



Methods for Quantifying Biofouling: An Initial Examination of Optical and Acoustic Approaches

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Biofouling Background

- Potential transport of aquatic nuisance species
- Major concern for vessels with operating profiles with long periods of inactivity*
- Currently, international guidelines have been promulgated for the reduction and prevention of biofouling



<https://www.green4sea.com/amsa-revises-biofouling-and-in-water-cleaning-guidelines/>



*Hopkins and Forrest, 2010

Fouling Mitigation—Coatings

Before 1960

- Toxic biocides (for example arsenic, mercury, and DDT) used in coatings to kill attached organisms

After 1960

- Tributyltin (TBT)
 - Toxic and persistent in environment
 - Worldwide ban on application since 2003
- International Convention on the Control of Harmful Anti-fouling Systems on Ships (International Maritime Organization, 2001)
- Currently, coatings with other antifoulants are used (e.g., copper)
- Other approaches being used (e.g., fouling-release coatings)



Biofouling Organisms

Microorganisms

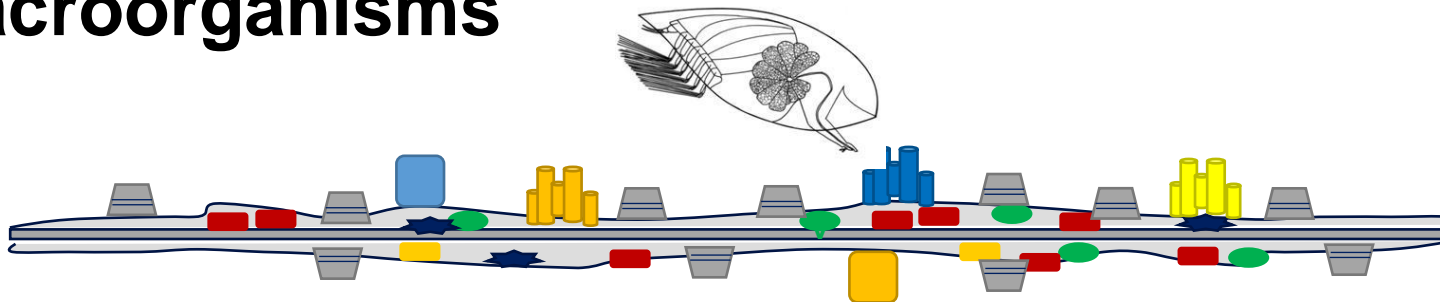


**Surface
immersed
in seawater**

Biofilm formation:

- Microorganisms (e.g., bacteria and microalgae) adhere to submerged surfaces and produce extracellular polymeric substances (EPS)
- Other organisms (protists, fungi, microinvertebrates) accumulate in the biofilm
- The thickness is on the scale of micrometers to millimeters

Macroorganisms



- Examples of hard foulers: Larval barnacles settle as cyprids – undergoes metamorphosis to adult barnacle, tube worms
- Examples of soft foulers: Macroalgae, tunicates, sponges

Goal of Work

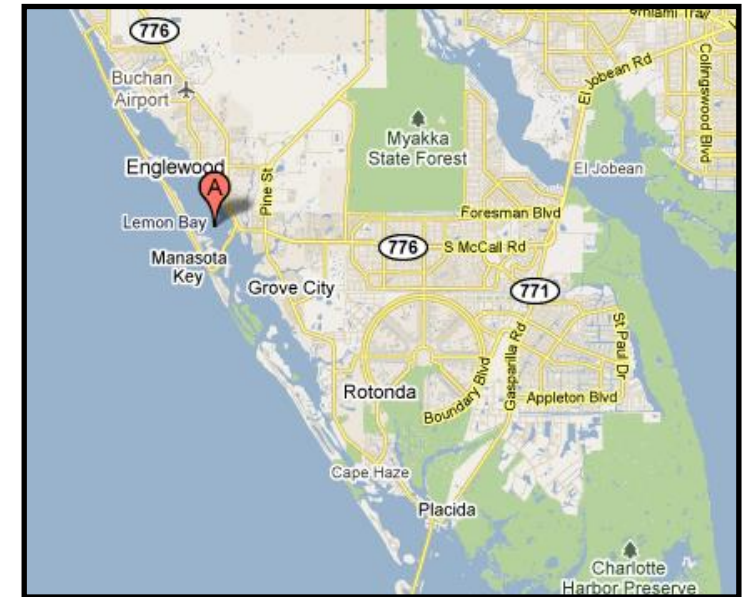
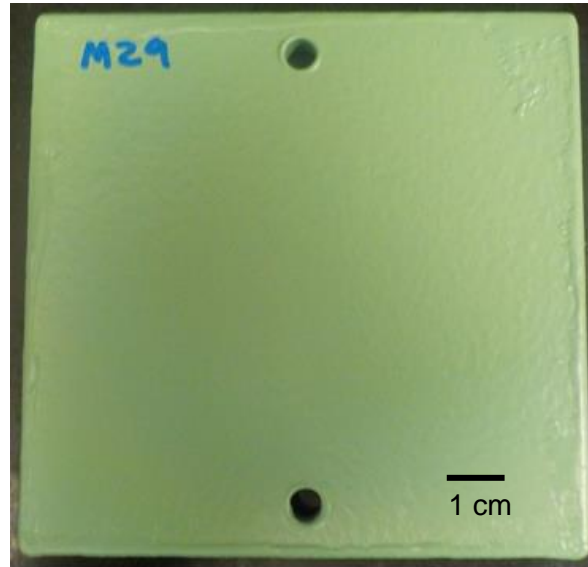
- The purpose of this pilot study was to evaluate optical and acoustic approaches to quantifying biofouling
- Methods Tested:
 - Imaging Fluorometry
 - Acoustic Imaging (with single and dual beam sonars)
 - Optical Imaging (using an underwater, digital camera)

Note: Methods not primarily designed for biofouling quantification

Methods—Test Panels and Exposure Sites

Test panels provide substrate for biofouling growth

Steel test panels
with inert epoxy
coating
(10 x 10 x 0.5
cm)



Methods—Imaging Fluorometry

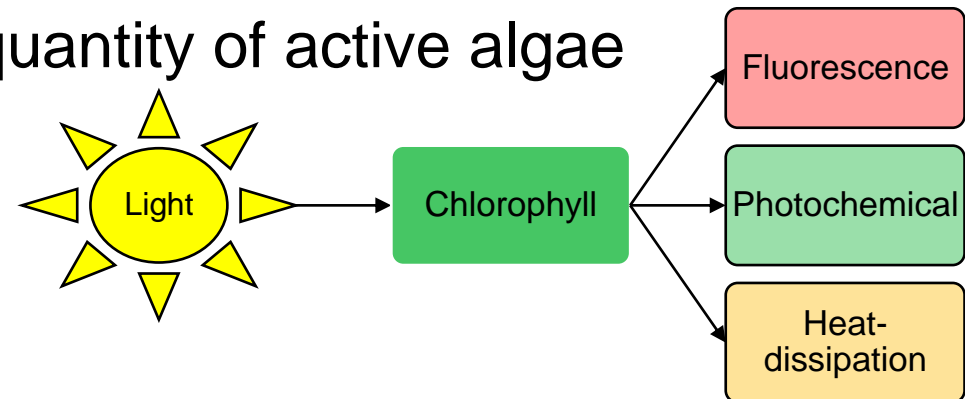
• Variable fluorescence fluorometry

• Saturation Pulse Method

- Basic assessment of photosynthetic performance of a sample
 - Photochemical yield (F_V/F_M)
- Can provide information on total quantity of active algae

• Imaging-PAM (MAXI version; WALZ, Germany)

- Maps fluorescence signals along surface using digital camera
- Collects fluorescence characteristics defined by pixels
- Generates high-resolution 2D map (algae biofilms)



http://www.walz.com/products/chl_p700/imaging-pam_ms/introduction.html

Methods—Acoustic Imaging

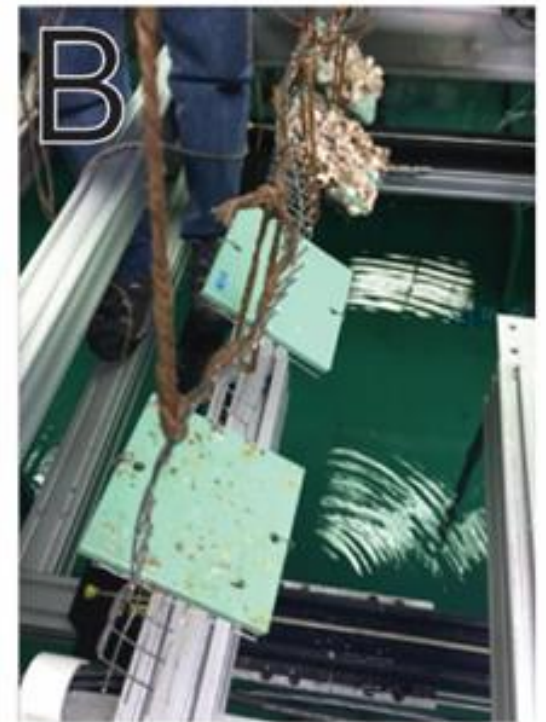
- 2D Imaging Sonar - Single Beam
 - M450 2D Imaging Sonar; BlueView; Bothell, WA
 - Single frequency – 450 kHz

Single beam sonar approach:

A. Test panels held in wire mesh



B. Lowered into tank



Methods—Acoustic Imaging - Continued

Single beam sonar approach:

C. Suspended panels (view through tank)

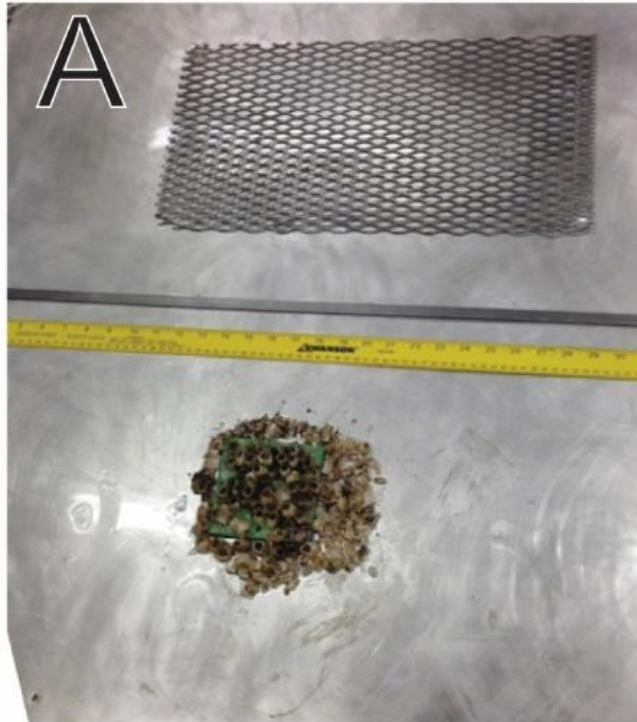
D. Outlined region is enlarged to show the panels



Methods—Acoustic Imaging - Continued

- 2D Imaging Sonar - Dual Beam
 - M900-2250 Imaging Sonar; BlueView, Bothell, WA
 - Mid frequency – 900 kHz; high frequency – 2250 kHz
- Dual beam sonar approach:**

- A. Wire mesh used as reference
- B. Barnacles scraped from one side allowing panel to sit flat; Scraped barnacles were piled on and near top of panel



Panel were submerged in the test tank for acoustic imaging with a dual-beam sonar

Methods—Optical Imaging

Methods

- Waterproof, digital camera used for underwater imaging
- Panels submerged for short (40 d) and long-term (640 d) exposures
- Periodic imaging in flow-through troughs to capture fouling rates
- Fouled panels compared to a non-fouled control surface

Lumix DMC-TS5
(Panasonic North America, Newark, NJ)

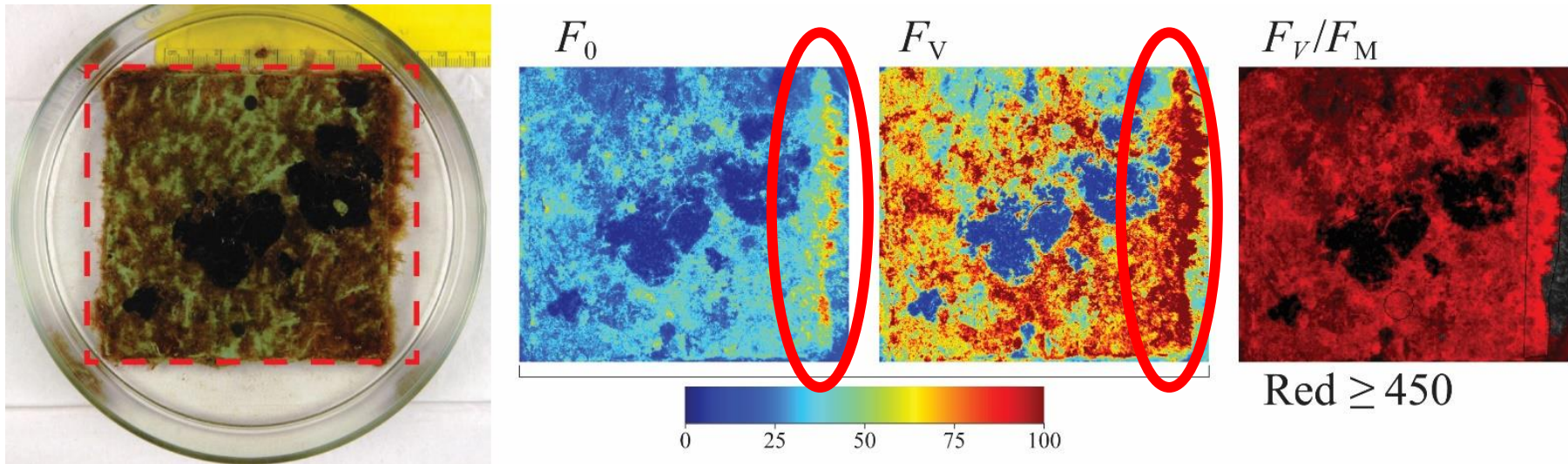


https://www.google.com/search?q=Panasonic+Lumix+DMC-TS5&source=lnms&tbm=isch&sa=X&ved=0ahUKEwiguZCY1u3VAhVF5yYKHdGLAOUQ_AUIDCgD&biw=2100&bih=1132#imgrc=k8ltZ4GuStv02M



Flow-through troughs used to hold panels during imaging in Key West, FL

Results—Imaging Fluorometry

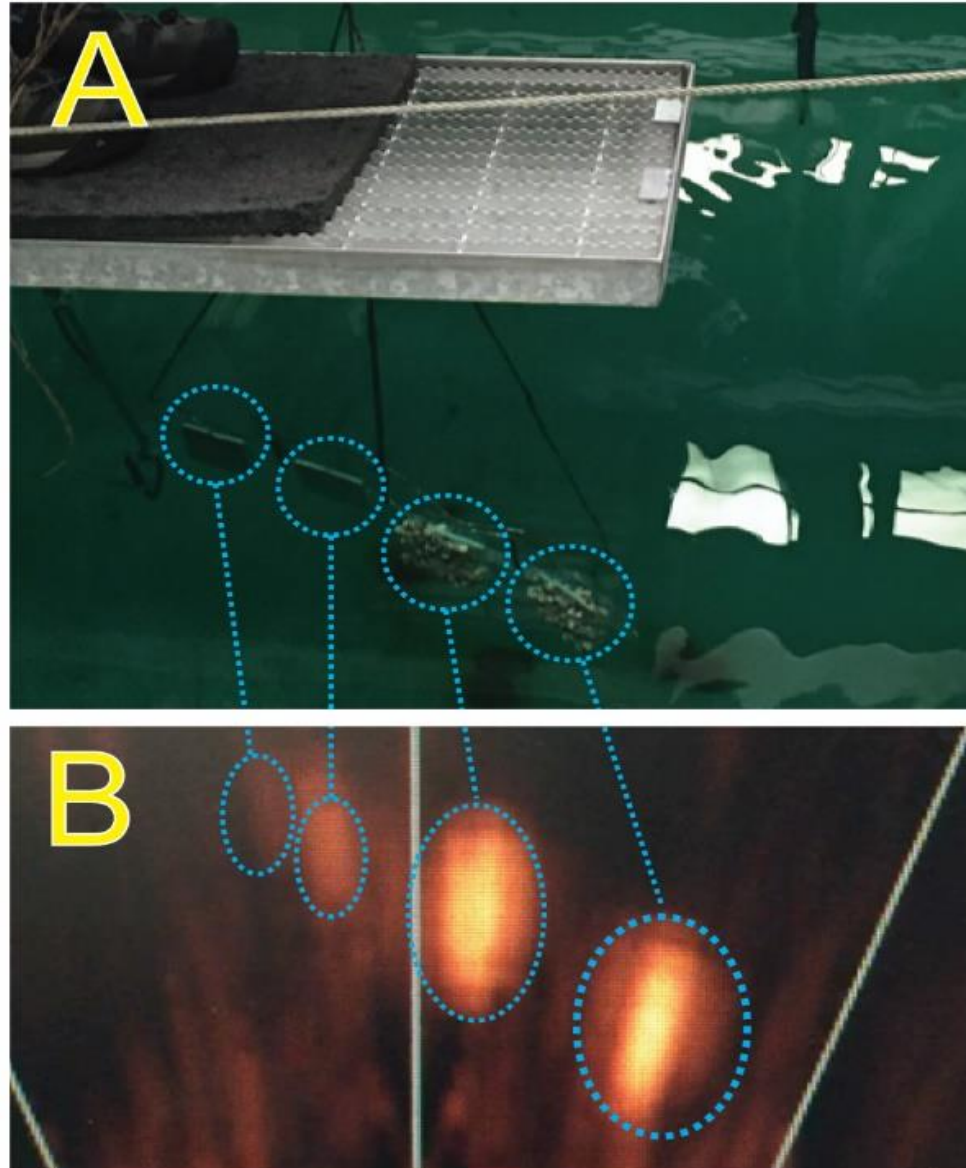


- Visible distribution of algal films throughout panel
- Dark region in center of panel (colonial tunicate) displayed low fluorescence values
- Filamentous algae on right edge of test panel displayed high relative fluorescence intensities
 - Pixels with $F_V/F_M > 450$ (no units) were labeled red

Results—Acoustic Imaging

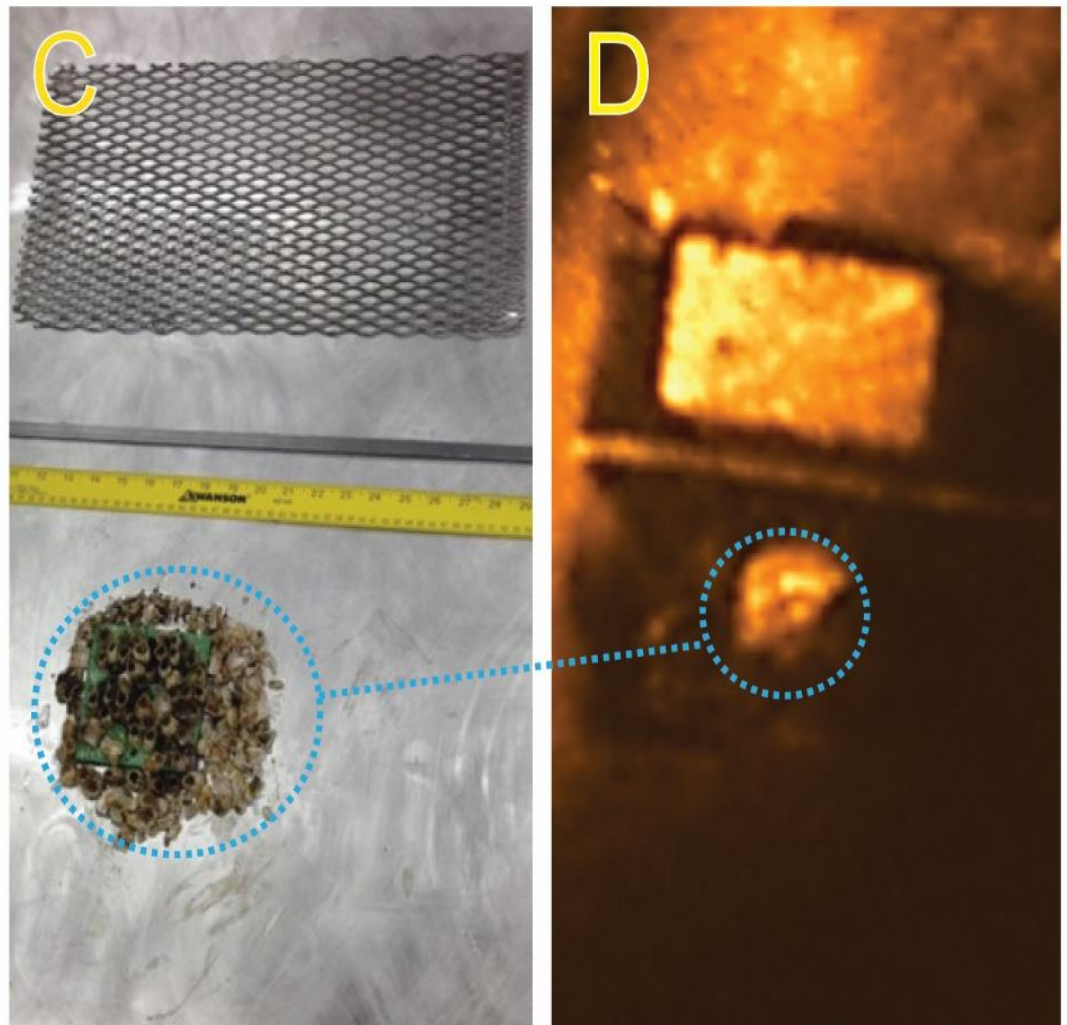
A – B.) Single-beam acoustic imaging (low frequency) could distinguish among panels based on signal intensity

The resolution was not high enough to distinguish difference within a panel

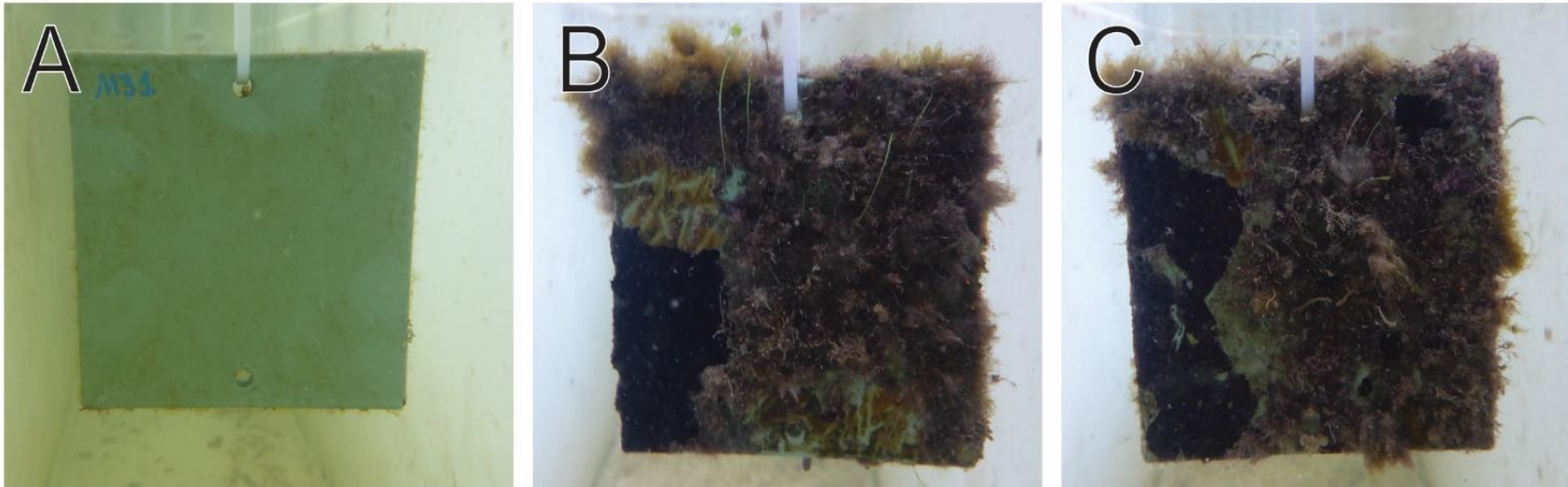


Results—Acoustic Imaging - Continued

C- D.) The dual-beam acoustic imaging (high-frequency) showed higher resolution, and regions within the panel could be distinguished based upon the signal intensity



Results—Optical Imaging



- High resolution images of both algal biofilms and mixed assemblages were collected (A: 40 d incubation; B-C: 640 d incubation)
- Differences between images reflects the affect of water quality on imaging
- Work is currently underway with colleagues from Old Dominion University (ODU) to develop image processing techniques to categorize general taxa and measure surface roughness based on pixel values

Conclusions

- Imaging fluorometry requires test panels to be evaluated *ex situ*; targets only phototrophic organisms
- Technologies may be suited for different tasks
 - Imaging fluorometry measures photochemical yield; could be used to monitor the efficacy of surface treatments or it could be combined with other approaches used to interrogate surfaces
- Single-beam acoustic imaging could distinguish difference among panels (minimal – heavy fouling), but could not distinguish difference within a panel – not enough resolution

Conclusions—Continued

- Regional differences within a panel could be distinguished based on signal intensity due to the higher resolution of the dual-beam acoustic imaging
- Optical images collected under ideal conditions indicate the degree of fouling and allowed for identification of organisms in a mature community (data in prep.)
- Cameras are more available, cheaper, and easier to operate than acoustic-based systems
 - Still require software development to measure quantity of biofouling

Acknowledgements

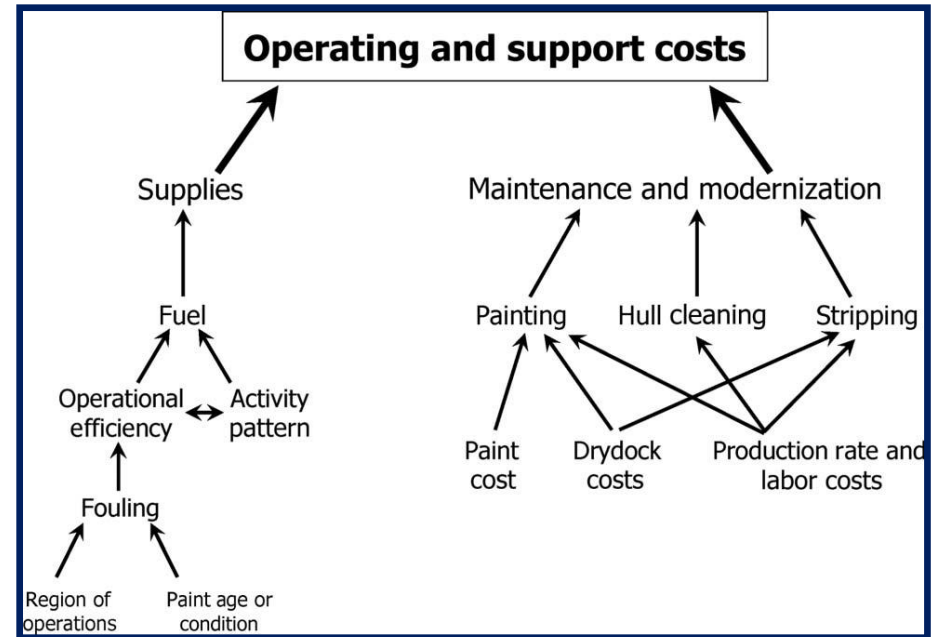
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Background

- Increased frictional drag
→ increased fuel costs
- **Typical fouling levels → ~10% increase in fuel consumption***
- Hull maintenance costs (cleaning, applying new coatings)
- Estimated overall costs of hull fouling: \$56M y⁻¹ for DDG-51 class*

* Schultz et al., 2011



(Schultz et al., 2011)

USS *Arleigh Burke* (DDG-51)



https://en.wikipedia.org/wiki/USS_Arleigh_Burke

Fouling Mitigation—Cleaning

- Dry docking
- Diver cleanings
(‘full’, ‘interim’, or ‘partial’)
- ROV cleanings

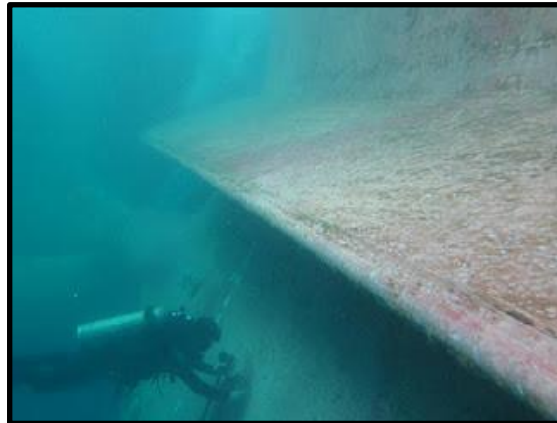
Years

Interval between
cleanings

Days



<https://www.prlog.org/11529528-chemical-tanker-sanmar-majesty-major-drydocking-and-lay-up-repairs-completed-sucessfully.html>



<http://antipodeanmariner.blogspot.com/2011/12/clean-bottom.html>



<http://embient.de/en-index.html>

Issues:

- Cost
- Logistics
- No specified interval for hull cleaning