A Decade of Gene Diversification by Viral Hemorrhagic Septicemia (VHS) since its first appearance in the Laurentian Great Lakes







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(now at Cleve. Clinic)



Viral Hemorrhagic Septicemia virus (VHSv)

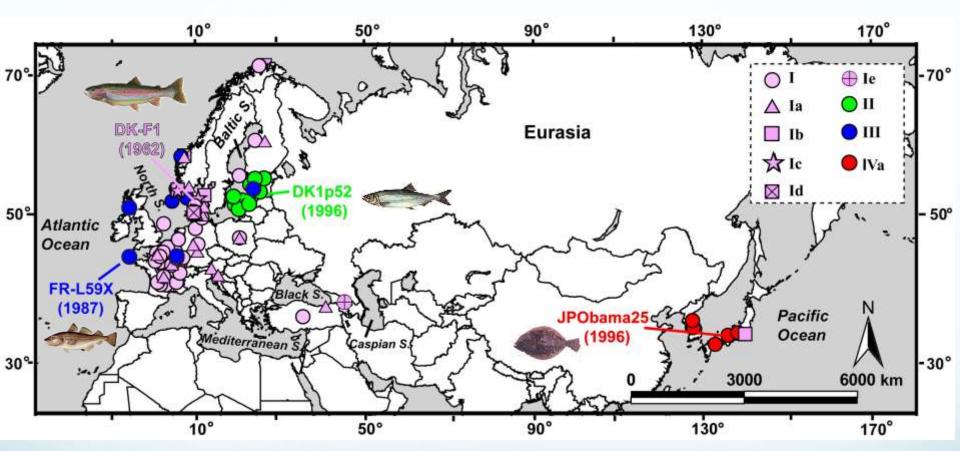


- RNA bullet-shaped Novirhabdovirus
- 11,158 nt, 6 genes: 3'N-P-M-G-Nv-L'5
 - Unique Nv (Nonvirion) gene to this group
 - Related to IHNV, Snakehead rhabdovirus
- One of world's most important finfish diseases
- External & internal hemorrhages
- Infects >80 fish species acrossN. Hemisphere
- New substrain (IVb) emerged in the Great Lakes~ decade ago
 - Large outbreaks across GL
 - ~30 species killed





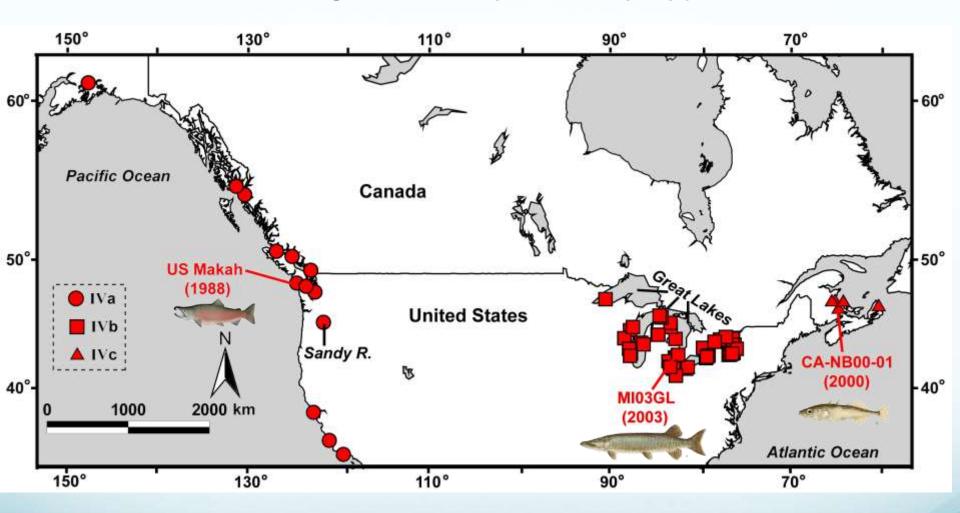
VHSv Distribution in Eurasia



(Dates=first known occurrences)

- First discovered in 1938 European rainbow trout aquaculture
- Now: Four strains (I, II, III, & IV) & several substrains (adapted from Pierce & Stepien 2012, Mol Phyl Evol)

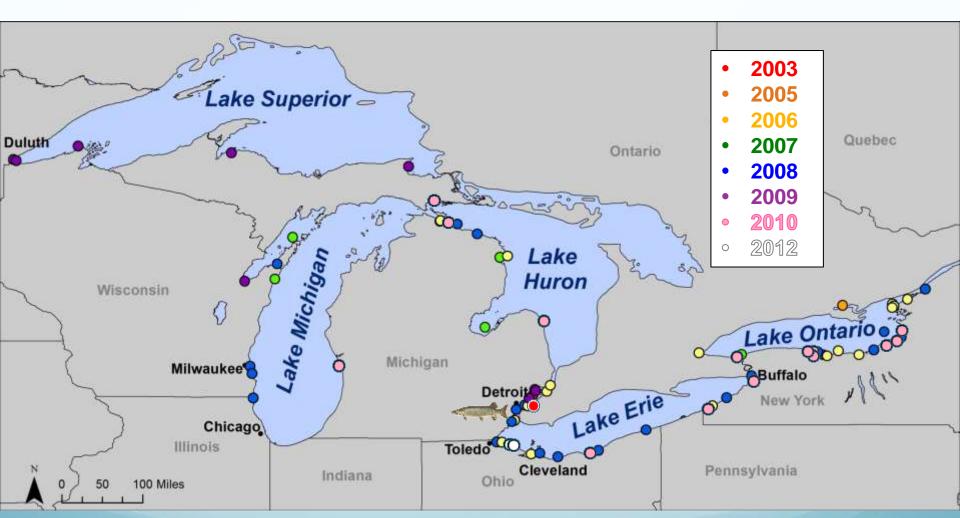
VHSv-IV in North America



(Dates=first known occurrences)

Three geographically separated substrains of IV (IVa, b, c) (adapted from Pierce & Stepien 2012, Mol Phyl Evol)

VHSv-IVb Spread in the Great Lakes



Data compiled from: J. Sieracki & J. Bossenbroek, UT (2003-2009); Cornwell et al. 2015 (2010 samples); Stepien et al. 2015 (2012 samples)

Major Species Infected by VHSv-IVb in the Great Lakes





Brown bullhead Ameiurus nebulosus





Burbot Lota lota

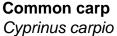


Emerald shiner Notropis atherinoides



Freshwater drum Aplodinotus grunniens







Channel catfish Ictalurus punctatus



Gizzard shad Dorosoma cepedianum



Lake herring Coregonus artedi



Oncorhynchus tshawytscha



Oncorhynchus mykiss





Trout perch Percopsis omiscomavcus



Walleye Sander vitreus

VHSv Impacts on Great Lakes Region

- Commercial & sport fishing
- Tourism & public perception
- Secondary pathogens & biological wastes
- Aquaculture, hatchery & baitfish industries





VHSv Characteristics & Spread

Ecology:

- Stable 3-15°C in water
- Long-lived in water (~14 days)
- Fish shed virus up to 15 weeks post infection
- Continue to shed virus after recovery in times of of stress

Transport:

- Fish migration (spawning)
- Ballast water from fishing boats
- Bait fish may transport
- Birds' feet
- Benthic invertebrates (leeches & Diporeia spp.)
- Aquatic turtles





Study Objectives

- 1. Evaluate the overall phylogenetic & biogeographic history & relationships of VHSv
- 2. Survey/ Test for current infection across the GL with our PLOS-One (2013) assay
- Discern the evolutionary diversification patterns & virus-host coevolution of IVb in the Great Lakes



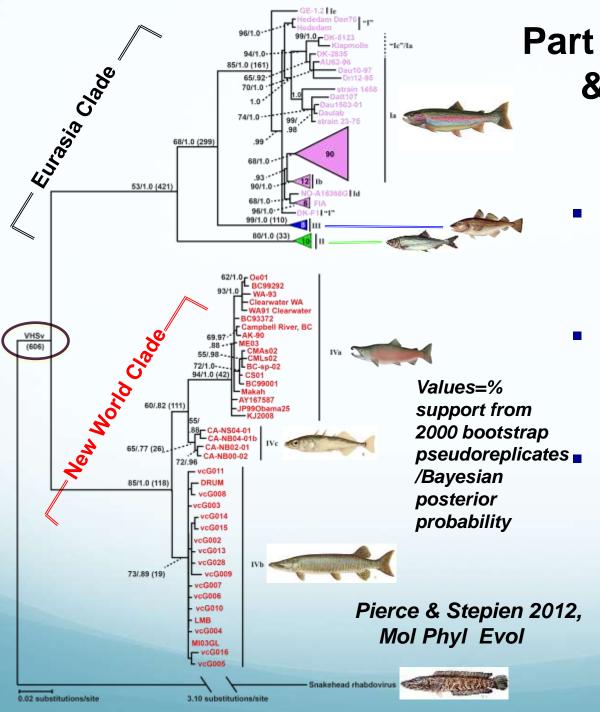












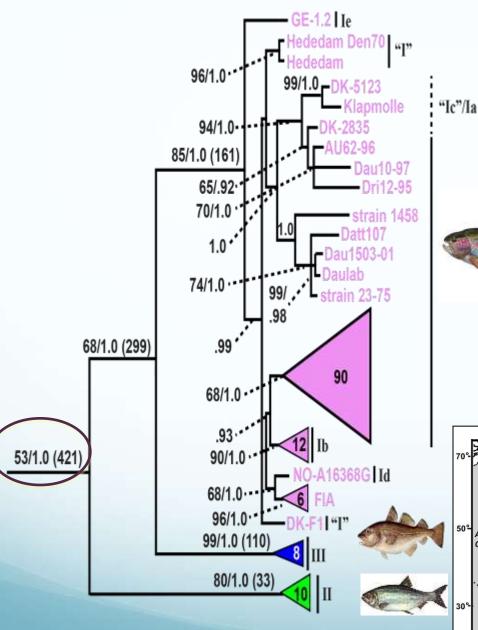
Part 1. Biogeographic & Phylogenetic Results

(G, Nv, N, M, P-genes)

- Phylogenetic analyses: Maximum Likelihood & Mr. Bayes
 - VHS likely originated from a marine ancestor in N Atlantic

Diverged ~600ya into 2 primary clades:

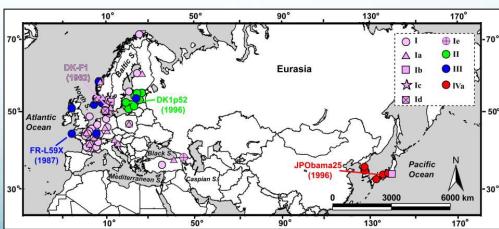
- I, II, III (Eurasia)
- IV (New World)
- IV has 3 substrains, which also are geographically distinct



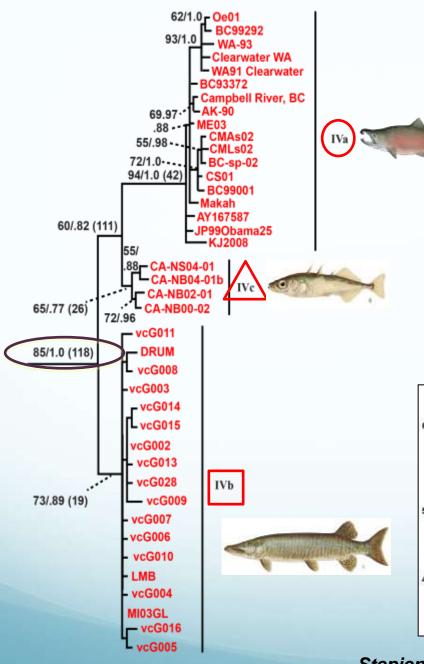
VHSv Eurasian Clade: Strains I, II & III

 The I-III clade then diverged ~420ya into 2 clades:

- II (marine) Baltic Sea region
- I (marine/estuarine/freshwater)
 - + III (marine/estuarine)
- Strains | & III diverged ~300ya
 - III separated in the North Atlantic Ocean & North Sea
 - I radiated in European salmonids

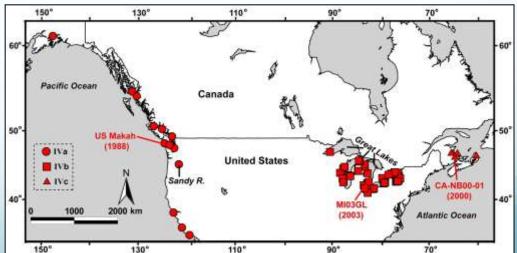


Pierce & Stepien 2012, MPE



VHSv New World Clade: Strain IV

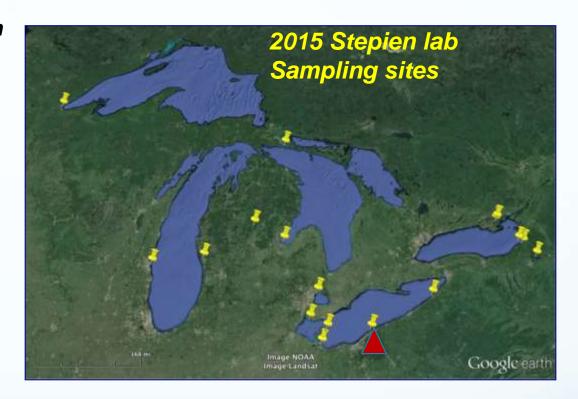
- Likely originated in marine fishes in N Atlantic
- IV diverged ~120ya into3 distinct substrains:
 - IVa = (~40ya) N Pacific Ocean
 - IVc = (~25ya) NW Atlantic Ocean
 - IVb = (~20ya) Great Lakes

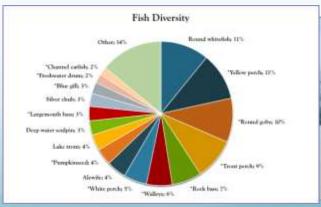


Stepien et al. 2015, PLOS One;, Pierce & Stepien 2012, Molecular Phylogenetics & Evolution

Part 2: 2015 VHSv-IVb Sampling Results

- Students (PhD candidate Megan Niner & undergraduate Shelby Edwards) sampled 1,193 fishes with federal & state agencies
 - April-July 2015
 - 40 fish species
 - Positives only at Fairport Harbor, central Lake Erie
- Screened pooled groups & individuals with our SYBR Green qPCR Assay (PLOS ONE 2013)
- Followed by our 2-color fluorescence quantitative assay
 - Uses synthetic internal standards
 - Pierce, Willey, Leaman, Shepherd, Stepien 2013a,b (J Virological Methods, PLOS ONE 2013)

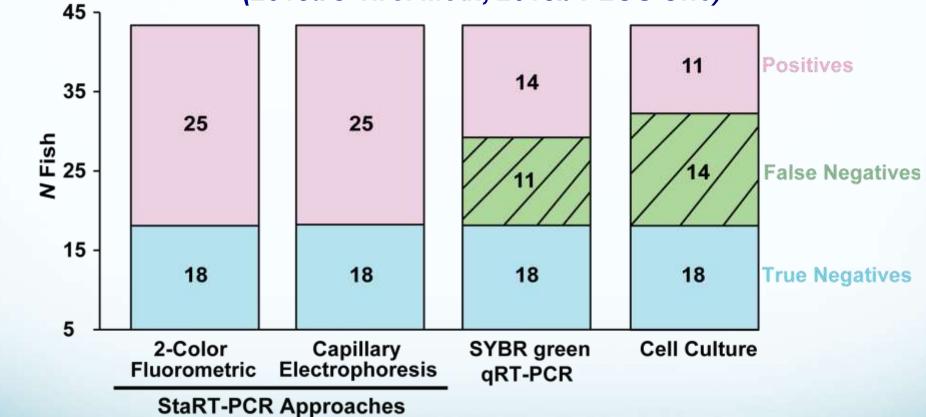






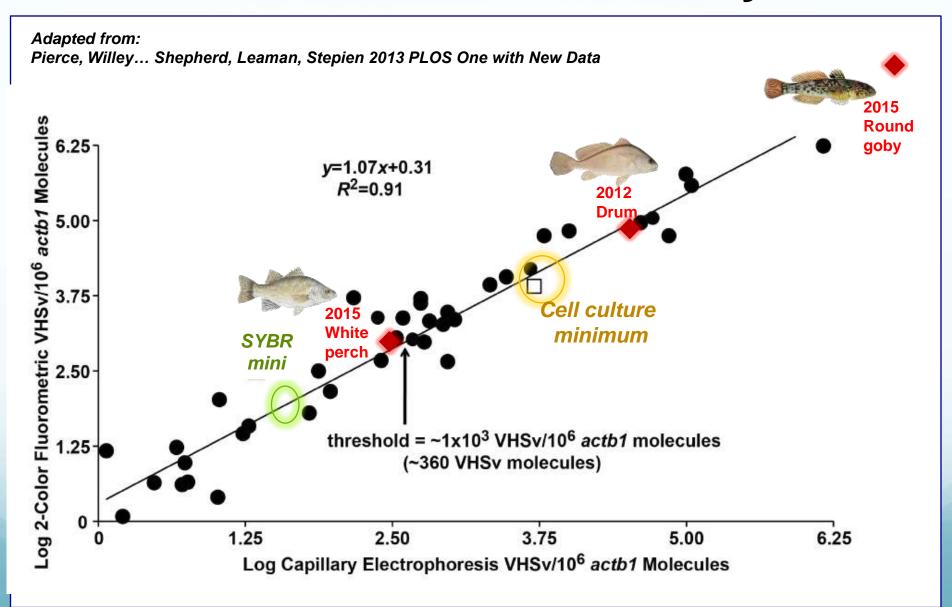
Developed New Accurate Assays to Detect & Quantify VHSv

Pierce, Willey... Leaman, Shepherd, Stepien (2013a J Virol Meth, 2013b PLOS One)

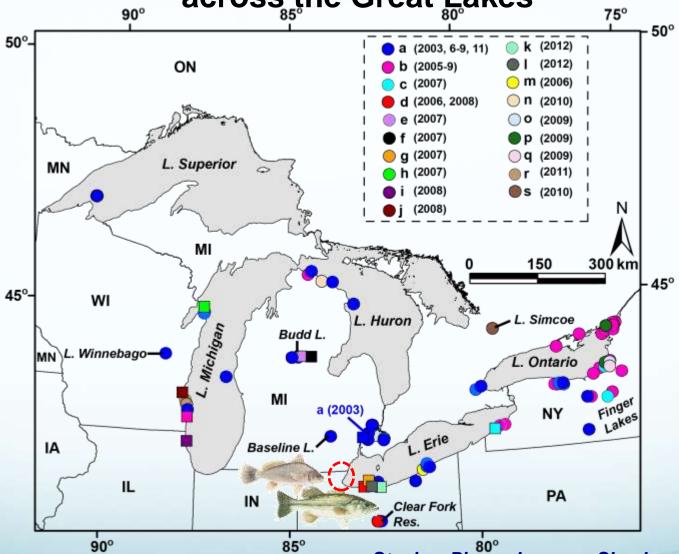


- 1) Both StaRT-PCR assays detected the same # of positives & negatives
- 2) Both were more sensitive than SYBR green qRT-PCR (χ^2 =5.68, p=0.02) & Cell Culture (χ^2 =9.36, p=0.002)
- 3) False negative rates=38% SYBR green & 44% Cell Culture

VHSv Concentration Analyses



Part 3: Genetics/Genomics over Time & Space across the Great Lakes



21 G-gene haplotypes identified to date

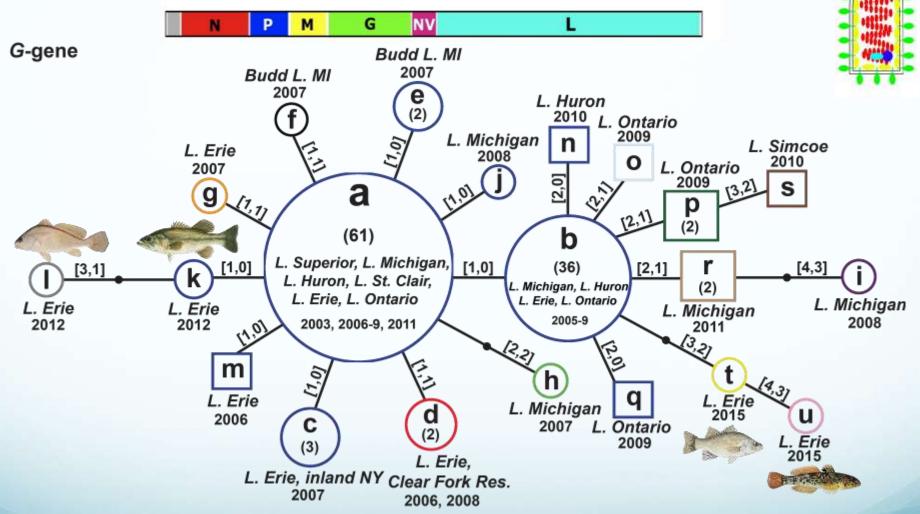
We are sequencing these for other genes

Stepien, Pierce, Leaman, Shepherd, Niner 2015 PLOS One

With info from Dr. Gael Kurath, USGS, Seattle

3. Evolutionary Patterns of VHSv-IVb:

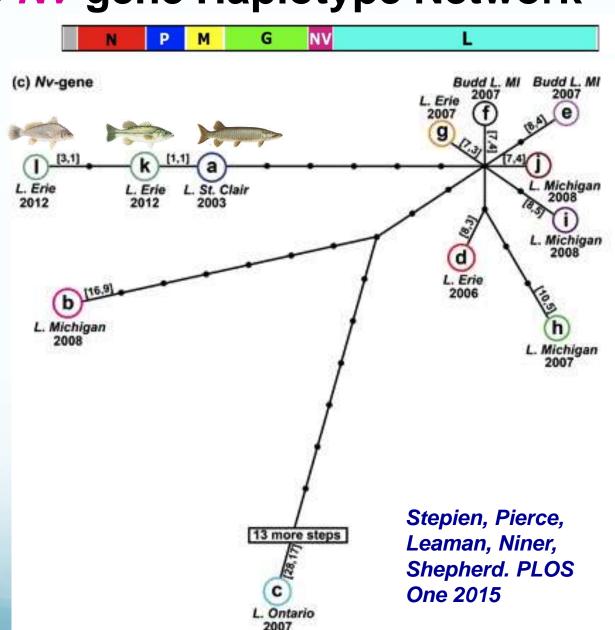
G-(glycoprotein) gene Haplotype Network



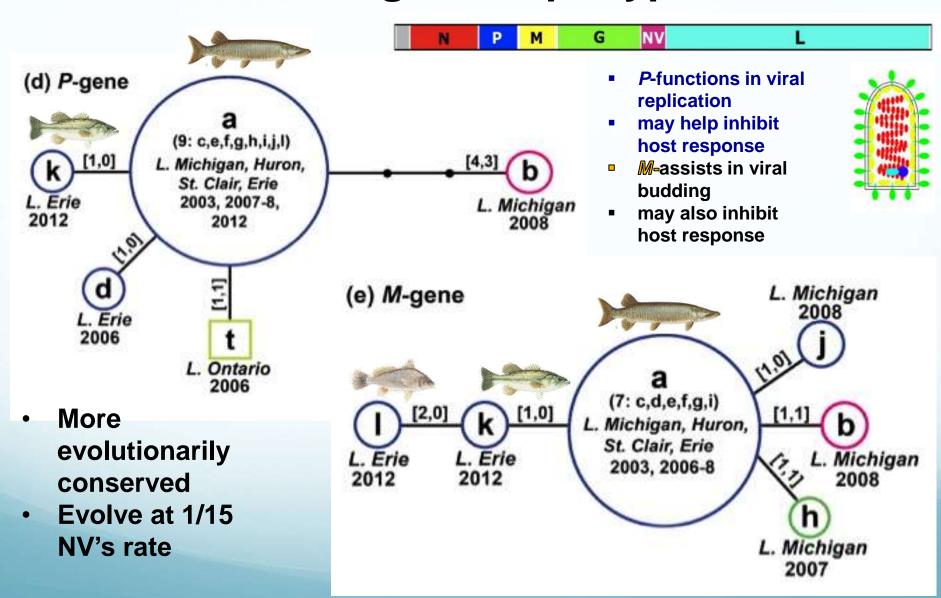
- Two central haplotypes -a & b- abundant & widespread; likely the oldest
- Several haplotypes radiated in a "star-like" pattern (i.e., quasispecies)
- •G evolves at ~1/10 rate of Nv
- G functions in attachment & entry into host cells

VHSv-IVb Nv-gene Haplotype Network

- Nv evolves fastest
 μ=2.1x10⁻³
- All G-gene variants also have unique Nv sequences
- All but one Nv variants differed in amino acids
- Functions in viral replication
- Anti-apoptotic role in early infection
- Involved in pathogenicity
- New paper: influences interferon pathway in fish host (Cano et al. 2016, Veterinary Microbiology)

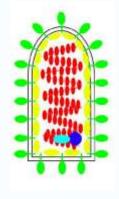


VHSv-IVb P & M-gene Haplotype Networks



VHS Summary to Date

- VHSv likely originated in the N Atlantic
 - Marine reservoir
 - Split into 2 primary clades: Eurasia & New World
 - Successful invader of freshwater systems
- VHSv-IVb may have invaded the Great Lakes via St. Lawrence Seaway
- IVb has diversified rapidly into many unique variants
- Follows a quasispecies mode of evolution
- Nv gene evolves the fastest (>N>G>P>M)
- Current directions:
 - Testing for new variants, sampling across GL
 - Whole genome sequencing
 - Analyzing virulence of variants in challenge experiments
 - Testing knock-out gene variants (w/ Dr. Vikram Vakharia, U Md)





Thank You:

Virus & Samples:

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