

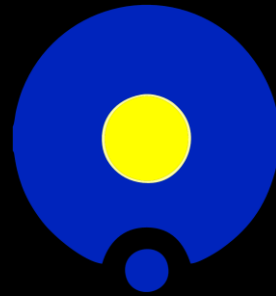
WHAT REALLY SCARES ZEBRA MUSSELS?

A few words about the impact of biotic factors on valve movement reactions of the zebra mussel *Dreissena polymorpha*

Anna Dzierżyńska-Białończyk

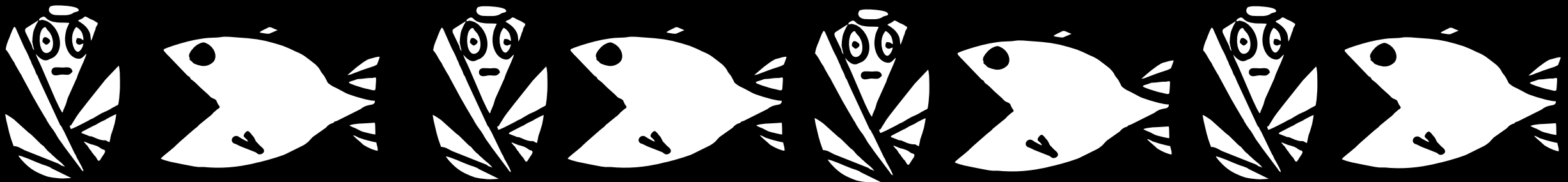
Jarosław Kobak,

Łukasz Jermacz



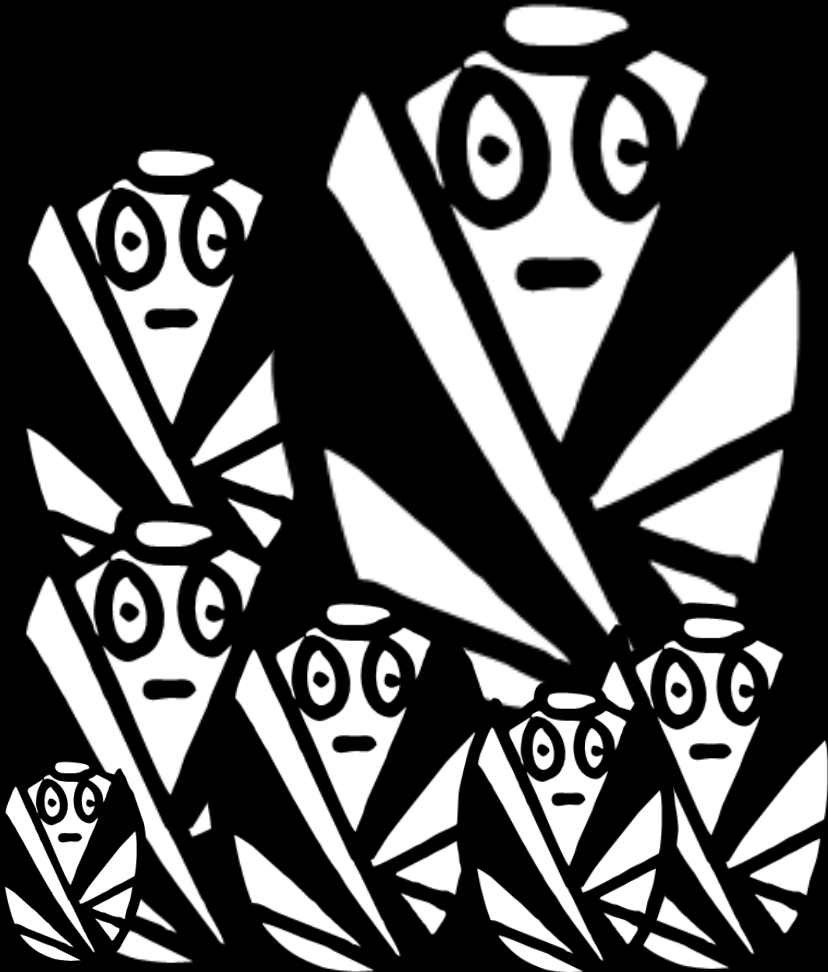
Nicolaus Copernicus University

in Toruń, Poland



ZEBRA MUSSEL *DREISSENA POLYMORPHA*

- ❖ Invasive species from the Ponto-Caspian region
- ❖ Used in early warning systems based on mussel valve gaping



Ecotoxicology 6, 153–165 (1997)

Valve movement response of the mussel *Dreissena polymorpha* – the influence of pH and turbidity on the acute toxicity of pentachlorophenol under laboratory and field conditions

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Pergamon

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TECHNICAL NOTE

SHELL VALVE MOVEMENT RESPONSE OF DARK FALSE MUSSEL, *MYTILOPSIS LEUCOPHAETA*, TO CHLORINATION

SANJEEVI RAJAGOPAL¹*, GERARD VAN DER VELDE¹ and HENK A. JENNER²

ELSEVIER

Arch. Environ. Contam. Toxicol. 40, 497–504 (2001)
DOI: 10.1007/s002440010202

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and Toxicology**
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The Influence of Suspended Particles on the Acute Toxicity of 2-Chloro-4-Nitro-Aniline, Cadmium, and Pentachlorophenol on the Valve Movement Response of the Zebra Mussel (*Dreissena polymorpha*)

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Received: 4 July 2000/Accepted: 14 November 2000

Valve movement responses of *Velesunio angasi* (Bivalvia: Hyriidae) to manganese and uranium: An exception to the free ion activity model

Scott J. Markich^{a,b,*}, Paul L. Brown^a, Ross A. Jeffree^a, Richard P. Lim^b

TOXICOLOGY

Aquatic Toxicology 66 (2004) 333–343

www.elsevier.com/locate/aquatox

Copper detection in the Asiatic clam *Corbicula fluminea*: optimum valve closure response

Damien Tran^{a,b,*}, Elodie Fournier^{a,b}, Gilles Durrieu^b, Jean-Charles Massabau^b

THE AIM OF OUR STUDY

- ❖ To check valve responses of zebra mussels to biotic factors
- ❖ 6 variants with chemical cues:

1 control



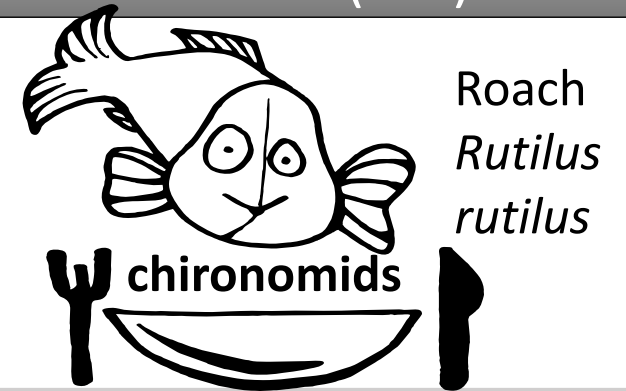
aerated
clean
water

2 with alarm substance

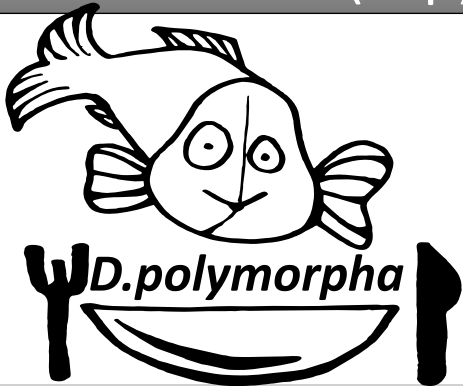


from
crushed
mussels

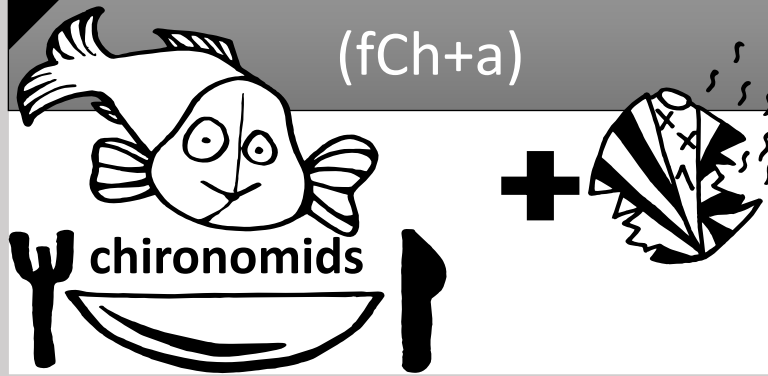
3 with fish scent (fCh)



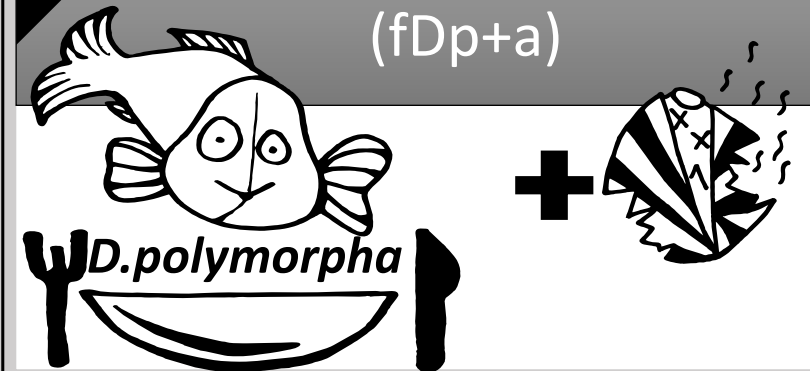
4 with fish scent (fDp)



5 with fish scent and alarm
(fCh+a)



6 with fish scent and alarm
(fDp+a)



THE AIM OF OUR STUDY

- ❖ To check valve responses of zebra mussels to biotic factors
- ❖ 2 variants with the physical presence of:

7 with the presence of
Dikerogammarus villosus



3 individuals
per tank

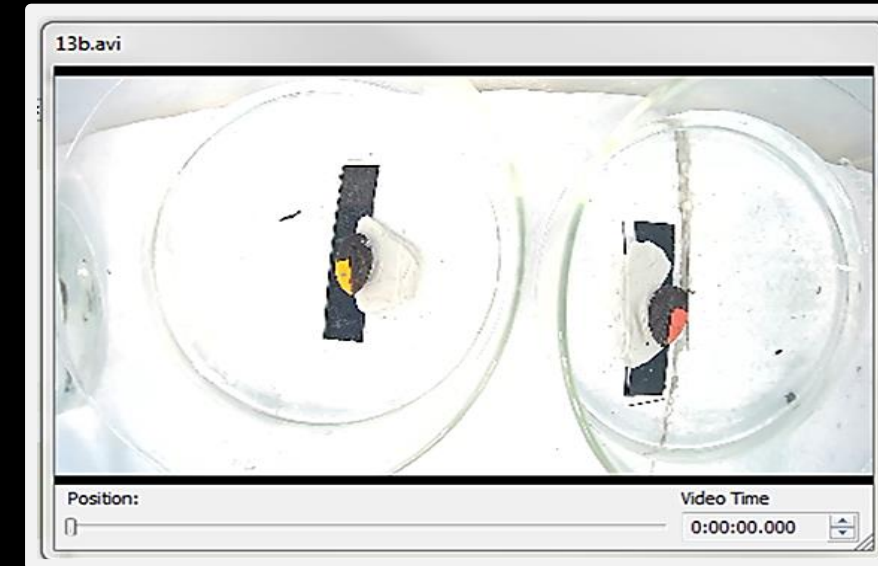
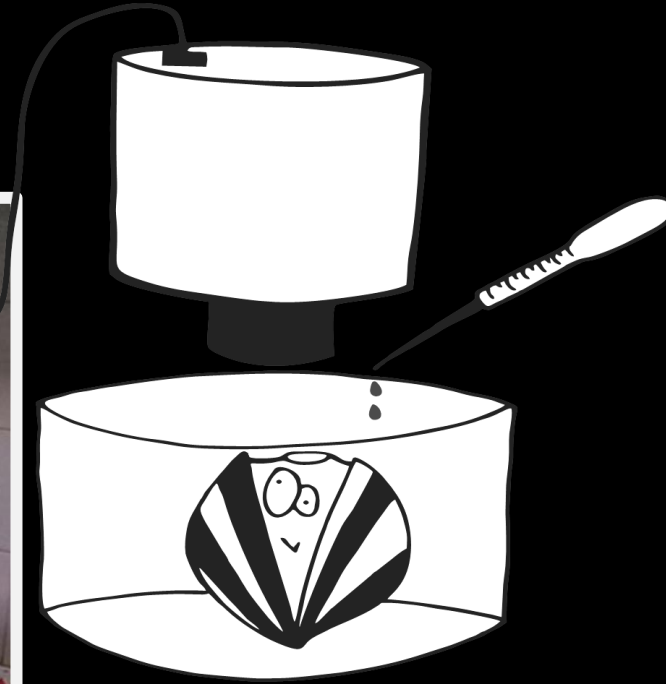
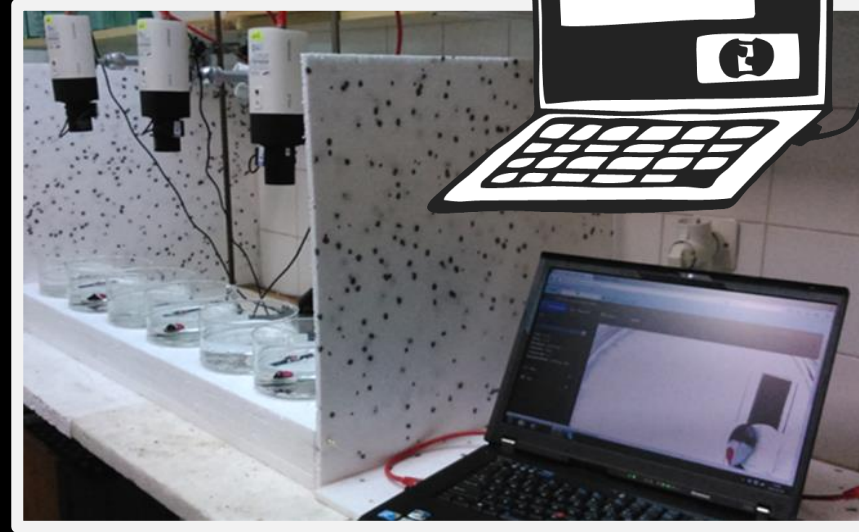
8 control for variant 7



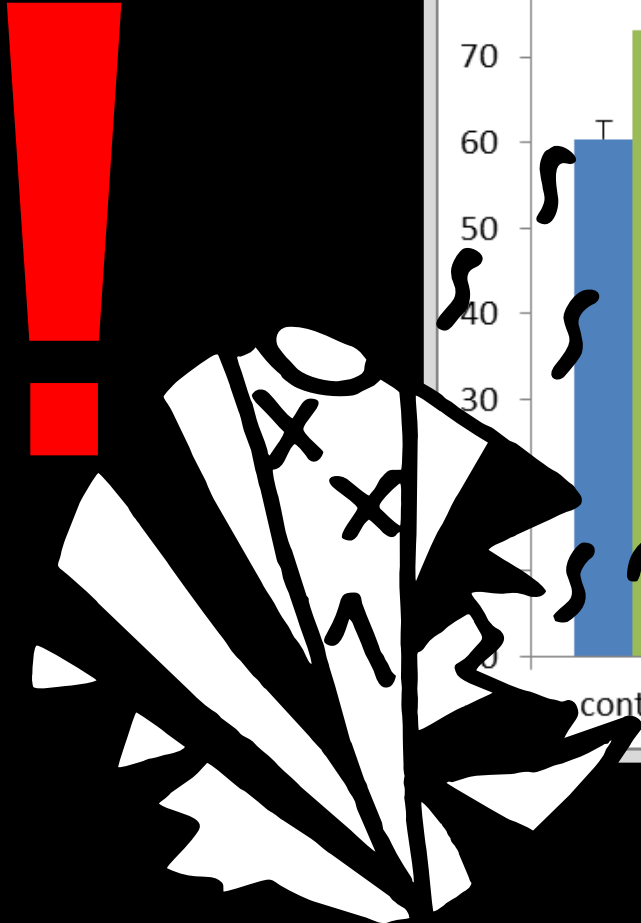
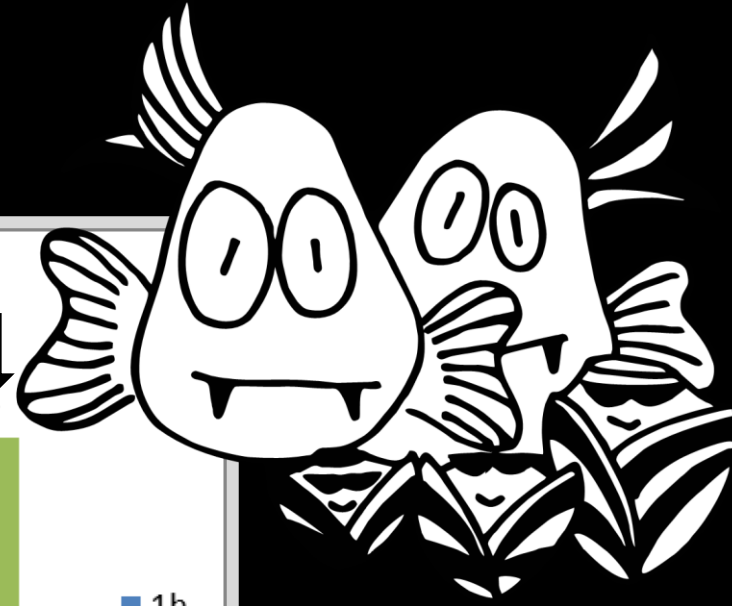
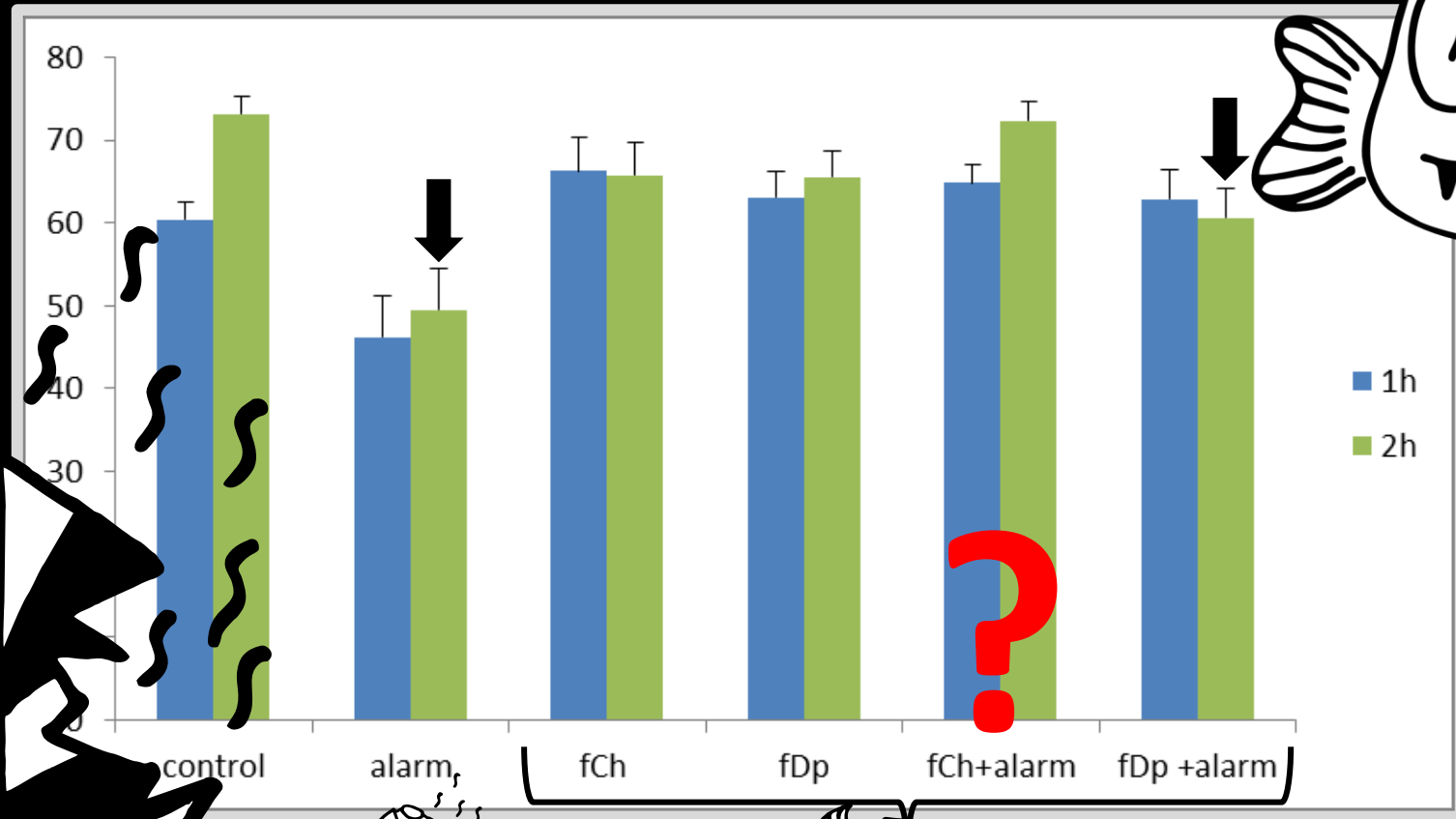
aerated
clean
water

METHODS

- ❖ Experiments under laboratory conditions
- ❖ Recorded by video cameras
- ❖ Tested individuals:
 - one per treatment
 - specially marked
- ❖ Duration: 2 hours
- ❖ Noldus Ethovision XT
 - the software for analysing recorded video files
- ❖ Checked parameters:
 - mean opening (%)
 - total duration of particular valve gaping levels
 - number of valve opening events



RESULTS – MEAN OPENING [%]



Alarm substance = lower mean opening

Kruskal-Wallis test:

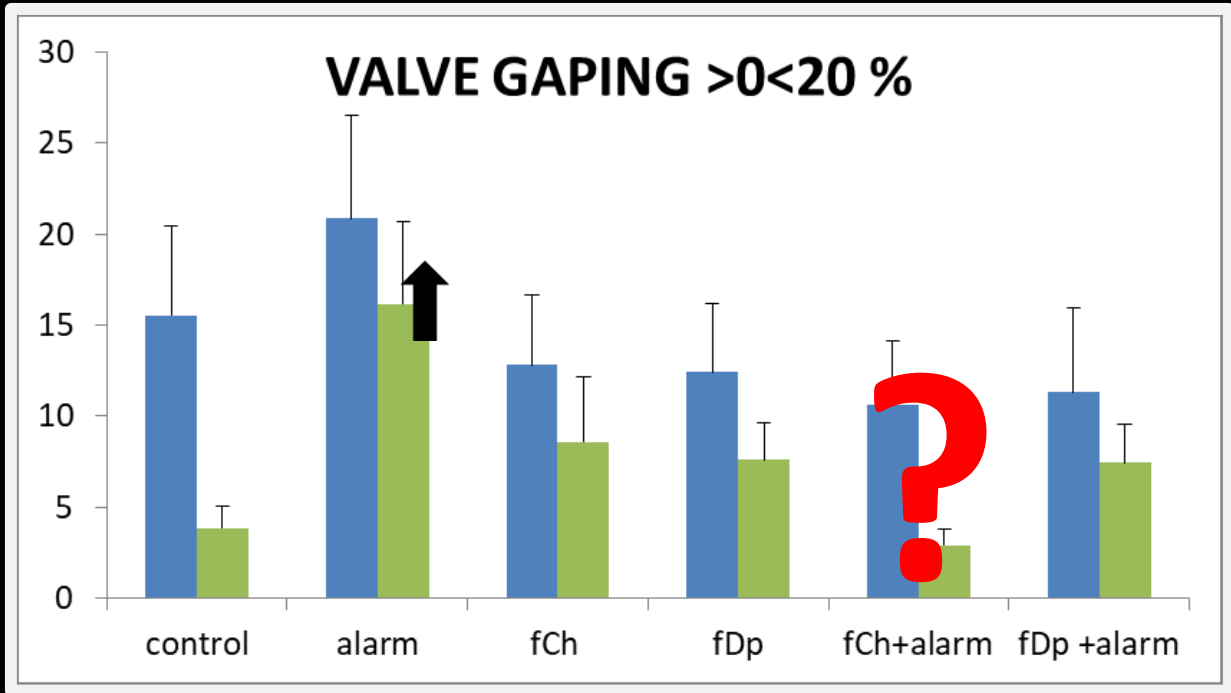
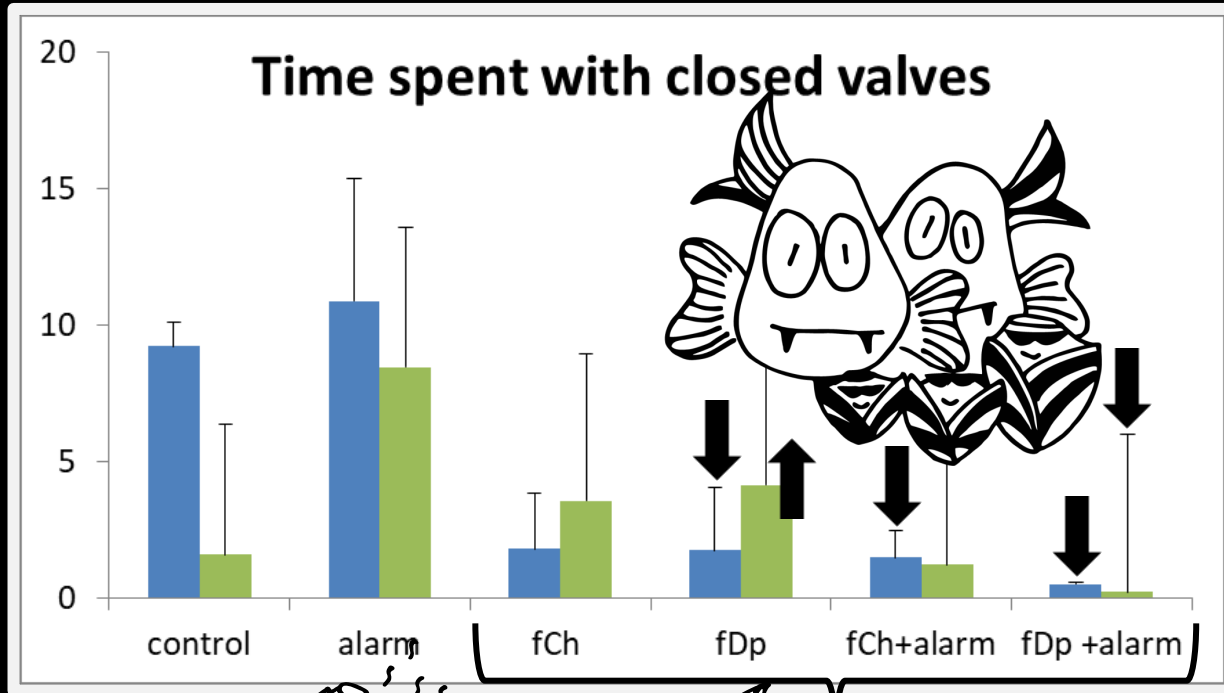
1h: $\chi^2=8.4$, df=5, P=0.135

2h: $\chi^2=21.9$, df=5, P=0.001

RESULTS – DURATION OF PARTICULAR GAPING LEVELS

(% OF TOTAL TIME)

1h 2h



Kruskal-Wallis test:

1h: $\chi^2=37.7$, df=5, $P<0.001$

2h: $\chi^2=22.0$, df=5, $P<0.001$

Kruskal-Wallis test:

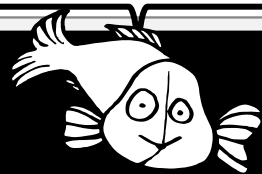
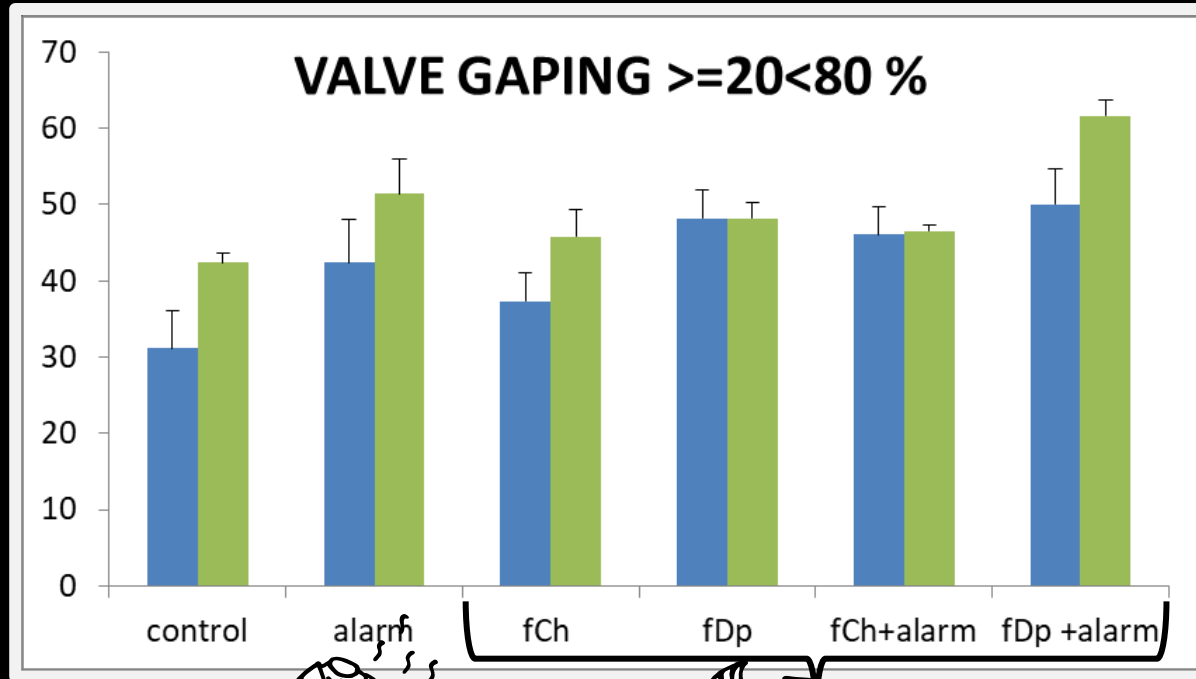
1h: $\chi^2=10.1$, df=5, $P=0.071$

2h: $\chi^2=21.9$, df=5, $P=0.006$

RESULTS – DURATION OF PARTICULAR GAPING LEVELS

(% OF TOTAL TIME)

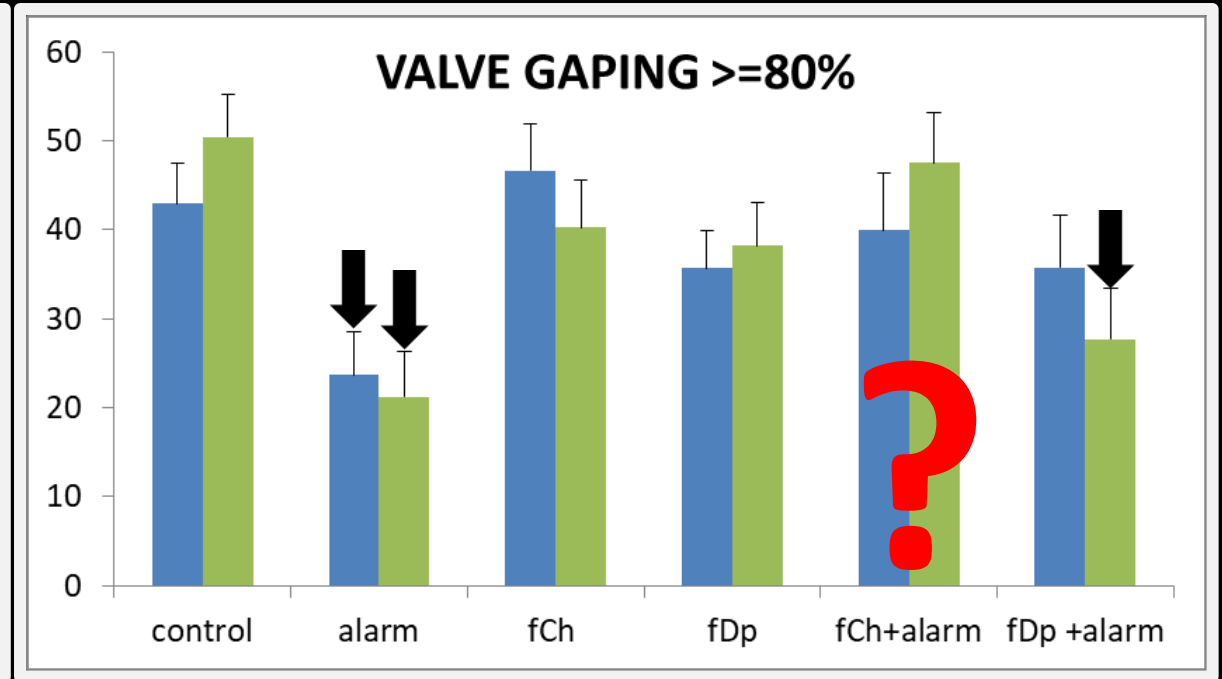
1h 2h



Kruskal-Wallis test:

1h: $\chi^2=8.6$, $df=5$, $P=0.127$

2h: $\chi^2=9.4$, $df=5$, $P=0.095$

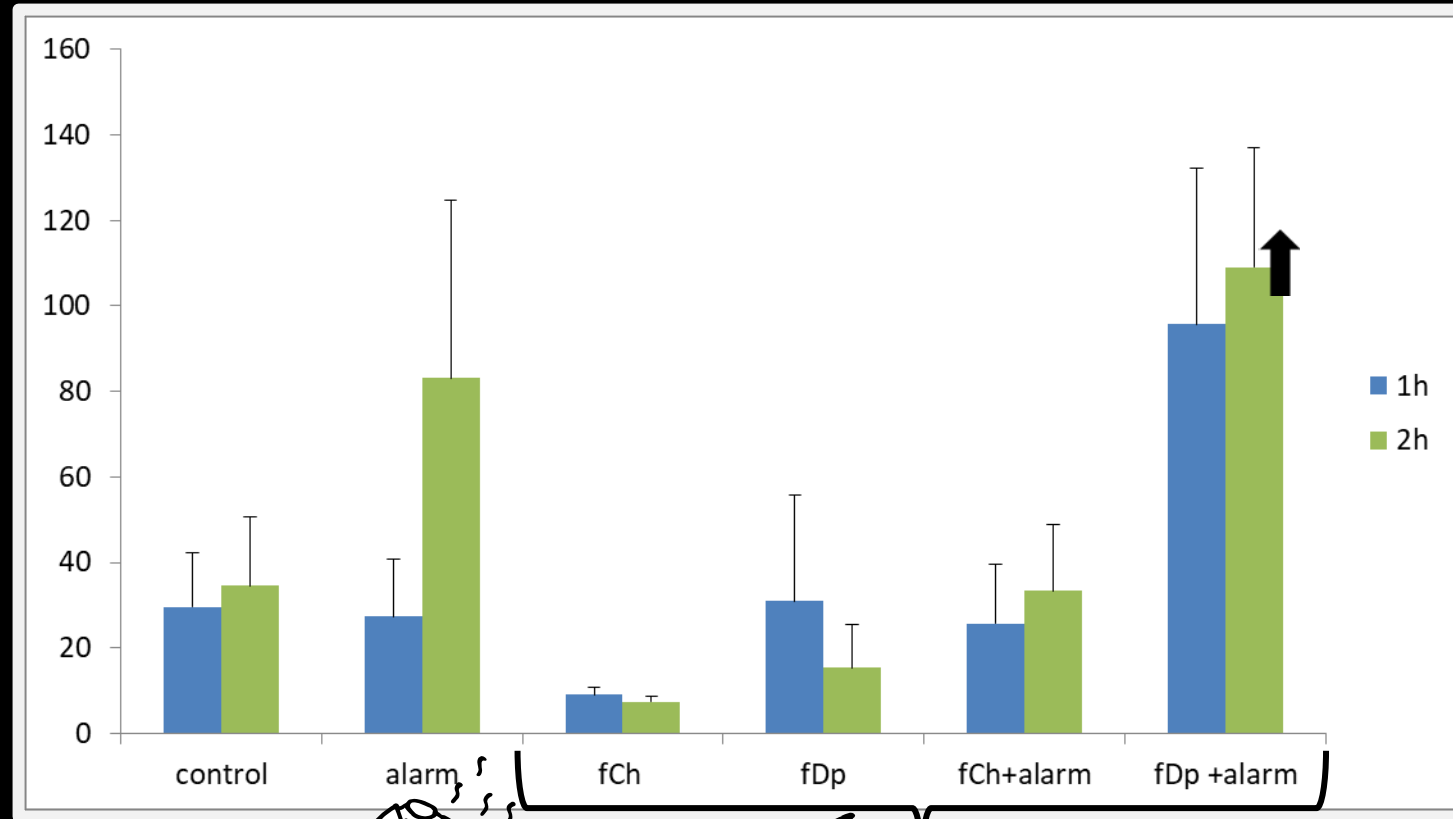


Kruskal-Wallis test:

1h: $\chi^2=11.3$, $df=5$, $P=0.045$

2h: $\chi^2=19.4$, $df=5$, $P=0.002$

RESULTS – NUMBER OF THE VALVE OPENING EVENTS <20% OF THE MAXIMUM OF VALVE GAPING



Kruskal-Wallis test:

1h: $\chi^2=12.1$, df=5, $P=0.034$

2h: $\chi^2=18.4$, df=5, $P=0.002$

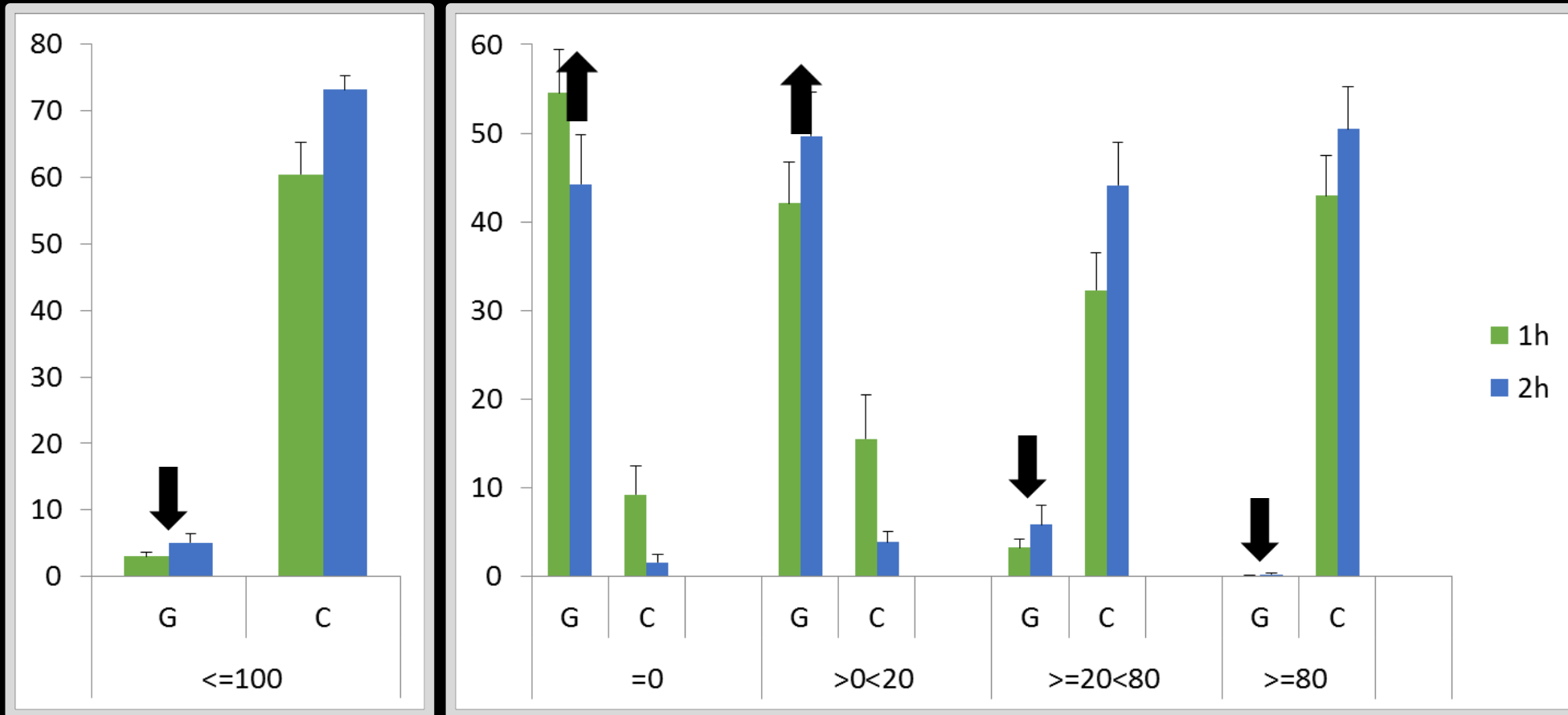
Alarm substance = more narrow valve openings

RESULTS – IMPACT OF D. VILLOSUS

DURATION OF PARTICULAR GAPING LEVELS

MEAN OPENING [%]

(% OF TOTAL TIME)



D. villosus = smaller mean opening, longer time spent in small gaping, shorter time spent in large gaping

G – presence of *D. villosus*
C - control

CONCLUSIONS

- ❖ Zebra mussels exhibited a clear response to the conspecific alarm substance:
 - narrower mean opening during the entire experiment
 - shorter time spent with valves gaping $\geq 80\%$ of the maximum opening and longer time spent with valves gaping $>0 < 20\%$.
- ❖ Tested mussels did not clearly respond to the scent of the roach. We suppose that roach kairomones may:
 - ? - stimulate specific responses to the alarm substance (e.g. number of opening events)
 - - „mask” alarm substance what can be beneficial for the predator.
- ❖ The physical presence of *Dikerogammarus villosus* is a strong stress factor for the zebra mussel (mussels spent more than 50% of time with closed valves).



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