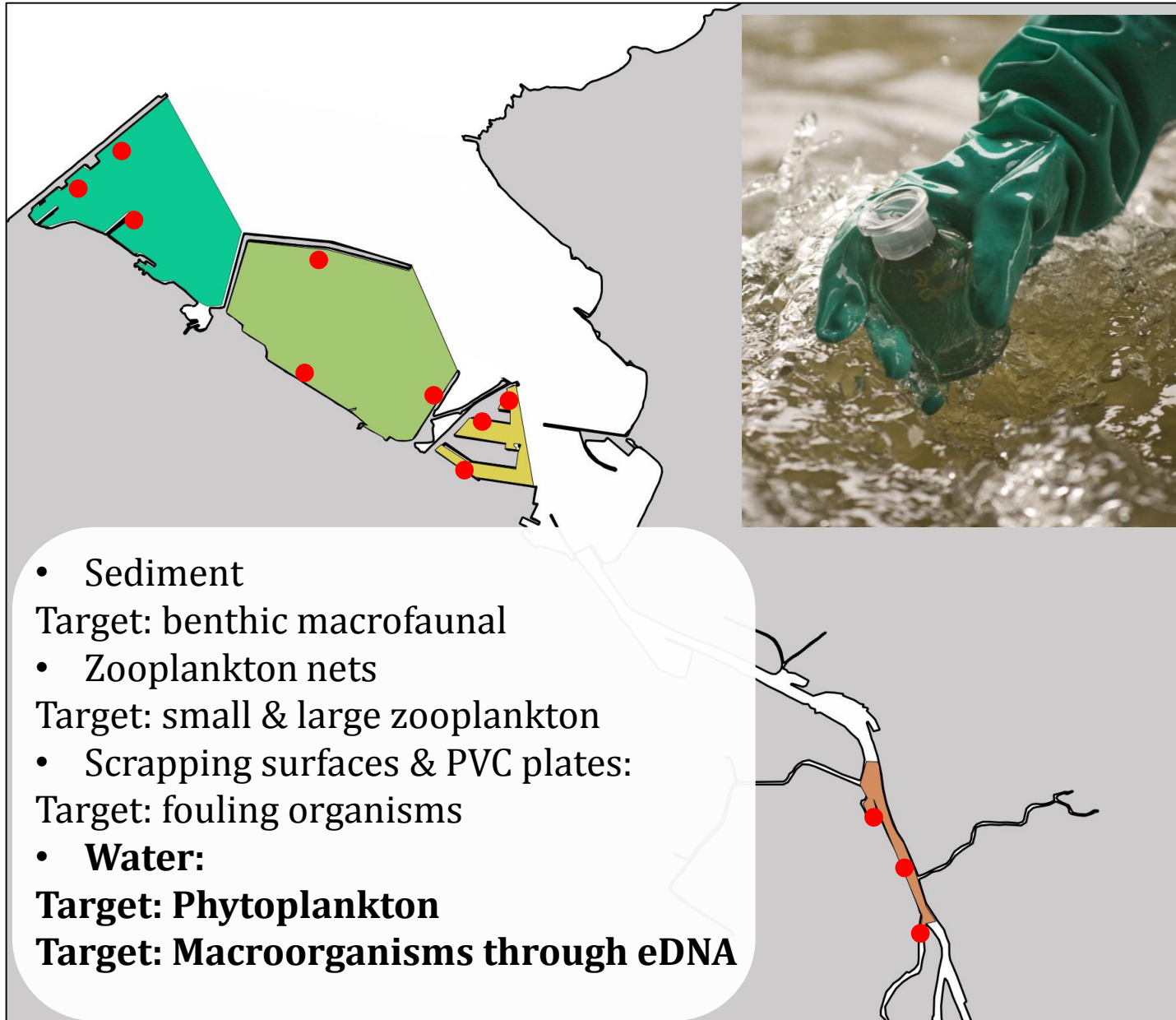


- Sediment
Target: benthic macrofaunal
- Zooplankton nets
Target: small & large zooplankton
- **Scrapping surfaces & PVC plates:**
Target: fouling organisms



2 Sampling periods:
Winter to Spring 2017,
Spring to Summer 2017

Materials and methods: The sampling design



4 Sampling periods:

Autumn 2016

Winter 2017,

Spring 2017

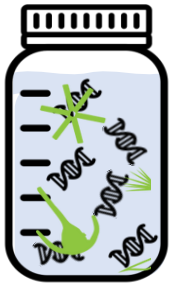
Summer 2017

Materials and methods: The metabarcoding workflow

- In total, 160 samples were processed
- From filtered water:

To retrieve macroorganisms

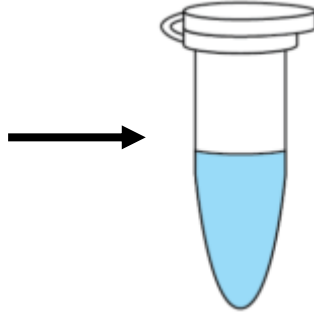
Environmental sample



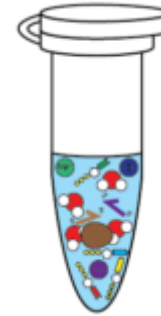
Filtration



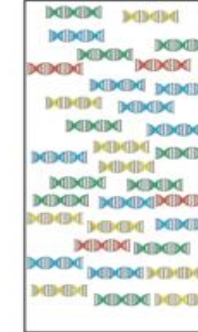
DNA extraction



Amplification of
DNA barcode:
COI



Next-Generation
Sequencing:
Illumina MiSeq



Bioinformatic
processing

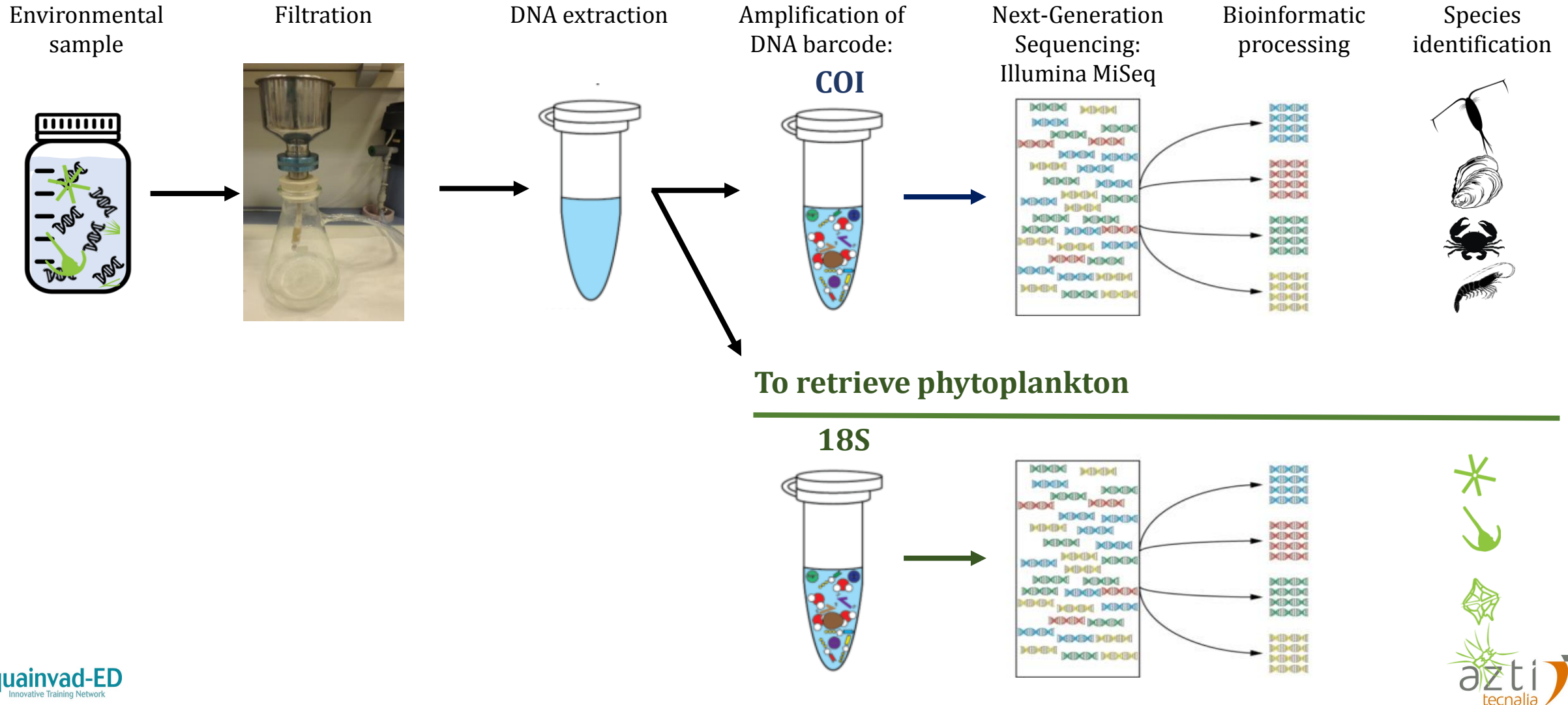


Species
identification



Materials and methods: The metabarcoding workflow

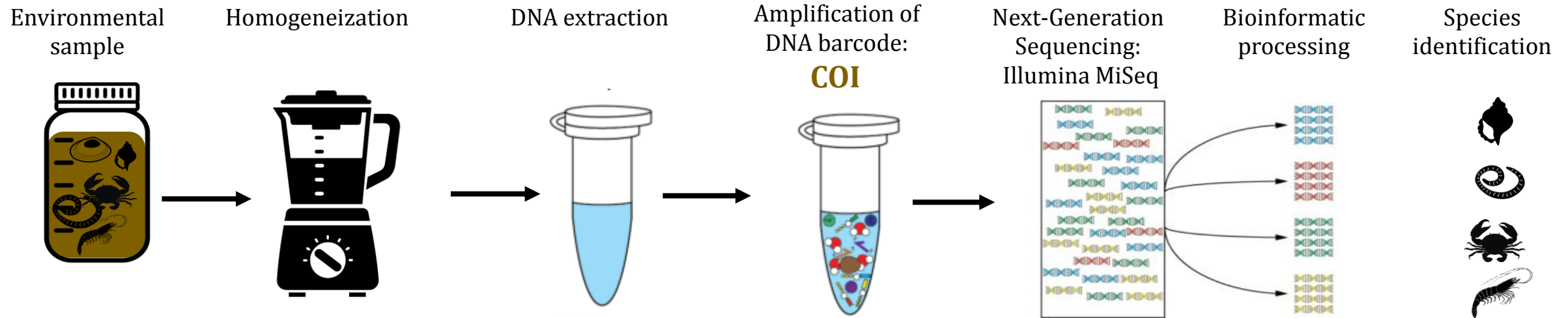
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- From sediment, PVC plates and plankton nets → Similarly processed

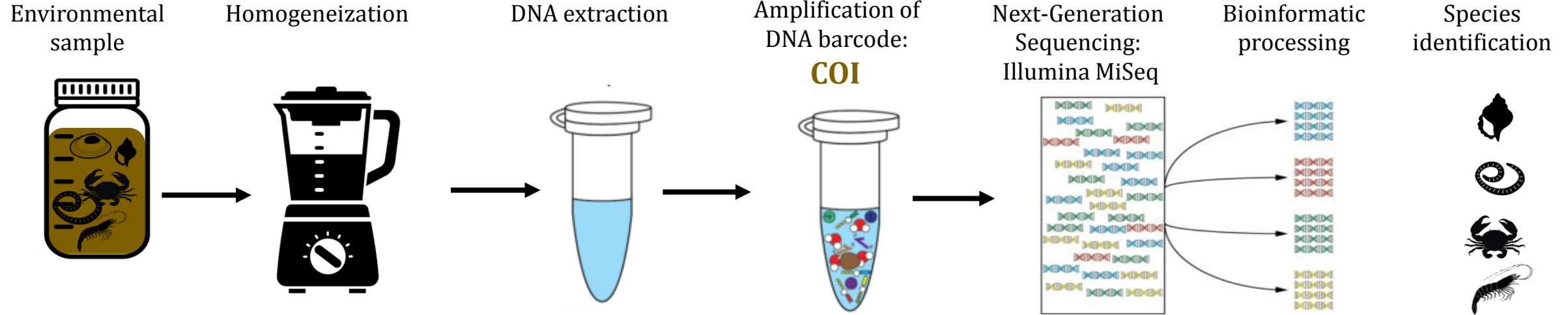
To retrieve benthic macrofaunal



Materials and methods: The metabarcoding workflow

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To retrieve benthic macrofaunal

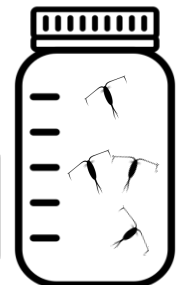


To retrieve benthic macrofaunal:
COI
To retrieve harmful algae species:
18S

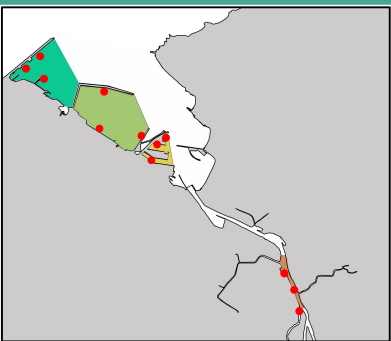


To retrieve fouling organisms:
COI & 18S

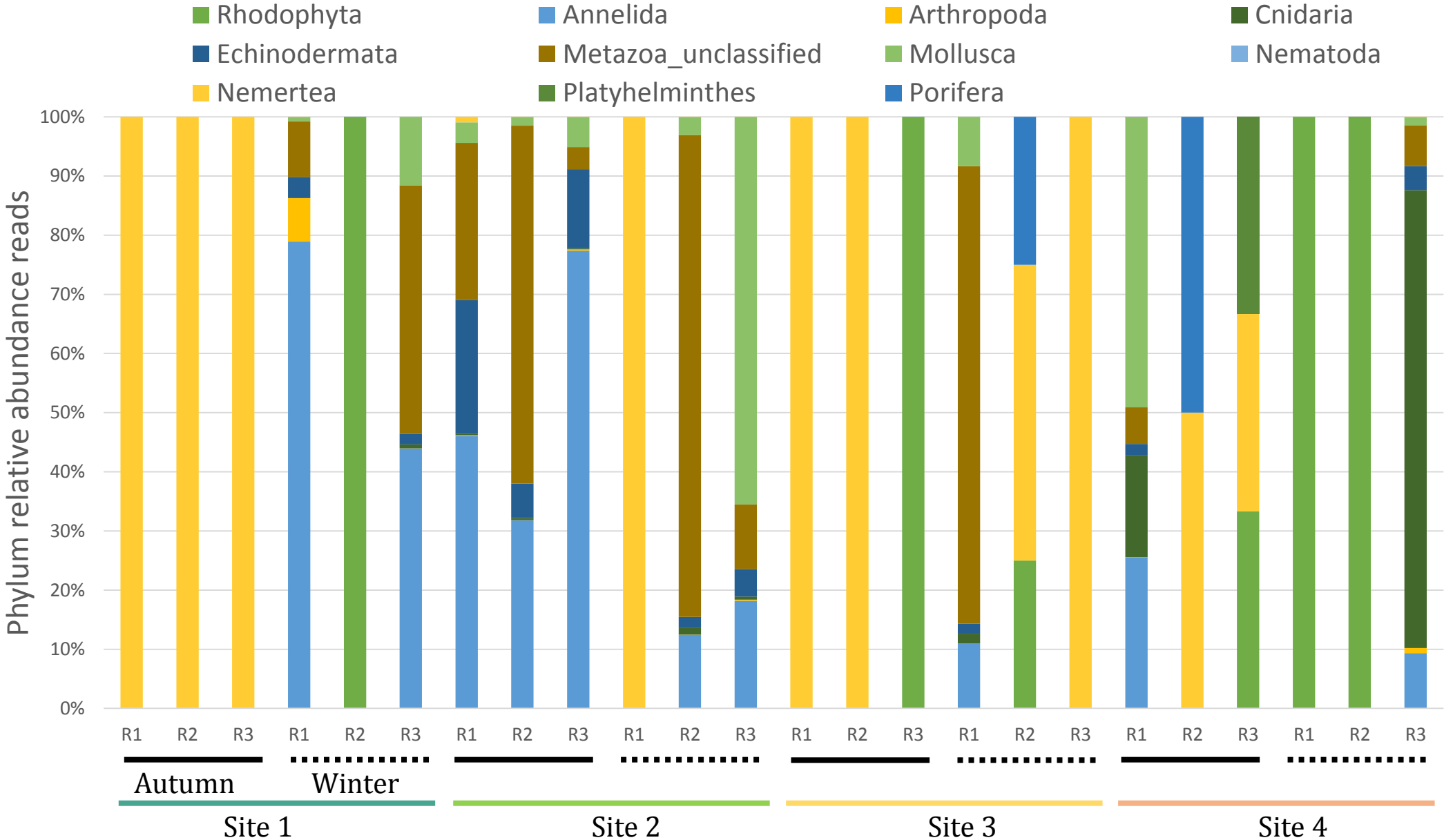
To retrieve zooplankton:
COI and 18S



Preliminary results for the benthic macrofauna sampling:



Autumn and Winter periods :



Preliminary results for the benthic macrofauna sampling:

Total OTU α -Diversity : 104

- **Autumn period : 88 (84%)**
- **Winter period : 78 (75%)**
- **Shared OTU between 2 periods : 62 (60%)**

OTU identified to :

- **Species level \rightarrow 73 (70 %)**



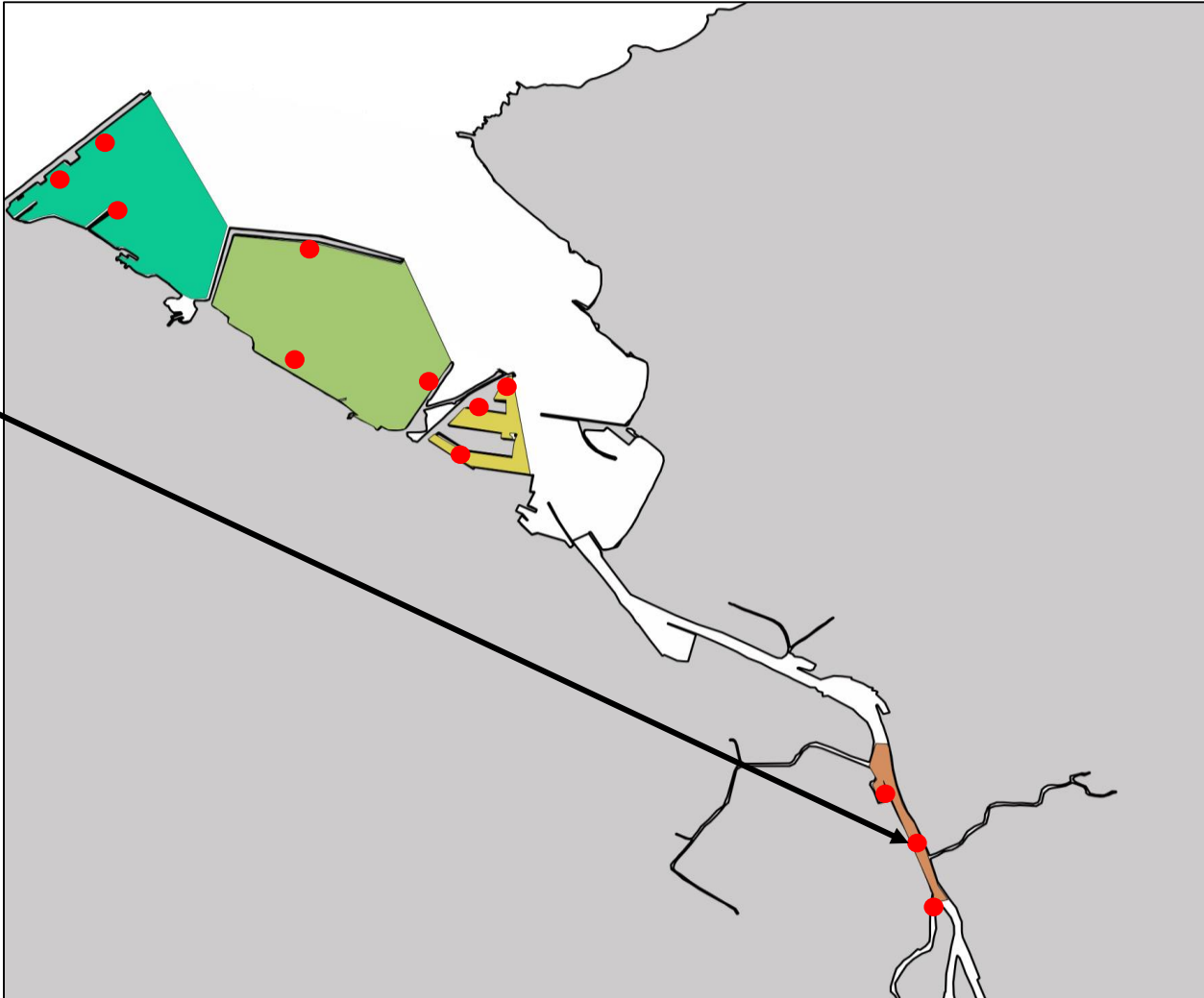
Preliminary results for the benthic macrofauna sampling:

Non-Indigenous Species detected in the benthic macrofauna communities:



Black-pygmy mussel
(*Xenostrobus securis*)

- First report in 2010
- Native from Japan
- Likely maritime transport

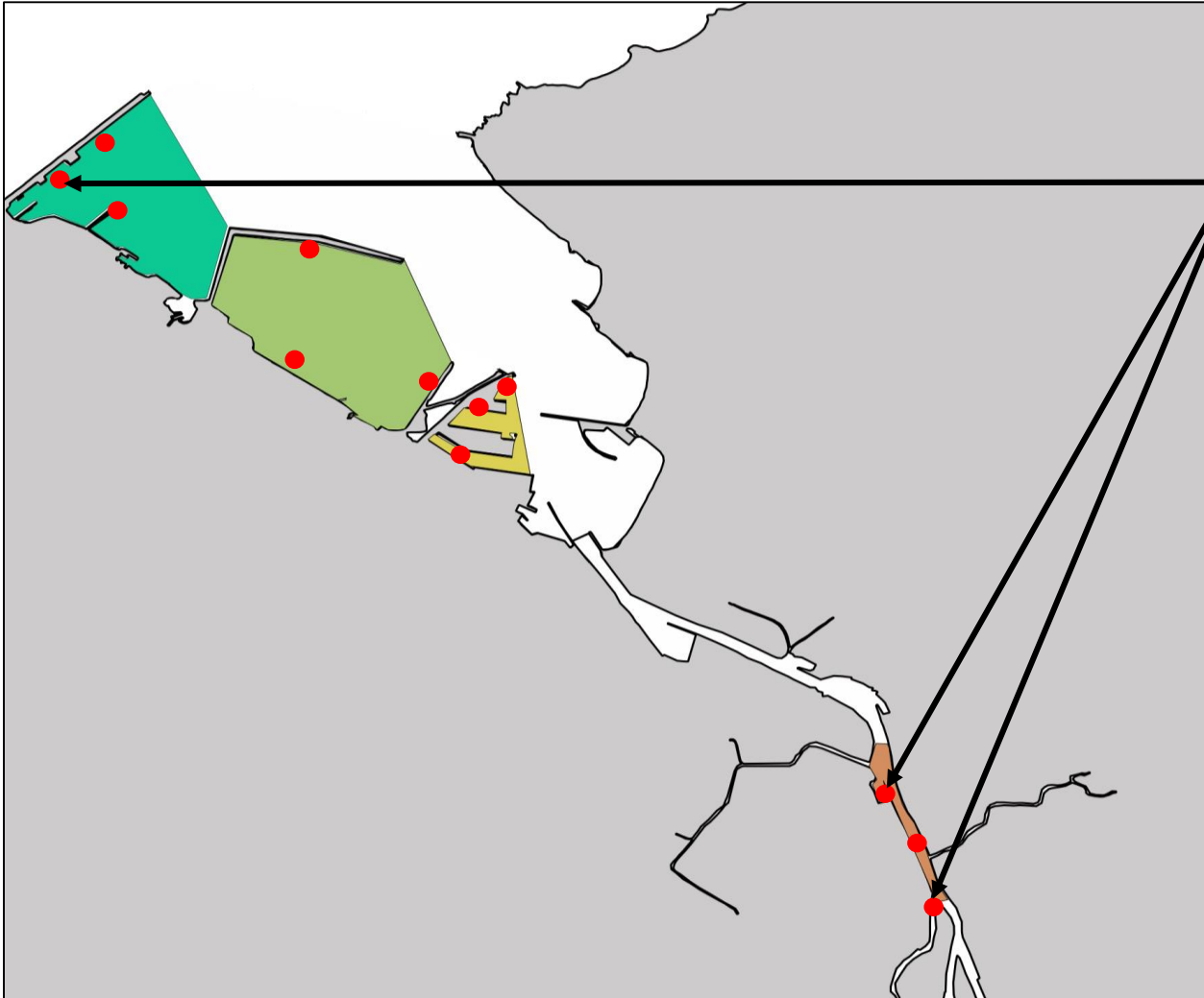


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Manila clam (*Ruditapes philippinarum*)

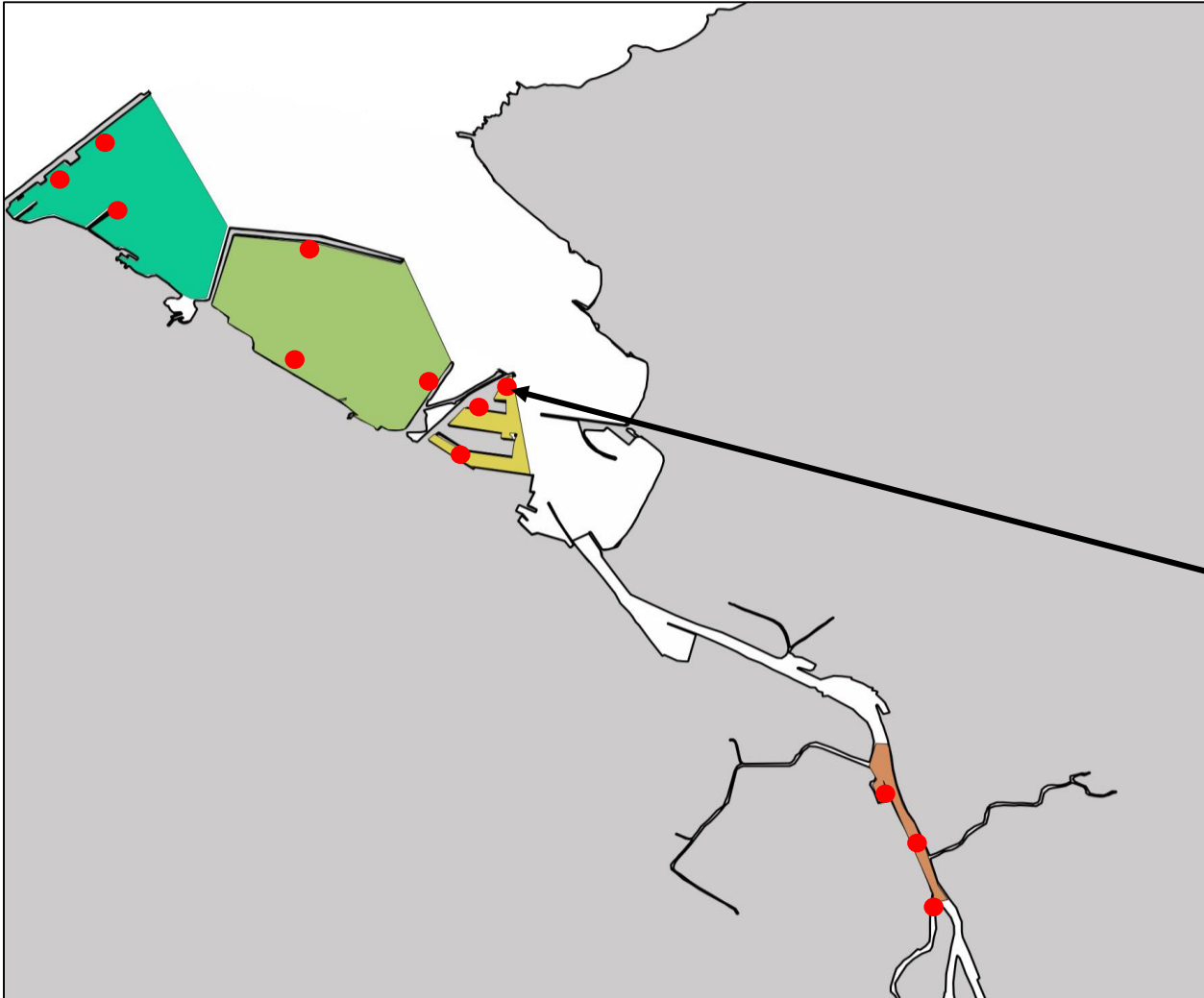
- First report 2005
- Native from Pacific coast of Asia
- Introduced with aquaculture in Europe

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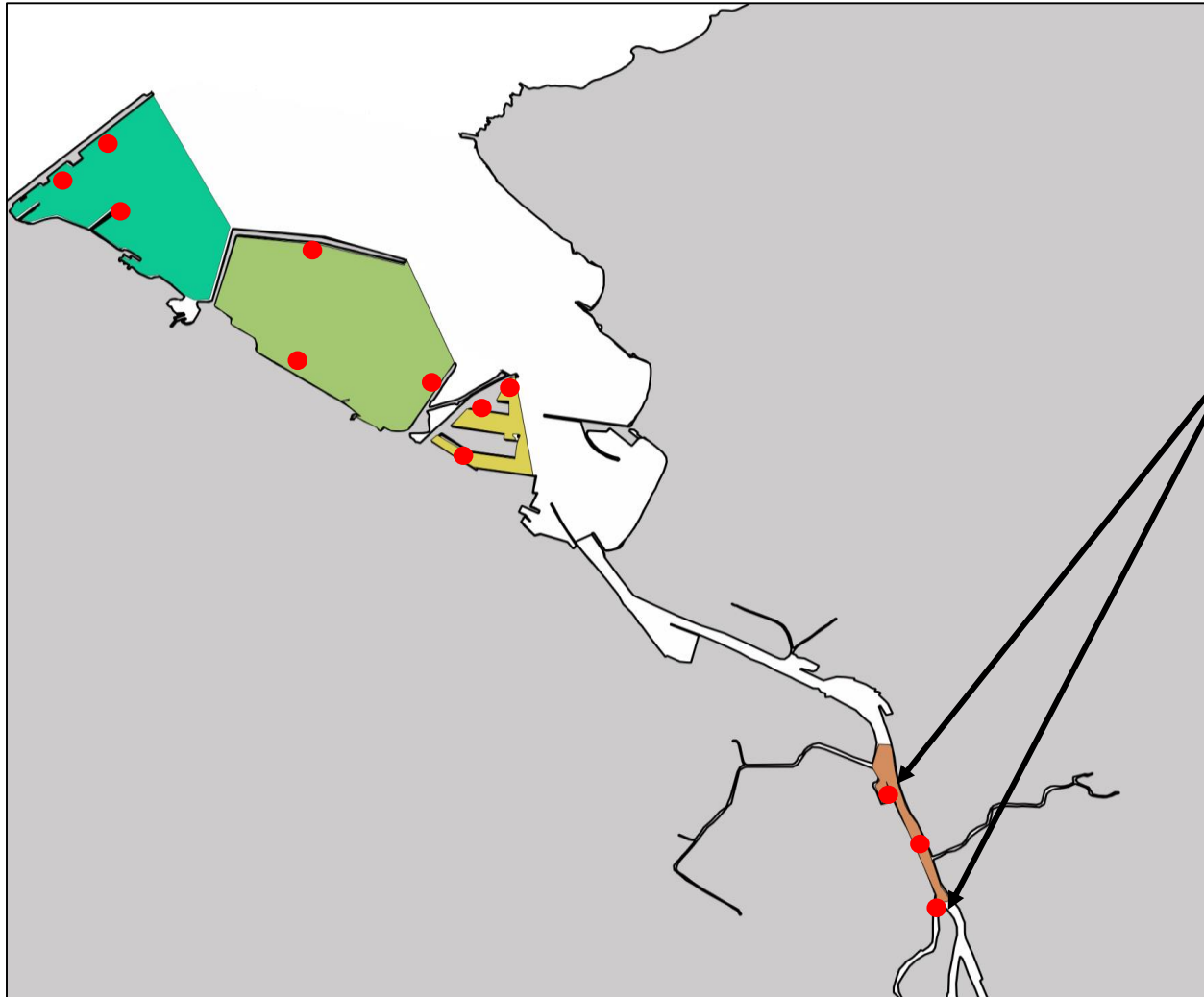


Amphipoda (*Grandidierella japonica*)

- **FIRST RECORD in Northern Spain**
- Native from Japan
- Likely maritime transport

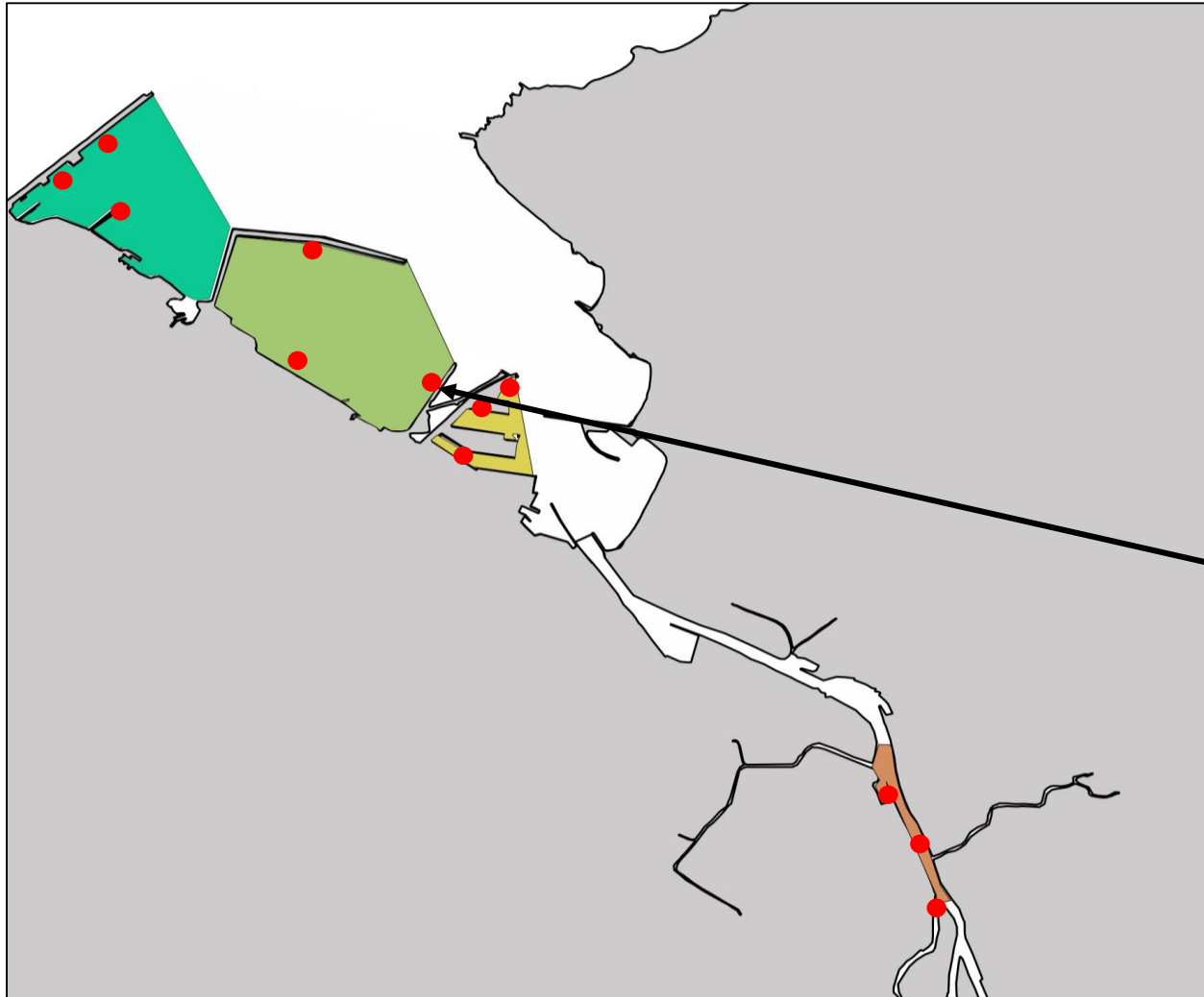
Preliminary results for the benthic macrofauna sampling:

Non-benthic macrofauna Non-Indigenous species recovered with the sampling:



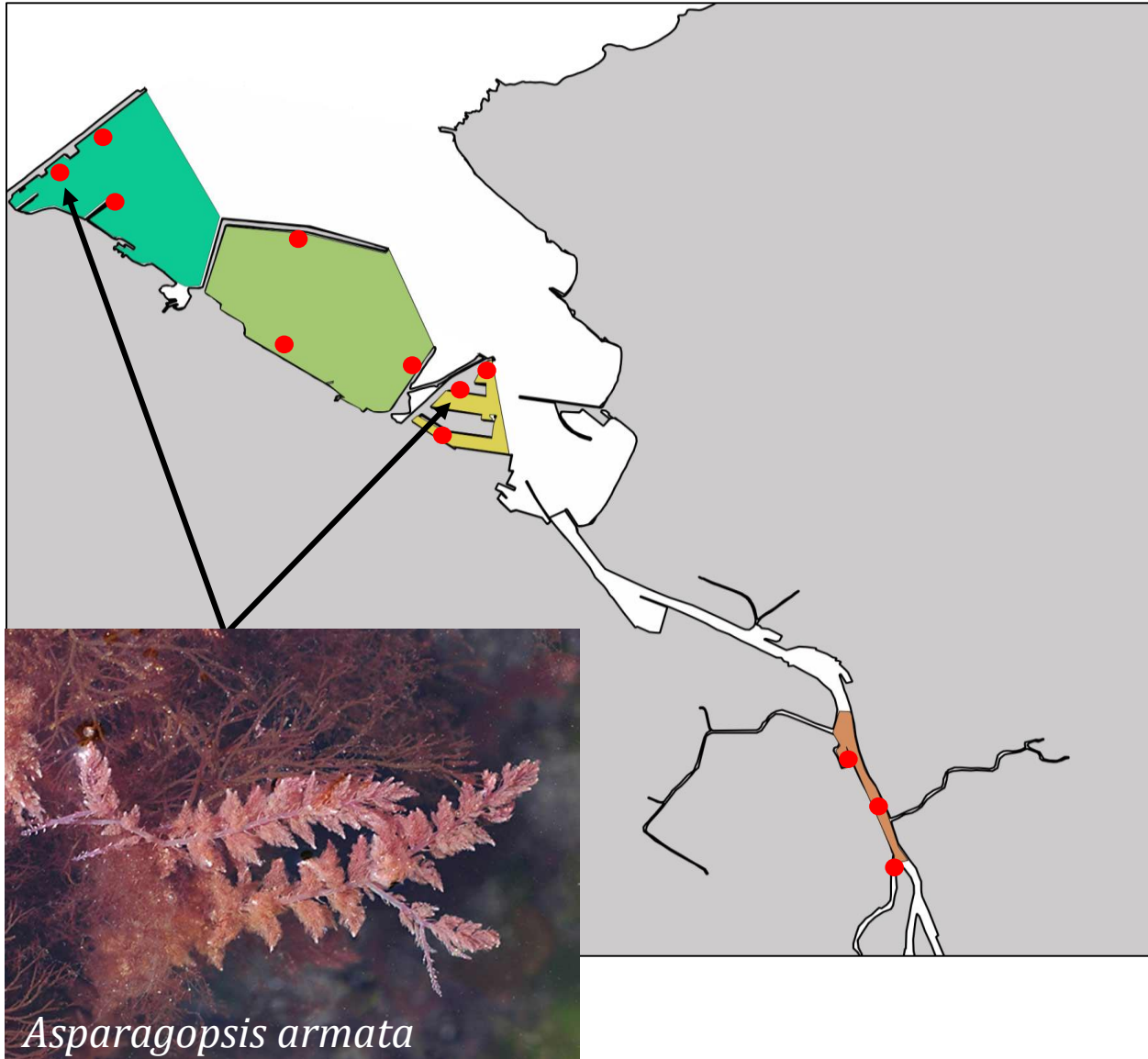
Preliminary results for the benthic macrofauna sampling:

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DISCUSSION & CONCLUSION:

What have we learned so far?



- First results suggest DNA metabarcoding as cost-effective tool to provide biological data for port baseline survey
- Good species level recovery in general
- Seasonal sampling is needed
- Some samples are dominated by one phylum → Representativity?

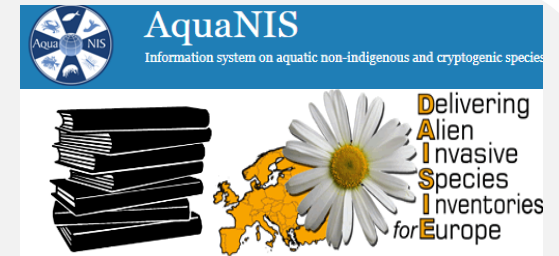
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What is next?

- Work in progress 
- Comparison of the DNA metabarcoding port baseline survey with previous monitoring surveys and Non Indigenous Species databases
- Assessment of environmental DNA (from water and sediment) to retrieve the community of the biodiversity components
- Cost and time analysis 
- Propose the next steps to optimize:
 - Barcoding of Non-Indigenous Species?
 - Further benchmarking of metabarcoding protocols to minimize overlooking taxa?





Thank you for your attention!

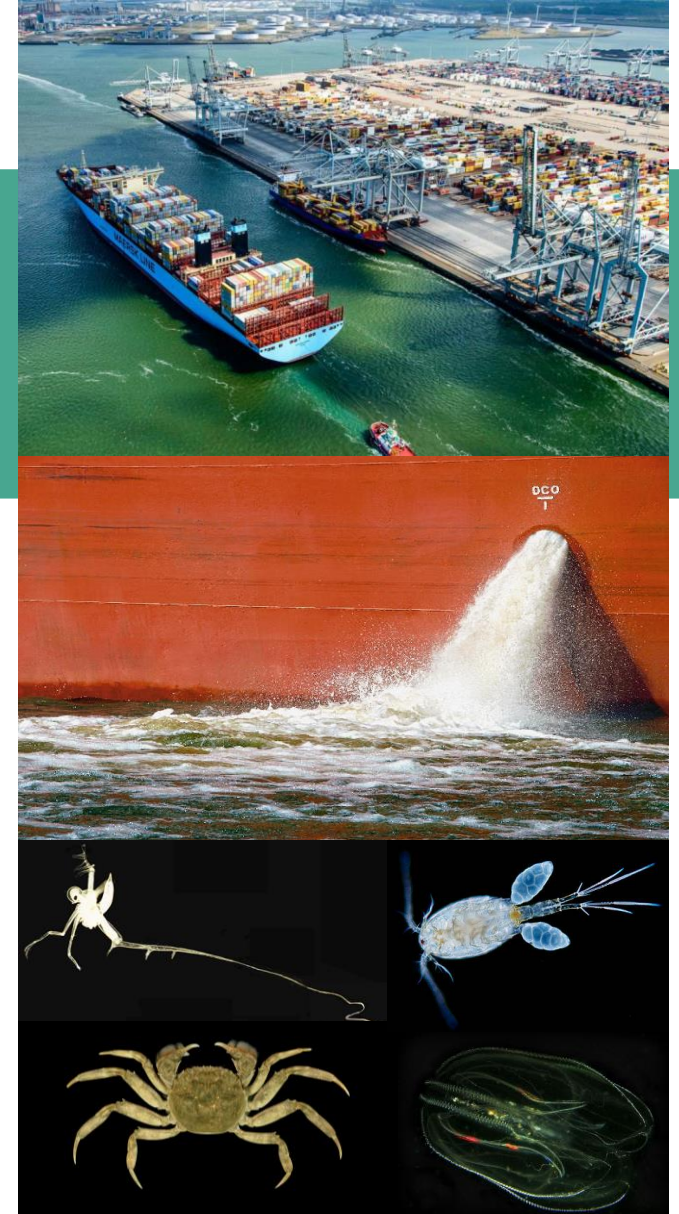
Questions?

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Dr Oihane C. Basurko: ocabezas@azti.es

<http://www.aquainvad-ed.com/>



Introduction: Making the link between ballast water monitoring & genetic tools

Methodology: Review of the literature, guidelines and regulations of the BWM Convention



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The challenges and promises of genetic approaches for ballast water management

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Overview

- **36 studies in ballast water and ports which targeted:**
Pathogens, Bacteria, Protists
Zooplankton, Phytoplankton
Invertebrates in sediment
- **Approaches:**
Targeted species and biodiversity assessment surveillance



Monitoring applications extracted from Guidelines

- Test the efficacy of ballast water management
- Assess the alive biodiversity discharged via ballast water
- Risk assessment for granting exemptions: port baseline survey & target species detection



Future investments before possible implementation

- Standardization
- Increasing portability of genetic tools
- Improvement of reference databases
- Exploring alternative sampling strategies (environmental DNA)